

**STATA META-ANALYSIS REFERENCE
MANUAL
RELEASE 16**



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Cross-referencing the documentation

When reading this manual, you will find references to other Stata manuals, for example, [U] [27 Overview of Stata estimation commands](#); [R] [regress](#); and [D] [reshape](#). The first example is a reference to chapter 27, *Overview of Stata estimation commands*, in the *User's Guide*; the second is a reference to the [regress](#) entry in the *Base Reference Manual*; and the third is a reference to the [reshape](#) entry in the *Data Management Reference Manual*.

All the manuals in the Stata Documentation have a shorthand notation:

[GSM]	<i>Getting Started with Stata for Mac</i>
[GSU]	<i>Getting Started with Stata for Unix</i>
[GSW]	<i>Getting Started with Stata for Windows</i>
[U]	<i>Stata User's Guide</i>
[R]	<i>Stata Base Reference Manual</i>
[BAYES]	<i>Stata Bayesian Analysis Reference Manual</i>
[CM]	<i>Stata Choice Models Reference Manual</i>
[D]	<i>Stata Data Management Reference Manual</i>
[DSGE]	<i>Stata Dynamic Stochastic General Equilibrium Models Reference Manual</i>
[ERM]	<i>Stata Extended Regression Models Reference Manual</i>
[FMM]	<i>Stata Finite Mixture Models Reference Manual</i>
[FN]	<i>Stata Functions Reference Manual</i>
[G]	<i>Stata Graphics Reference Manual</i>
[IRT]	<i>Stata Item Response Theory Reference Manual</i>
[LASSO]	<i>Stata Lasso Reference Manual</i>
[XT]	<i>Stata Longitudinal-Data/Panel-Data Reference Manual</i>
[META]	<i>Stata Meta-Analysis Reference Manual</i>
[ME]	<i>Stata Multilevel Mixed-Effects Reference Manual</i>
[MI]	<i>Stata Multiple-Imputation Reference Manual</i>
[MV]	<i>Stata Multivariate Statistics Reference Manual</i>
[PSS]	<i>Stata Power, Precision, and Sample-Size Reference Manual</i>
[P]	<i>Stata Programming Reference Manual</i>
[RPT]	<i>Stata Reporting Reference Manual</i>
[SP]	<i>Stata Spatial Autoregressive Models Reference Manual</i>
[SEM]	<i>Stata Structural Equation Modeling Reference Manual</i>
[SVY]	<i>Stata Survey Data Reference Manual</i>
[ST]	<i>Stata Survival Analysis Reference Manual</i>
[TS]	<i>Stata Time-Series Reference Manual</i>
[TE]	<i>Stata Treatment-Effects Reference Manual: Potential Outcomes/Counterfactual Outcomes</i>
[I]	<i>Stata Glossary and Index</i>
[M]	<i>Mata Reference Manual</i>

Description

Meta-analysis ([Glass 1976](#)) is a statistical technique for combining the results from several similar studies. The results of multiple studies that answer similar research questions are often available in the literature. It is natural to want to compare their results and, if sensible, provide one unified conclusion. This is precisely the goal of the meta-analysis, which provides a single estimate of the effect of interest computed as the weighted average of the study-specific effect estimates. When these estimates vary substantially between the studies, meta-analysis may be used to investigate various causes for this variation.

Another important focus of the meta-analysis may be the exploration and impact of small-study effects, which occur when the results of smaller studies differ systematically from the results of larger studies. One of the common reasons for the presence of small-study effects is publication bias, which arises when the results of published studies differ systematically from all the relevant research results.

Comprehensive overview of meta-analysis may be found in [Sutton and Higgins \(2008\)](#); [Cooper, Hedges, and Valentine \(2009\)](#); [Borenstein et al. \(2009\)](#); [Higgins and Green \(2017\)](#); [Hedges and Olkin \(1985\)](#); [Sutton et al. \(2000a\)](#); and [Palmer and Sterne \(2016\)](#). A book dedicated to addressing publication bias was written by [Rothstein, Sutton, and Borenstein \(2005\)](#).

This entry presents a general introduction to meta-analysis and describes relevant statistical terminology used throughout the manual. For how to perform meta-analysis in Stata, see [\[META\] meta](#).

Remarks and examples

Remarks are presented under the following headings:

Brief overview of meta-analysis

Meta-analysis models

Common-effect (“fixed-effect”) model

Fixed-effects model

Random-effects model

Comparison between the models and interpretation of their results

Meta-analysis estimation methods

Forest plots

Heterogeneity

Assessing heterogeneity

Addressing heterogeneity

Subgroup meta-analysis

Meta-regression

Publication bias

Funnel plots

Tests for funnel-plot asymmetry

The trim-and-fill method

Cumulative meta-analysis

Brief overview of meta-analysis

The term meta-analysis refers to the analysis of the data obtained from a collection of studies that answer similar research questions. These studies are known as *primary studies*. Meta-analysis uses statistical methods to produce an overall estimate of an effect, explore between-study heterogeneity, and investigate the impact of publication bias or, more generally, small-study effects on the final results. [Pearson \(1904\)](#) provides the earliest example of what we now call meta-analysis. In that reference, the average of study-specific correlation coefficients was used to estimate an overall effect of vaccination against smallpox on subjects' survival.

There is a lot of information reported by a myriad of studies, which can be intimidating and difficult to absorb. Additionally, these studies may report conflicting results in terms of the magnitudes and even direction of the effects of interest. For example, many studies that investigated the effect of taking aspirin for preventing heart attacks reported contradictory results. Meta-analysis provides a principled approach for consolidating all of this overwhelming information to provide an overall conclusion or reasons for why such a conclusion cannot be reached.

Meta-analysis has been used in many fields of research. See the Cochrane Collaboration (<https://us.cochrane.org/>) for a collection of results from meta-analysis that address various treatments from all areas of healthcare. Meta-analysis has also been used in econometrics (for example, [Dalhuisen et al. \[2003\]](#); [Woodward and Wui \[2001\]](#); [Hay, Knechel, and Wang \[2006\]](#); [Card, Kluve, and Weber \[2010\]](#)); education (for example, [Bernard et al. \[2004\]](#); [Fan and Chen \[2001\]](#)); psychology (for example, [Sin and Lyubomirsky \[2009\]](#); [Barrick and Mount \[1991\]](#); [Harter, Schmidt, and Hayes \[2002\]](#)); psychiatry (for example, [Hanji 2017](#)); criminology (for example, [Gendreau, Little, and Goggin \[1996\]](#); [Pratt and Cullen \[2000\]](#)); and ecology (for example, [Hedges, Gurevitch, and Curtis \[1999\]](#); [Gurevitch, Curtis, and Jones \[2001\]](#); [Winfree et al. \[2009\]](#); [Arnqvist and Wooster \[1995\]](#)).

Meta-analysis is the statistical-analysis step of a systematic review. The term systematic review refers to the entire process of integrating the empirical research to achieve unified and potentially more general conclusions. Meta-analysis provides the theoretical underpinning of a systematic review and sets it apart from a narrative review; in the latter, an area expert summarizes the study-specific results and provides final conclusions, which could lead to potentially subjective and difficult-to-replicate findings. The theoretical soundness of meta-analysis made systematic reviews the method of choice for integrating empirical evidence from multiple studies. See [Cooper, Hedges, and Valentine \(2009\)](#) for more information as well as for various stages of a systematic review.

In what follows, we briefly describe the main components of meta-analysis: effect sizes, forest plots, heterogeneity, and publication bias.

Effect sizes. Effect sizes (or various measures of outcome) and their standard errors are the two most important components of a meta-analysis. They are obtained from each of the primary studies prior to the meta-analysis. Effect sizes of interest depend on the research objective and type of study. For example, in a meta-analysis of binary outcomes, odds ratios and risk ratios are commonly used, whereas in a meta-analysis of continuous outcomes, Hedges's g and Cohen's d measures are commonly used. An overall effect size is computed as a weighted average of study-specific effect sizes, with more precise (larger) studies having larger weights. The weights are determined by the chosen meta-analysis model; see [Meta-analysis models](#). Also see [\[META\] meta esize](#) for how to compute various effect sizes in a meta-analysis.

Meta-analysis models. Another important consideration for meta-analysis is that of the underlying model. Three commonly used models are a common-effect, fixed-effects, and random-effects models. The models differ in how they estimate and interpret parameters. See [Meta-analysis models](#) for details.

Meta-analysis summary—forest plots. The results of meta-analysis are typically summarized on a forest plot, which plots the study-specific effect sizes and their corresponding confidence intervals, the combined estimate of the effect size and its confidence interval, and other summary measures such as heterogeneity statistics. See *Forest plots* for details.

Heterogeneity. The estimates of effect sizes from individual studies will inherently vary from one study to another. This variation is known as a study heterogeneity. Two types of heterogeneity described by [Deeks, Higgins, and Altman \(2017\)](#) are methodological, when the studies differ in design and conduct, and clinical, when the studies differ in participants, treatments, and exposures or outcomes. The authors also define statistical heterogeneity, which exists when the observed effects differ between the studies. It is typically a result of clinical heterogeneity, methodological heterogeneity, or both. There are methods for assessing and addressing heterogeneity that we discuss in detail in *Heterogeneity*.

Publication bias. The selection of studies in a meta-analysis is an important step. Ideally, all studies that meet prespecified selection criteria must be included in the analysis. This is rarely achievable in practice. For instance, it may not be possible to have access to some unpublished results. So some of the relevant studies may be omitted from the meta-analysis. This may lead to what is known in statistics as a sample-selection problem. In the context of meta-analysis, this problem is known as publication bias or, more generally, reporting bias. Reporting bias arises when the omitted studies are systematically different from the studies selected in the meta-analysis. For details, see *Publication bias*.

Finally, you may ask, Does it make sense to combine different studies? According to [Borenstein et al. \(2009, chap. 40\)](#), “in the early days of meta-analysis, Robert Rosenthal was asked whether it makes sense to perform a meta-analysis, given that the studies differ in various ways and that the analysis amounts to combining apples and oranges. Rosenthal answered that *combining apples and oranges makes sense if your goal is to produce a fruit salad*.”

Meta-analysis would be of limited use if it could combine the results of identical studies only. The appeal of meta-analysis is that it actually provides a principled way of combining a broader set of studies and can answer broader questions than those originally posed by the included primary studies. The specific goals of the considered meta-analysis should determine which studies can be combined and, more generally, whether a meta-analysis is even applicable.

Meta-analysis models

The role of a meta-analysis model is important for the computation and interpretation of the meta-analysis results. Different meta-analysis models make different assumptions and, as a result, estimate different parameters of interest. In this section, we describe the available meta-analysis models and point out the differences between them.

Suppose that there are K independent studies. Each study reports an estimate, $\hat{\theta}_j$, of the unknown true effect size θ_j and an estimate, $\hat{\sigma}_j$, of its standard error, $j = 1, 2, \dots, K$. The goal of a meta-analysis is to combine these estimates in a single result to obtain valid inference about the population parameter of interest, θ_{pop} .

Depending on the research objective and assumptions about studies, three approaches are available to model the effect sizes: a common-effect model (historically known as a fixed-effect model—notice the singular “effect”), a fixed-effects model (notice the plural “effects”), and a random-effects model. We briefly define the three models next and describe them in more detail later.

Consider the model

$$\widehat{\theta}_j = \theta_j + \epsilon_j \quad j = 1, 2, \dots, K \quad (1)$$

where ϵ_j 's are sampling errors and $\epsilon_j \sim N(0, \sigma_j^2)$. Although σ_j^2 's are unknown, meta-analysis does not estimate them. Instead, it treats the estimated values, $\widehat{\sigma}_j^2$'s, of these variances as known and uses them during estimation. In what follows, we will thus write $\epsilon_j \sim N(0, \widehat{\sigma}_j^2)$.

A common-effect model, as suggested by its name, assumes that all study effect sizes in (1) are the same and equal to the true effect size θ ; that is, $\theta_j = \theta_{j'} = \theta$ for $j \neq j'$. The research questions and inference relies heavily on this assumption, which is often violated in practice.

A fixed-effects model assumes that the study effect sizes in (1) are different, $\theta_j \neq \theta_{j'}$ for $j \neq j'$, and “fixed”. That is, the studies included in the meta-analysis define the entire population of interest. So the research questions and inference concern only the specific K studies included in the meta-analysis.

A random-effects model also assumes that the study effect sizes in (1) are different, $\theta_j \neq \theta_{j'}$ for $j \neq j'$, but that they are “random”. That is, the studies in the meta-analysis represent a sample from a population of interest. The research questions and inference extend beyond the K studies included in the meta-analysis to the entire population of interest.

The models differ in the population parameter, θ_{pop} , they estimate; see [Comparison between the models and interpretation of their results](#). Nevertheless, they all use the weighted average as the estimator for θ_{pop} :

$$\widehat{\theta}_{\text{pop}} = \frac{\sum_{j=1}^K w_j \widehat{\theta}_j}{\sum_{j=1}^K w_j} \quad (2)$$

However, they differ in how they define the weights w_j .

We describe each model and the parameter they estimate in more detail below.

Common-effect (“fixed-effect”) model

As we mentioned earlier, a common-effect (CE) meta-analysis model ([Hedges 1982](#); [Rosenthal and Rubin 1982](#)) is historically known as a fixed-effect model. The term “fixed-effect model” is easy to confuse with the “fixed-effects model” (plural), so we avoid it in our documentation. The term “common-effect”, as suggested by [Rice, Higgins, and Lumley \(2018\)](#), is also more descriptive of the underlying model assumption. A CE model assumes a common (one true) effect for all studies in (1):

$$\widehat{\theta}_j = \theta + \epsilon_j \quad j = 1, 2, \dots, K$$

The target of interest in a CE model is an estimate of a common effect size, $\theta_{\text{pop}} = \theta$. The CE model generally uses the weights $w_j = 1/\widehat{\sigma}_j$ in (2) to estimate θ .

CE models are applicable only when the assumption that the same parameter underlies each study is reasonable, such as with pure replicate studies.

Fixed-effects model

A fixed-effects (FE) meta-analysis model (Hedges and Vevea 1998; Rice, Higgins, and Lumley 2018) is defined by (1); it assumes that different studies have different effect sizes ($\theta_1 \neq \theta_2 \neq \dots \neq \theta_K$) and that the effect sizes are fixed quantities. By fixed quantities, we mean that the studies included in the meta-analysis define the entire population of interest. FE models are typically used whenever the analyst wants to make inferences only about the included studies.

The target of interest in an FE model is an estimate of the weighted average of true study-specific effect sizes,

$$\theta_{\text{pop}} = \text{Ave}(\theta_j) = \frac{\sum_{j=1}^K W_j \theta_j}{\sum_{j=1}^K W_j}$$

where W_j 's represent true, unknown weights, which are defined in Rice, Higgins, and Lumley (2018, eq. 3). The estimated weights, $w_j = 1/\hat{\sigma}_j$, are generally used in (2) to estimate θ_{pop} .

Based on Rice, Higgins, and Lumley (2018), an FE model answers the question, “What is the magnitude of the average true effects in the set of K studies included in the meta-analysis?” It is appropriate when the true effects sizes are different across studies and the research interest lies in their average estimate.

Random-effects model

A random-effects (RE) meta-analysis model (Hedges 1983; DerSimonian and Laird 1986) assumes that the study effect sizes are different and that the collected studies represent a random sample from a larger population of studies. (The viewpoint of random effect sizes is further explored by Bayesian meta-analysis; see, for example, *Random-effects meta-analysis of clinical trials* in [BAYES] **bayesmh**.) The goal of RE meta-analysis is to provide inference for the population of studies based on the sample of studies used in the meta-analysis.

The RE model may be described as

$$\hat{\theta}_j = \theta_j + \epsilon_j = \theta + u_j + \epsilon_j$$

where $u_j \sim N(0, \tau^2)$ and, as before, $\epsilon_j \sim N(0, \hat{\sigma}_j^2)$. Parameter τ^2 represents the between-study variability and is often referred to as the heterogeneity parameter. It estimates the variability among the studies, beyond the sampling variability. When $\tau^2 = 0$, the RE model reduces to the CE model.

Here the target of inference is $\theta_{\text{pop}} = E(\theta_j)$, the mean of the distribution of effect sizes θ_j 's. θ_{pop} is estimated from (2) with $w_j = 1/(\hat{\sigma}_j^2 + \hat{\tau}^2)$.

Comparison between the models and interpretation of their results

CE and FE models are computationally identical but conceptually different. They differ in their target of inference and the interpretation of the overall effect size. In fact, all three models have important conceptual and interpretation differences. Table 1 summarizes the different interpretations of θ_{pop} under the three models.

Table 1. Interpretation of θ_{pop} under various meta-analysis models

Model	Interpretation of θ_{pop}
common-effect	common effect ($\theta_1 = \theta_2 = \dots = \theta_K = \theta$)
fixed-effects	weighted average of the K true study effects
random-effects	mean of the distribution of $\theta_j = \theta + u_j$

A CE meta-analysis model estimates the true effect size under the strong assumption that all studies share the same effect and thus all the variability between the studies is captured by the sampling errors. Under that assumption, the weighted average estimator indeed estimates the true common effect size, θ .

In the presence of additional variability unexplained by sampling variations, the interpretation of the results depends on how this variability is accounted for in the analysis.

An FE meta-analysis model uses the same weighted average estimator as a CE model, but the latter now estimates the weighted average of the K true study-specific effect sizes, $\text{Ave}(\theta_j)$.

An RE meta-analysis model assumes that the study contributions, u_j 's, are random. It decomposes the variability of the effect sizes into the between-study and within-study components. The within-study variances, $\hat{\sigma}_j^2$'s, are assumed known by design. The between-study variance, τ^2 , is estimated from the sample of the effect sizes. Thus, the extra variability attributed to τ^2 is accounted for during the estimation of the mean effect size, $E(\theta_j)$.

So which model should you choose? The literature recommends to start with a random-effects model, which is Stata's default for most meta-analyses. If you are willing to assume that the studies have different true effect sizes and you are interested only in providing inferences about these specific studies, then the FE model is appropriate. If the assumption of study homogeneity is reasonable for your data, a CE model may be considered.

Meta-analysis estimation methods

Depending on the chosen meta-analysis model, various methods are available to estimate the weights w_j in (2). The meta-analysis models from the previous sections assumed the inverse-variance estimation method (Whitehead and Whitehead 1991) under which the weights are inversely related to the variance. The inverse-variance estimation method is applicable to all meta-analysis models and all types of effect sizes. Thus, it can be viewed as the most general approach.

For binary data, CE and FE models also support the Mantel–Haenszel estimation method, which can be used to combine odds ratios, risk ratios, and risk differences. The classical Mantel–Haenszel method (Mantel and Haenszel 1959) is used for odds ratios, and its extension by Greenland and Robins (1985) is used for risk ratios and risk differences. The Mantel–Haenszel method is recommended with sparse data. Fleiss, Levin, and Paik (2003) also suggests that it be used with small studies provided that there are many.

In RE models, the weights are inversely related to the total variance, $w_j = 1/(\hat{\sigma}_j^2 + \hat{\tau}^2)$. Different methods are proposed for estimating the between-study variability, τ^2 , which is used in the expression for the weights. These include the restricted maximum likelihood (REML), maximum likelihood (ML), empirical Bayes (EB), DerSimonian–Laird (DL), Hedges (HE), Sidik–Jonkman (SJ), and Hunter–Schmidt (HS).

REML, ML, and EB are iterative methods, whereas other methods are noniterative (have closed-form expressions). The former estimators produce nonnegative estimates of τ^2 . The other estimators, except SJ, may produce negative estimates and are thus truncated at zero when this happens. The SJ estimator always produces a positive estimate of τ^2 .

REML, ML, and EB assume that the distribution of random effects is normal. The other estimators make no distributional assumptions about random effects. Below, we briefly describe the properties of each method. See Sidik and Jonkman (2007), Viechtbauer (2005), and Veroniki et al. (2016) for a detailed discussion and the merit of each estimation method.

The REML method (Raudenbush 2009) produces an unbiased, nonnegative estimate of τ^2 and is commonly used in practice. (It is the default estimation method in Stata because it performs well in most scenarios.)

When the number of studies is large, the ML method (Hardy and Thompson 1998; Thompson and Sharp 1999) is more efficient than the REML method but may produce biased estimates when the number of studies is small, which is a common case in meta-analysis.

The EB estimator (Berkey et al. 1995), also known as the Paule–Mandel estimator (Paule and Mandel 1982), tends to be less biased than other RE methods, but it is also less efficient than REML or DL (Knapp and Hartung 2003).

The DL method (DerSimonian and Laird 1986), historically, is one of the most popular estimation methods because it does not make any assumptions about the distribution of the random effects and does not require iteration. But it may underestimate τ^2 , especially when the variability is large and the number of studies is small. However, when the variability is not too large and the studies are of similar sizes, this estimator is more efficient than other noniterative estimators HE and SJ. See Veroniki et al. (2016) for details and relevant references.

The SJ estimator (Sidik and Jonkman 2005), along with the EB estimator, is the best estimator in terms of bias for large τ^2 (Sidik and Jonkman 2007). This method always produces a positive estimate of τ^2 and thus does not need truncating at 0, unlike the other noniterative methods.

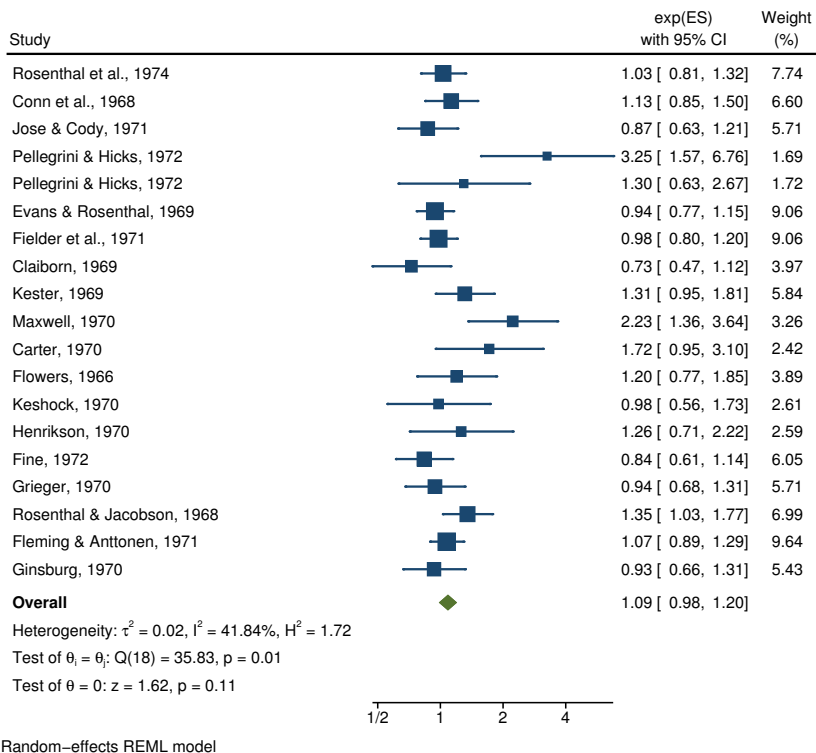
Like DL, the HE estimator (Hedges 1983) is a method of moments estimator, but, unlike DL, it does not weight effect-size variance estimates (DerSimonian and Laird 1986). Veroniki et al. (2016) note, however, that this method is not widely used in practice.

The HS estimator (Schmidt and Hunter 2015) is negatively biased and thus not recommended when unbiasedness is important (Viechtbauer 2005). Otherwise, the mean squared error of HS is similar to that of ML and is smaller than those of HE, DL, and REML.

Forest plots

Meta-analysis results are often presented using a forest plot (for example, Lewis and Ellis [1982]). A forest plot shows study-specific effect sizes and an overall effect size with their respective confidence intervals. The information about study heterogeneity and the significance of the overall effect size are also typically presented. This plot provides a convenient way to visually compare the study effect sizes, which can be any summary estimates available from primary studies, such as standardized and unstandardized mean differences, (log) odds ratios, (log) risk ratios, and (log) hazard ratios.

Below is an example of a forest plot.



A blue square is plotted for each study, with the size of the square being proportional to the study weight; that is, larger squares correspond to larger (more precise) studies. Studies' CIs are plotted as whiskers extending from each side of the square and spanning the width of the CI. The estimate of the overall effect size, depicted here by a green diamond, is typically plotted following the individual effect sizes. The diamond is centered at the estimate of the overall effect size and the width of the diamond represents the corresponding CI width. Heterogeneity measures such as the I^2 and H^2 statistics, homogeneity test, and the significance test of the overall effect sizes are also commonly reported.

Two further variations of forest plots are for cumulative and subgroup meta-analyses; see [Cumulative meta-analysis](#) and [Subgroup meta-analysis](#).

For further details about forest plots, see [\[META\] meta forestplot](#).

Heterogeneity

The exposition below is based on [Deeks, Higgins, and Altman \(2017\)](#) and references therein.

It is natural for effect sizes of studies collected in a meta-analysis to vary between the studies because of sampling variability. However, when this variation exceeds the levels that could be explained by sampling variation, it is referred to as the between-study heterogeneity. Between-study heterogeneity may arise for different reasons and is generally divided into two types: clinical and methodological ([Thompson 1994](#); [Deeks, Higgins, and Altman 2017](#)). Clinical heterogeneity is the variability in

the intervention strategies, outcomes, and study participants. Methodological heterogeneity is the variability in the study design and conduct. Statistical heterogeneity refers to the cases when the variability between the observed effects cannot be explained by sampling variability alone. It arises when the true effects in each study are different and may be the result of clinical heterogeneity, methodological heterogeneity, or both. In what follows, we refer to statistical heterogeneity simply as heterogeneity.

Assessing heterogeneity

Forest plots are useful for visual examination of heterogeneity. Its presence can be evaluated by looking at the plotted CIs, which are represented as horizontal lines on the plot. Heterogeneity is suspect if there is a lack of overlap between the CIs.

You can also test for heterogeneity more formally by using Cochrane's homogeneity test. Additionally, various heterogeneity measures such as the I^2 statistic, which estimates the percentage of the between-study variability, are available to quantify heterogeneity.

See [META] [meta summarize](#) for details.

Addressing heterogeneity

There are several strategies to address heterogeneity when it is present. Below, we summarize some of the recommendations from [Deeks, Higgins, and Altman \(2017\)](#):

1. *“Explore heterogeneity”*. [Subgroup analyses](#) and [meta-regression](#) are commonly used to explore heterogeneity. For such analyses to be proper, you must prespecify upfront (before your meta-analysis) the study attributes you would like to explore. Often, meta-analysts are already familiar with the studies, so the genuine prestudy specification may not be possible. In that case, you should use caution when interpreting the results. Once heterogeneity is established, its exploration after the fact is viewed as data snooping and should be avoided.
2. *“Perform an RE meta-analysis”*. After careful consideration of subgroup analysis and meta-regression, you may consider an RE meta-analysis to account for the remaining unexplained between-study heterogeneity. See [Deeks, Higgins, and Altman \(2017, sec. 9.5.4\)](#) for details.
3. *“Exclude studies”*. Generally, you should avoid excluding studies from a meta-analysis because this may lead to bias. You may consider doing this in the presence of a few outlying studies when the reasons for the outlying results are well understood and are unlikely to interfere with your research objectives. Even then, you still need to perform sensitivity analysis and report both the results with and without the outlying studies.
4. *“Do not perform a meta-analysis”*. In the presence of substantial variation that cannot be explained, you may have to abandon the meta-analysis altogether. In this case, it will be misleading to report a single overall estimate of an effect, especially if there is a disagreement among the studies about the direction of the effect.

Below, we discuss ways of exploring heterogeneity via subgroup meta-analysis and meta-regression.

Subgroup meta-analysis

It is not uncommon for the studies in a meta-analysis to report varying effect-size estimates. But it is important to understand and account for such variation during the meta-analysis to obtain reliable results ([Thompson 1994](#); [Berlin 1995](#)). In the presence of substantial between-study variability, meta-analysis may be used to explore the relationship between the effect sizes and study-level covariates of

interest, known in the meta-analysis literature as moderators. For example, the effect of a particular vaccine may depend on a study location, the effect of a particular drug may depend on the studies' dosages, and so on.

Depending on the type of covariates, subgroup meta-analysis or meta-regression may be used to explore the between-study heterogeneity. Subgroup meta-analysis is commonly used with categorical covariates, whereas meta-regression is used when at least one of the covariates is continuous.

In subgroup meta-analysis or simply subgroup analysis, the studies are grouped based on study or participants' characteristics, and an overall effect-size estimate is computed for each group. The goal of subgroup analysis is to compare these overall estimates across groups and determine whether the considered grouping helps explain some of the observed between-study heterogeneity. Note that subgroup analysis can be viewed as a special case of a meta-regression with only one categorical moderator.

For more details about subgroup analysis, see the `subgroup()` option in [META] [meta summarize](#) and [META] [meta forestplot](#).

Meta-regression

Meta-regression explores a relationship between the study-specific effect sizes and the study-level covariates, such as a latitude of a study location or a dosage of a drug. These covariates are often referred to as moderators. See, for instance, [Greenland \(1987\)](#), [Berkey et al. \(1995\)](#), [Thompson and Sharp \(1999\)](#), [Thompson and Higgins \(2002\)](#), and [Viechtbauer et al. \(2015\)](#) for more information about meta-regression.

Two types of meta-regression are commonly considered in the meta-analysis literature: fixed-effects meta-regression and random-effects meta-regression.

An FE meta-regression ([Greenland 1987](#)) assumes that all heterogeneity between the study outcomes can be accounted for by the specified moderators. Let \mathbf{x}_j be a $p \times 1$ vector of moderators with the corresponding unknown coefficient vector, β . An FE meta-regression is given by

$$\hat{\theta}_j = \mathbf{x}_j\beta + \epsilon_j \quad \text{weighted by } w_j = \frac{1}{\hat{\sigma}_j^2}, \text{ where } \epsilon_j \sim N(0, \hat{\sigma}_j^2)$$

A traditional FE meta-regression does not model residual heterogeneity, but it can be incorporated by multiplying each of the variances, $\hat{\sigma}_j^2$, by a common factor. This model is known as an FE meta-regression with a multiplicative dispersion parameter or a multiplicative FE meta-regression ([Thompson and Sharp 1999](#)).

An RE meta-regression ([Berkey et al. 1995](#)) can be viewed as a meta-regression that incorporates the residual heterogeneity via an additive error term, which is represented in a model by a study-specific random effect. These random effects are assumed to be normal with mean zero and variance τ^2 , which estimates the remaining between-study heterogeneity that is unexplained by the considered moderators. An RE meta-regression is

$$\hat{\theta}_j = \mathbf{x}_j\beta + u_j + \epsilon_j \quad \text{weighted by } w_j^* = \frac{1}{\hat{\sigma}_j^2 + \hat{\tau}^2}, \text{ where } u_j \sim N(0, \tau^2) \text{ and } \epsilon_j \sim N(0, \hat{\sigma}_j^2)$$

For more details about meta-regression, see [META] [meta regress](#) and [META] [meta regress postestimation](#).

Publication bias

Publication bias or, more generally, reporting bias occurs when the studies selected for a scientific review are systematically different from all available relevant studies. Specifically, publication bias is known in the meta-analysis literature as an association between the likelihood of a publication and the statistical significance of a study result. The rise of systematic reviews for summarizing the results of scientific studies elevated the importance of acknowledging and addressing publication bias in research. Publication bias typically arises when nonsignificant results are being underreported in the literature (for example, [Rosenthal \[1979\]](#); [Iyengar and Greenhouse \[1988\]](#); [Begg and Berlin \[1988\]](#); [Hedges \[1992\]](#); [Stern and Simes \[1997\]](#); [Givens, Smith, and Tweedie \[1997\]](#); [Sutton et al. \[2000b\]](#); and [Kicinski, Springate, and Kontopantelis \[2015\]](#)).

Suppose that we are missing some of the studies in our meta-analysis. If these studies are simply a random sample of all the studies that are relevant to our research question, our meta-analytic results will remain valid but will not be as precise. That is, we will likely obtain wider confidence intervals and less powerful tests. However, if the missing studies differ systematically from our observed studies, such as when smaller studies with nonsignificant findings are suppressed from publication, our meta-analytic results will be biased toward a significant result. Any health-policy or clinical decisions based on them will be invalid.

[Dickersin \(2005\)](#) notes that to avoid potentially serious consequences of publication bias, many researchers (for example, [Simes \[1986\]](#); [Dickersin \[1988\]](#); [Hetherington et al. \[1989\]](#); [Dickersin and Rennie \[2003\]](#); [Antes and Chalmers \[2003\]](#); and [Krakovsky \[2004\]](#)) called for the registration of clinical trials worldwide at the outset to keep track of the findings, whether or not significant, from all trials. Although this may not necessarily eradicate the problem of publication bias, this will make it more difficult for the results of smaller trials to go undetected. Generally, when one selects the studies for meta-analysis, the review of the literature should be as comprehensive as possible, including searching the grey literature to uncover the relevant unpublished studies.

See [Borenstein et al. \(2009, chap. 30\)](#) for the summary of other factors for publication bias such as language bias and cost bias.

Funnel plots

The funnel plot ([Light and Pillemer 1984](#)) is commonly used to explore publication bias ([Sterne, Becker, and Egger 2005](#)). It is a scatterplot of the study-specific effect sizes versus measures of study precision. In the absence of publication bias, the shape of the scatterplot should resemble a symmetric inverted funnel. The funnel-plot asymmetry, however, may be caused by factors other than publication bias such as a presence of a moderator correlated with the study effect and study size or, more generally, the presence of substantial between-study heterogeneity ([Egger et al. 1997](#); [Peters et al. 2008](#); [Sterne et al. 2011](#)). The so-called contour-enhanced funnel plots have been proposed to help discriminate between the funnel-plot asymmetry because of publication bias versus other reasons.

See [\[META\] meta funnelplot](#) for details.

Tests for funnel-plot asymmetry

Graphical evaluation of funnel plots is useful for data exploration but may be subjective when detecting the asymmetry. Statistical tests provide a more formal evaluation of funnel-plot asymmetry. These tests are also known as tests for small-study effects ([Sterne, Gavaghan, and Egger 2000](#)) and, historically, as tests for publication bias. The tests are no longer referred to as “tests for publication bias” because, as we commented earlier, the presence of the funnel-plot asymmetry may not necessarily be attributed to publication bias, particularly in the presence of substantial between-study variability. See [Harbord, Harris, and Sterne \(2016\)](#) for a summary of these tests.

Two types of tests for funnel-plot asymmetry are considered in the literature: regression-based tests (Egger et al. 1997; Harbord, Egger, and Sterne 2006; and Peters et al. 2006) and a nonparametric rank-based test (Begg and Mazumdar 1994). These tests explore the relationship between the study-specific effect sizes and study precision. The presence of the funnel-plot asymmetry is declared when the association between the two measures is greater than what would have been observed by chance.

For more details regarding the tests of funnel-plot asymmetry, see [META] [meta bias](#).

The trim-and-fill method

Tests for funnel-plot asymmetry are useful for detecting publication bias but are not able to estimate the impact of this bias on the final meta-analysis results. The nonparametric trim-and-fill method of Duval and Tweedie (2000a, 2000b) provides a way to assess the impact of missing studies because of publication bias on the meta-analysis. It evaluates the amount of potential bias present in meta-analysis and its impact on the final conclusion. This method is typically used as a sensitivity analysis to the presence of publication bias.

See [META] [meta trimfill](#) for more information about the trim-and-fill method.

Cumulative meta-analysis

Cumulative meta-analysis provides the results of multiple meta-analyses, where each analysis is performed by adding one study at a time. It is useful to identify various trends in the overall effect sizes. For example, when the studies are ordered chronologically, one can determine the point in time of the potential change in the direction or significance of the effect size. A well-known example of a cumulative meta-analysis is presented in *Cumulative meta-analysis* of [META] [meta](#) for the study of the efficacy of streptokinase after a myocardial infarction (Lau et al. 1992). Also see the `cumulative()` option in [META] [meta summarize](#) and [META] [meta forestplot](#).

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Also see

[META] [meta](#) — Introduction to meta

[META] [Glossary](#)

Title

meta — Introduction to meta

Description
Also see

Remarks and examples

Acknowledgments

References

Description

The `meta` command performs meta-analysis. In a nutshell, you can do the following:

1. Compute or specify effect sizes; see [META] [meta esize](#) and [META] [meta set](#).
2. Summarize meta-analysis data; see [META] [meta summarize](#) and [META] [meta forestplot](#).
3. Perform meta-regression to address heterogeneity; see [META] [meta regress](#).
4. Explore small-study effects and publication bias; see [META] [meta funnelplot](#), [META] [meta bias](#), and [META] [meta trimfill](#).

For software-free introduction to meta-analysis, see [META] [Intro](#).

Declare, update, and describe `meta data`

<code>meta data</code>	Declare meta-analysis data
<code>meta esize</code>	Compute effect sizes and declare <code>meta data</code>
<code>meta set</code>	Declare <code>meta data</code> using precalculated effect sizes
<code>meta update</code>	Update current settings of <code>meta data</code>
<code>meta query</code>	Describe current settings of <code>meta data</code>
<code>meta clear</code>	Clear current settings of <code>meta data</code>

Summarize `meta data` by using a table

<code>meta summarize</code>	Summarize meta-analysis data
<code>meta summarize, subgroup()</code>	Perform subgroup meta-analysis
<code>meta summarize, cumulative()</code>	Perform cumulative meta-analysis

Summarize `meta data` by using a forest plot

<code>meta forestplot</code>	Produce meta-analysis forest plots
<code>meta forestplot, subgroup()</code>	Produce subgroup meta-analysis forest plots
<code>meta forestplot, cumulative()</code>	Produce cumulative meta-analysis forest plots

Explore heterogeneity and perform meta-regression

<code>meta labbeplot</code>	Produce L'Abbé plots for binary data
<code>meta regress</code>	Perform meta-regression
<code>estat bubbleplot</code>	Produce bubble plots after meta-regression

Explore and address small-study effects (funnel-plot asymmetry, publication bias)

<code>meta funnelplot</code>	Produce funnel plots
<code>meta funnelplot, contours()</code>	Produce contour-enhanced funnel plots
<code>meta bias</code>	Test for small-study effects or funnel-plot asymmetry
<code>meta trimfill</code>	Perform trim-and-fill analysis of publication bias

Remarks and examples

This entry describes Stata's suite of commands, `meta`, for performing meta-analysis. For a software-free introduction to meta-analysis, see [META] [Intro](#).

Remarks are presented under the following headings:

Introduction to meta-analysis using Stata

Example datasets

Effects of teacher expectancy on pupil IQ (pupiliq.dta)

Effect of streptokinase after a myocardial infarction (strepto.dta)

Efficacy of BCG vaccine against tuberculosis (bcg.dta)

Effectiveness of nonsteroidal anti-inflammatory drugs (nsaids.dta)

Tour of meta-analysis commands

Prepare your data for meta-analysis in Stata

Basic meta-analysis summary

Subgroup meta-analysis

Cumulative meta-analysis

Heterogeneity: Meta-regression and bubble plot

Funnel plots for exploring small-study effects

Testing for small-study effects

Trim-and-fill analysis for addressing publication bias

Introduction to meta-analysis using Stata

Stata's `meta` command offers full support for meta-analysis from computing various effect sizes and producing basic meta-analytic summary and forest plots to accounting for between-study heterogeneity and potential publication bias. Random-effects, common-effect, and fixed-effects meta-analyses are supported.

Standard effect sizes for binary data, such as the log odds-ratio, or for continuous data, such as Hedges's g , may be computed using the `meta esize` command; see [META] [meta esize](#). Generic (precalculated) effect sizes may be specified by using the `meta set` command; see [META] [meta set](#).

`meta esize` and `meta set` are part of the meta-analysis declaration step, which is the first step of meta-analysis in Stata. During this step, you specify the main information about your meta-analysis such as the study-specific effect sizes and their corresponding standard errors and the meta-analysis model and method. This information is then automatically used by all subsequent `meta` commands

for the duration of your meta-analysis session. You can use `meta update` to easily update some of the specified information during the session; see [META] **meta update**. And you can use `meta query` to remind yourself about the current `meta settings` at any point of your meta-analysis; see [META] **meta update**. For more information about the declaration step, see [META] **meta data**. Also see *Prepare your data for meta-analysis in Stata*.

Random-effects, common-effect, and fixed-effects meta-analysis models are supported. You can specify them during the declaration step and use the same model throughout your meta-analysis or you can specify a different model temporarily with any of the `meta` commands. You can also switch to a different model for the rest of your meta-analysis by using `meta update`. See *Declaring a meta-analysis model* in [META] **meta data** for details.

Traditionally, meta-analysis literature and software used the term “fixed-effect model” (notice singular effect) to refer to the model that assumes a common effect for all studies. To avoid potential confusion with the term “fixed-effects model” (notice plural effects), which is commonly used in various disciplines to refer to the model whose effects vary from one group to another, we adopted the terminology from Rice, Higgins, and Lumley (2018) of the “common-effect model”. This terminology is also reflected in the option names for specifying the corresponding models with `meta` commands: `common` specifies a common-effect model and `fixed` specifies a fixed-effects model. (Similarly, `random` specifies a random-effects model.) Although the overall effect-size estimates from the common-effect and fixed-effects models are computationally identical, their interpretation is different. We provide the two options to emphasize this difference and to encourage proper interpretation of the final results given the specified model. See *common-effect versus fixed-effects models* in [META] **meta data** and *Meta-analysis models* in [META] **Intro** for more information.

Depending on the chosen meta-analysis model, various estimation methods are available: inverse-variance and Mantel–Haenszel for the common-effect and fixed-effects models and seven different estimators for the between-study variance parameter for the random-effects model. See *Declaring a meta-analysis estimation method* in [META] **meta data**.

Also see *Default meta-analysis model and method* in [META] **meta data** for the default model and method used by the `meta` commands.

Results of a basic meta-analysis can be summarized numerically in a table by using `meta summarize` (see [META] **meta summarize**) or graphically by using forest plots; see [META] **meta forestplot**. See *Basic meta-analysis summary*.

To evaluate the trends in the estimates of the overall effect sizes, you can use the `cumulative()` option with `meta summarize` or `meta forestplot` to perform cumulative meta-analysis. See *Cumulative meta-analysis*.

In the presence of `subgroup heterogeneity`, you can use the `subgroup()` option with `meta summarize` or `meta forestplot` to perform single or multiple subgroup analyses. See *Subgroup meta-analysis*.

Heterogeneity can also be explored by performing meta-regression using the `meta regress` command; see [META] **meta regress**. After meta-regression, you can produce bubble plots (see [META] **estat bubbleplot**) and perform other postestimation analysis (see [META] **meta regress postestimation**). With binary data, you can also use `meta labbeplot` to explore heterogeneity visually; see [META] **meta labbeplot**. Also see *Heterogeneity: Meta-regression and bubble plot*.

Publication bias, or more accurately, `small-study effects` or funnel-plot asymmetry, may be explored graphically via standard or contour-enhanced funnel plots (see [META] **meta funnelplot**). Regression-based and other tests for detecting small-study effects are available with the `meta bias` command; see [META] **meta bias**. The trim-and-fill method for assessing the potential impact of publication bias on the meta-analysis results is implemented in the `meta trimfill` command; see [META] **meta trimfill**.

See *Funnel plots for exploring small-study effects*, *Testing for small-study effects*, and *Trim-and-fill analysis for addressing publication bias*.

Example datasets

We present several datasets that we will use throughout the documentation to demonstrate the `meta` suite. Feel free to skip over this section to *Tour of meta-analysis commands* and come back to it later for specific examples.

Example datasets are presented under the following headings:

Effects of teacher expectancy on pupil IQ (pupiliq.dta)
Effect of streptokinase after a myocardial infarction (strepto.dta)
Efficacy of BCG vaccine against tuberculosis (bcg.dta)
Effectiveness of nonsteroidal anti-inflammatory drugs (nsaids.dta)

Effects of teacher expectancy on pupil IQ (pupiliq.dta)

This example describes a well-known study of [Rosenthal and Jacobson \(1968\)](#) that found the so-called Pygmalion effect, in which expectations of teachers affected outcomes of their students. A group of students was tested and then divided randomly into experimentals and controls. The division may have been random, but the teachers were told that the students identified as experimentals were likely to show dramatic intellectual growth. A few months later, a test was administered again to the entire group of students. The experimentals outperformed the controls.

Subsequent researchers attempted to replicate the results, but many did not find the hypothesized effect.

[Raudenbush \(1984\)](#) did a meta-analysis of 19 studies and hypothesized that the Pygmalion effect might be mitigated by how long the teachers had worked with the students before being told about the nonexistent higher expectations for the randomly selected subsample of students. We explore this hypothesis in *Subgroup meta-analysis*.

The data are saved in `pupiliq.dta`. Below, we describe some of the variables that will be used in later analyses.

```
. use https://www.stata-press.com/data/r16/pupiliq
(Effects of teacher expectancy on pupil IQ)

. describe
Contains data from https://www.stata-press.com/data/r16/pupiliq.dta
  obs:          19                Effects of teacher expectancy
                                on pupil IQ
  vars:         14                24 Apr 2019 08:28
                                (_dta has notes)
```

variable name	storage type	display format	value label	variable label
study	byte	%9.0g		Study number
author	str20	%20s		Author
year	int	%9.0g		Publication year
nexper	int	%9.0g		Sample size in experimental group
ncontrol	int	%9.0g		Sample size in control group
stdmdiff	double	%9.0g		Standardized difference in means
weeks	byte	%9.0g		Weeks of prior teacher-student contact
catweek	byte	%9.0g	catwk	Weeks of prior contact (categorical)
week1	byte	%9.0g	catweek1	Prior teacher-student contact > 1 week
se	double	%10.0g		Standard error of stdmdiff
se_c	float	%9.0g		se from Pubbias book, p.322
setting	byte	%8.0g	testtype	Test setting
tester	byte	%8.0g	tester	Tester (blind or aware)
studylbl	str26	%26s		Study label

Sorted by:

Variables `stdmdiff` and `se` contain the effect sizes (standardized mean differences between the experimental and control groups) and their standard errors, respectively. Variable `weeks` records the number of weeks of prior contact between the teacher and the students. Its dichotomized version, `week1`, records whether the teachers spent more than one week with the students (high-contact group, `week1=1`) or one week and less (low-contact group, `week1=0`) prior to the experiment.

We perform basic meta-analysis summary of this dataset in [Basic meta-analysis summary](#) and explore the between-study heterogeneity of the results with respect to the amount of the teacher-student contact in [Subgroup meta-analysis](#).

This dataset is also used in [Examples of using meta summarize](#) of [META] [meta summarize](#), example 4 of [META] [meta forestplot](#), example 8 of [META] [meta funnelplot](#), and [Examples of using meta bias](#) of [META] [meta bias](#).

See [example 1](#) for the declaration of the `pupiliq.dta`. You can also use its predeclared version, `pupiliqset.dta`.

Effect of streptokinase after a myocardial infarction (`strepto.dta`)

Streptokinase is a medication used to break down clots. In the case of myocardial infarction (heart attack), breaking down clots reduces damage to the heart muscle.

[Lau et al. \(1992\)](#) conducted a meta-analysis of 33 studies performed between 1959 and 1988. These studies were of heart attack patients who were randomly treated with streptokinase or a placebo.

Lau et al. (1992) introduced cumulative meta-analysis to investigate the time when the effect of streptokinase became statistically significant. Studies were ordered by time, and as each was added to the analysis, standard meta-analysis was performed. See [Cumulative meta-analysis](#) for details.

The data are saved in `strepto.dta`.

```
. use https://www.stata-press.com/data/r16/strepto
(Effect of streptokinase after a myocardial infarction)
. describe
Contains data from https://www.stata-press.com/data/r16/strepto.dta
obs:                33                Effect of streptokinase after a
                                myocardial infarction
vars:                7                14 May 2019 18:24
                                (_dta has notes)
```

variable name	storage type	display format	value label	variable label
<code>study</code>	<code>str12</code>	<code>%12s</code>		Study name
<code>year</code>	<code>int</code>	<code>%10.0g</code>		Publication year
<code>ndeadt</code>	<code>int</code>	<code>%10.0g</code>		Number of deaths in treatment group
<code>nsurvt</code>	<code>int</code>	<code>%9.0g</code>		Number of survivors in treatment group
<code>ndeadc</code>	<code>int</code>	<code>%10.0g</code>		Number of deaths in control group
<code>nsurvc</code>	<code>int</code>	<code>%9.0g</code>		Number of survivors in control group
<code>studyplus</code>	<code>str13</code>	<code>%13s</code>		Study label for cumulative MA

Sorted by:

The outcome of interest was death from myocardial infarction. Variables `ndeadt` and `nsurvt` contain the numbers of deaths and survivals, respectively, in the treatment group and `ndeadc` and `nsurvc` contain those in the control (placebo) group.

See [example 5](#) for the declaration of the `strepto.dta`. You can also use its predeclared version, `streptoset.dta`.

Efficacy of BCG vaccine against tuberculosis (`bcg.dta`)

BCG vaccine is a vaccine used to prevent tuberculosis (TB). The vaccine is used worldwide. Efficacy has been reported to vary. [Colditz et al. \(1994\)](#) performed meta-analysis on the efficacy using 13 studies—all randomized trials—published between 1948 and 1980. The dataset, shown below, has been studied by, among others, [Berkey et al. \(1995\)](#), who hypothesized that the latitude of the study location might explain the variations in efficacy. We explore this via meta-regression in [Heterogeneity: Meta-regression and bubble plot](#).

The data are saved in `bcg.dta`. Below, we describe some of the variables we will use in future analyses.

```
. use https://www.stata-press.com/data/r16/bcg
(Efficacy of BCG vaccine against tuberculosis)

. describe

Contains data from https://www.stata-press.com/data/r16/bcg.dta
  obs:                13                Efficacy of BCG vaccine against
                                         tuberculosis
  vars:                11                1 May 2019 14:40
                                         (_dta has notes)
```

variable name	storage type	display format	value label	variable label
<code>trial</code>	byte	%9.0g		Trial number
<code>trialloc</code>	str14	%14s		Trial location
<code>author</code>	str21	%21s		Author
<code>year</code>	int	%9.0g		Publication year
<code>npost</code>	int	%9.0g		Number of TB positive cases in treated group
<code>nnegt</code>	long	%9.0g		Number of TB negative cases in treated group
<code>npsc</code>	int	%9.0g		Number of TB positive cases in control group
<code>nnegc</code>	long	%9.0g		Number of TB negative cases in control group
<code>latitude</code>	byte	%9.0g		Absolute latitude of the study location (in degrees)
<code>alloc</code>	byte	%10.0g	alloc	Method of treatment allocation
<code>study1bl</code>	str27	%27s		Study label

Sorted by: `trial`

Variables `npost` and `nnegt` contain the numbers of positive and negative TB cases, respectively, in the treatment group (vaccinated group) and `npsc` and `nnegc` contain those in the control group. Variable `latitude` records the latitude of the study location, which is a potential moderator for the vaccine efficacy. Studies are identified by `study1bl`, which records the names of the authors and the year of the publication for each study.

This dataset is also used in [example 3](#) of [\[META\] meta data, Examples of using meta forestplot](#) of [\[META\] meta forestplot](#), [example 1](#) of [\[META\] meta labbeplot, Examples of using meta regress](#) of [\[META\] meta regress](#), [Remarks and examples](#) of [\[META\] meta regress postestimation](#), and [Examples of using estat bubbleplot](#) of [\[META\] estat bubbleplot](#).

See [example 7](#) for the declaration of the `bcg.dta`. You can also use its predeclared version, `bcgset.dta`.

Effectiveness of nonsteroidal anti-inflammatory drugs (`nsaids.dta`)

Strains and sprains cause pain, and nonsteroidal anti-inflammatory drugs (NSAIDs) are used to treat it. How well do they work? People who study such things define success as a 50-plus percent reduction in pain. [Moore et al. \(1998\)](#) performed meta-analysis of 37 randomized trials that looked into successful pain reduction via NSAIDs. Following their lead, we will explore publication bias or, more generally, small-study effects in these data. See [Funnel plots for exploring small-study effects](#), [Testing for small-study effects](#), and [Trim-and-fill analysis for addressing publication bias](#).

The data are saved in `nsaids.dta`.

```
. use https://www.stata-press.com/data/r16/nsaids
(Effectiveness of nonsteroidal anti-inflammatory drugs)
. describe
Contains data from https://www.stata-press.com/data/r16/nsaids.dta
  obs:                37                Effectiveness of nonsteroidal
                                anti-inflammatory drugs
  vars:                5                24 Apr 2019 17:09
                                (_dta has notes)
```

variable name	storage type	display format	value label	variable label
<code>study</code>	byte	%8.0g		Study ID
<code>nstreat</code>	byte	%8.0g		Number of successes in the treatment arm
<code>nftreat</code>	byte	%9.0g		Number of failures in the treatment arm
<code>nscontrol</code>	byte	%8.0g		Number of successes in the control arm
<code>nfcontrol</code>	byte	%9.0g		Number of failures in the control arm

Sorted by:

Variables `nstreat` and `nftreat` contain the numbers of successes and failures, respectively, in the experimental group and `nscontrol` and `nfcontrol` contain those in the control group.

This dataset is also used in [Examples of using meta funnelplot](#) of [META] [meta funnelplot](#) and [example 3](#) of [META] [meta bias](#).

See [example 10](#) for the declaration of the `nsaids.dta`. You can also use its predeclared version, `nsaidsset.dta`.

Tour of meta-analysis commands

In this section, we provide a tour of Stata's meta-analysis (`meta`) commands with applications to several real-world datasets. We demonstrate the basic meta-analysis summary and a forest plot and explore heterogeneity via subgroup analysis using the [pupil IQ dataset](#). We then demonstrate cumulative meta-analysis using the [streptokinase dataset](#). We continue with more heterogeneity analyses of the [BCG dataset](#). Finally, we explore and address publication bias for the [NSAIDS dataset](#).

Examples are presented under the following headings:

- [Prepare your data for meta-analysis in Stata](#)
- [Basic meta-analysis summary](#)
- [Subgroup meta-analysis](#)
- [Cumulative meta-analysis](#)
- [Heterogeneity: Meta-regression and bubble plot](#)
- [Funnel plots for exploring small-study effects](#)
- [Testing for small-study effects](#)
- [Trim-and-fill analysis for addressing publication bias](#)

Prepare your data for meta-analysis in Stata

The first step of meta-analysis in Stata is to declare your data as `meta data`. During this step, we specify the main information needed for meta-analysis such as effect sizes and their standard errors. We declare this information once by using either `meta set` or `meta esize`, and it is then used by all `meta` commands. If needed, we can update our initial settings throughout the meta-analysis session by using `meta update`. The declaration step helps minimize potential mistakes and typing; see [META] `meta data` for details.

► Example 1: Set up your data for meta-analysis in Stata

Consider the pupil IQ dataset described in *Effects of teacher expectancy on pupil IQ (pupiliq.dta)*.

```
. use https://www.stata-press.com/data/r16/pupiliq
(Effects of teacher expectancy on pupil IQ)
. describe studylbl stdmdiff se week1
```

variable name	storage type	display format	value label	variable label
studylbl	str26	%26s		Study label
stdmdiff	double	%9.0g		Standardized difference in means
se	double	%10.0g		Standard error of stdmdiff
week1	byte	%9.0g	catweek1	Prior teacher-student contact > 1 week

First, we prepare our data for use with `meta` commands. The dataset contains precomputed effect sizes, standardized mean differences stored in variable `stdmdiff`, and their standard errors stored in variable `se`. We will use `meta set` to declare these data. (If we needed to compute the individual effect sizes and their standard errors from the available `summary data`, we would have used [META] `meta esize`.)

We specify the effect sizes `stdmdiff` and their standard errors `se` with `meta set`. We also specify the variable that contains the study labels in the `studylabel()` option and the effect-size label in the `eslabel()` option. These are optional but useful for displaying the study and effect-size labels instead of generic study numbers and the generic label `Effect size`.

```
. meta set stdmdiff se, studylabel(studylbl) eslabel(Std. Mean Diff.)
Meta-analysis setting information
Study information
  No. of studies: 19
  Study label: studylbl
  Study size: N/A
Effect size
  Type: Generic
  Label: Std. Mean Diff.
  Variable: stdmdiff
Precision
  Std. Err.: se
  CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML
```

The header reports that there are $K = 19$ studies in the meta-analysis and which variables contain the study labels, the effect sizes, and the standard errors. The output also shows that we will be using the random-effects model with the REML estimation method for our meta-analysis. This can be

changed by specifying options with either `meta set` or the `meta` command of interest; see [Declaring a meta-analysis model](#) in [\[META\] meta data](#).

`meta set` creates some system variables beginning with `_meta_` and stores some data characteristics. For example, the system variables `_meta_cil` and `_meta_ciu` store the lower and upper limits of the CIs for the effect sizes. See [System variables](#) in [\[META\] meta data](#) for details.

See [\[META\] meta set](#) for more information about the command.



Basic meta-analysis summary

In this section, we focus on basic meta-analysis summary by using [\[META\] meta summarize](#) and [\[META\] meta forestplot](#). See [Introduction of \[META\] meta summarize](#) and [Overview of \[META\] meta forestplot](#) for an overview of the meta-analysis summary and forest plots.

► Example 2: Meta-analysis summary

Continuing with [example 1](#), we use `meta summarize` to combine the studies and estimate the overall effect size.

```
. meta summarize
      Effect-size label: Std. Mean Diff.
      Effect size:      stdmdiff
      Std. Err.:       se
      Study label:     studylbl

Meta-analysis summary                Number of studies =      19
Random-effects model                 Heterogeneity:
Method: REML                          tau2 = 0.0188
                                       I2 (%) = 41.84
                                       H2 = 1.72
```

Effect Size: Std. Mean Diff.

Study	Effect Size	[95% Conf. Interval]		% Weight
Rosenthal et al., 1974	0.030	-0.215	0.275	7.74
Conn et al., 1968	0.120	-0.168	0.408	6.60
Jose & Cody, 1971	-0.140	-0.467	0.187	5.71
Pellegrini & Hicks, 1972	1.180	0.449	1.911	1.69
Pellegrini & Hicks, 1972	0.260	-0.463	0.983	1.72
Evans & Rosenthal, 1969	-0.060	-0.262	0.142	9.06
Fielder et al., 1971	-0.020	-0.222	0.182	9.06
Claiborn, 1969	-0.320	-0.751	0.111	3.97
Kester, 1969	0.270	-0.051	0.591	5.84
Maxwell, 1970	0.800	0.308	1.292	3.26
Carter, 1970	0.540	-0.052	1.132	2.42
Flowers, 1966	0.180	-0.257	0.617	3.89
Keshock, 1970	-0.020	-0.586	0.546	2.61
Henrikson, 1970	0.230	-0.338	0.798	2.59
Fine, 1972	-0.180	-0.492	0.132	6.05
Grieger, 1970	-0.060	-0.387	0.267	5.71
Rosenthal & Jacobson, 1968	0.300	0.028	0.572	6.99
Fleming & Anttonen, 1971	0.070	-0.114	0.254	9.64
Ginsburg, 1970	-0.070	-0.411	0.271	5.43
theta	0.084	-0.018	0.185	

Test of theta = 0: z = 1.62

Prob > |z| = 0.1052

Test of homogeneity: Q = chi2(18) = 35.83

Prob > Q = 0.0074

The output from the standard meta-analysis summary includes heterogeneity statistics, the individual and overall effect sizes, and other information. The estimate of the overall effect size θ is reported at the bottom of the table and labeled as `theta`. It is computed as the weighted average of study-specific effect sizes (standardized mean differences in our example). For these data, the overall estimate is 0.084 with a 95% CI of $[-0.018, 0.185]$. The significance test of $H_0: \theta = 0$ is reported below the table and has a p -value of 0.1052, which suggests that the overall effect size is not statistically significantly different from zero.

We should be careful with our inferential conclusions about θ because of the presence of between-study heterogeneity, as indicated, for instance, by the homogeneity test of $H_0: \theta_1 = \theta_2 = \dots = \theta_{19} = \theta$ reported following the significance test. Its Q test statistic is 35.83 with a p -value of 0.0074, from which we can infer that there is significant heterogeneity between the individual studies.

The presence of heterogeneity among studies can be inferred also from the heterogeneity statistics reported in the header. For instance, $I^2 = 41.84$ indicates that about 42% of the variability in the effect-size estimates is due to the differences between studies. The between-study heterogeneity must be addressed before final meta-analytic conclusions; see [Subgroup meta-analysis](#).

The table also reports the study-specific effect-sizes and their corresponding 95% CIs, but this information can be suppressed, if desired, by specifying the `nostudies` option.

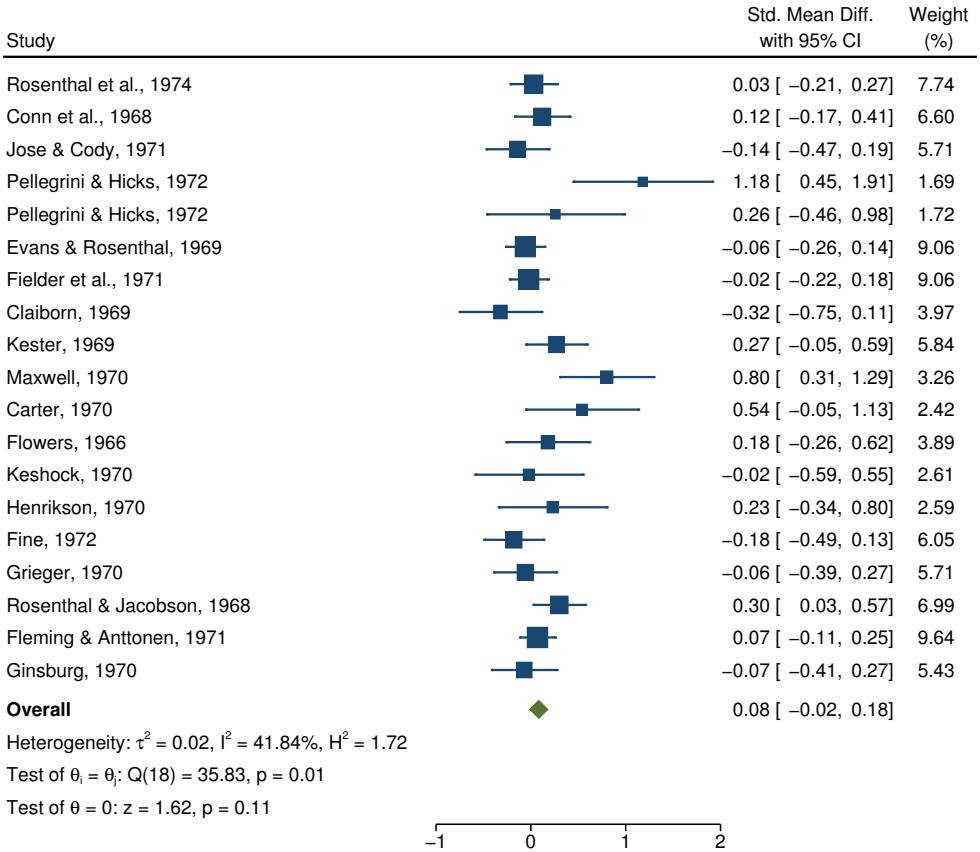
See [\[META\] meta summarize](#) for details.

◀

▷ Example 3: Forest plot

The results of meta-analysis are commonly displayed graphically using a forest plot. Continuing with [example 2](#), we can use `meta forestplot` to produce a meta-analysis forest plot for the pupil IQ data.

```
. meta forestplot
      Effect-size label: Std. Mean Diff.
            Effect size: stdmdiff
            Std. Err.: se
            Study label: studylbl
```



Random-effects REML model

We obtain the same meta-analysis summary as with `meta summarize` in [example 2](#), but it is now displayed on a graph. In addition to the estimated values, the effect sizes are displayed graphically as dark-blue squares centered at their estimates with areas proportional to the study weights and with horizontal lines or whiskers that represent the length of the corresponding CIs. The overall effect size is displayed as a green diamond with its width corresponding to the respective CI. (Notice that only the width and not the height of the diamond is relevant for the overall effect size.)

A forest plot provides an easy way to visually explore the agreement between the study-specific effect sizes and how close they are to the overall effect size. We can also spot the studies with large weights more easily by simply looking at the studies with large squares. In our example, the presence of between-study heterogeneity is evident—there are several studies whose effect-size estimates are very different from the overall estimate, and there are studies whose CIs do not even overlap.

See [\[META\] meta forestplot](#) for details.

Subgroup meta-analysis

In [example 2](#) and [example 3](#), we established the presence of between-study heterogeneity in the pupil IQ dataset. Sometimes, the differences between studies may be explained by study-level covariates available in the data. When these covariates are categorical, we can perform meta-analysis separately for each category, which is known as subgroup meta-analysis; see [Subgroup meta-analysis](#) of [META] [Intro](#).

▷ Example 4: Subgroup meta-analysis

[Raudenbush \(1984\)](#) suspected that the amount of time the teachers spent with students before the experiment could impact their susceptibility to researchers' test results about children's intellectual abilities. If so, we would expect the effect sizes to be negatively associated with the amount of contact.

Continuing with [example 2](#), we see that the dataset contains a binary variable `week1` that records whether the teachers spend more than one week with children (high-contact group) or one week and less (low-contact group). Let's perform meta-analysis separately for each group. Under Raudenbush's hypothesis, we should expect to see larger effect sizes in the low-contact group and smaller effect sizes in the high-contact group.

We use the `subgroup()` option with `meta summarize` to perform a separate analysis for each group of `week1`.


```
. meta summarize, subgroup(week1)
```

```
Effect-size label: Std. Mean Diff.
```

```
Effect size: stdmdiff
```

```
Std. Err.: se
```

```
Study label: studylbl
```

```
Subgroup meta-analysis summary
```

```
Number of studies = 19
```

```
Random-effects model
```

```
Method: REML
```

```
Group: week1
```

```
Effect Size: Std. Mean Diff.
```

Study	Effect Size	[95% Conf. Interval]		% Weight
Group: <= 1 week				
Pellegrini & Hicks, 1972	1.180	0.449	1.911	1.69
Pellegrini & Hicks, 1972	0.260	-0.463	0.983	1.72
Kester, 1969	0.270	-0.051	0.591	5.84
Maxwell, 1970	0.800	0.308	1.292	3.26
Carter, 1970	0.540	-0.052	1.132	2.42
Flowers, 1966	0.180	-0.257	0.617	3.89
Keshock, 1970	-0.020	-0.586	0.546	2.61
Rosenthal & Jacobson, 1968	0.300	0.028	0.572	6.99
theta	0.373	0.189	0.557	
Group: > 1 week				
Rosenthal et al., 1974	0.030	-0.215	0.275	7.74
Conn et al., 1968	0.120	-0.168	0.408	6.60
Jose & Cody, 1971	-0.140	-0.467	0.187	5.71
Evans & Rosenthal, 1969	-0.060	-0.262	0.142	9.06
Fielder et al., 1971	-0.020	-0.222	0.182	9.06
Claiborn, 1969	-0.320	-0.751	0.111	3.97
Henrikson, 1970	0.230	-0.338	0.798	2.59
Fine, 1972	-0.180	-0.492	0.132	6.05
Grieger, 1970	-0.060	-0.387	0.267	5.71
Fleming & Anttonen, 1971	0.070	-0.114	0.254	9.64
Ginsburg, 1970	-0.070	-0.411	0.271	5.43
theta	-0.021	-0.102	0.059	
Overall				
theta	0.084	-0.018	0.185	

```
Heterogeneity summary
```

Group	df	Q	P > Q	tau2	% I2	H2
<= 1 week	7	11.20	0.130	0.015	22.40	1.29
> 1 week	10	6.40	0.780	0.000	0.00	1.00
Overall	18	35.83	0.007	0.019	41.84	1.72

```
Test of group differences: Q_b = chi2(1) = 14.77
```

```
Prob > Q_b = 0.000
```

Indeed, if we look at the overall effect-size estimates for each group, the low-contact group has a larger estimate of 0.373 with a 95% CI of [0.189, 0.557], which suggests a statistically significant effect in this group, whereas the high-contact group has a smaller estimate of -0.021 with a 95% CI of $[-0.102, 0.059]$, which suggests that the effect in this group is not different from 0 at a 5% significance level. Clearly, the amount of teacher contact with students has an impact on the meta-analysis results.

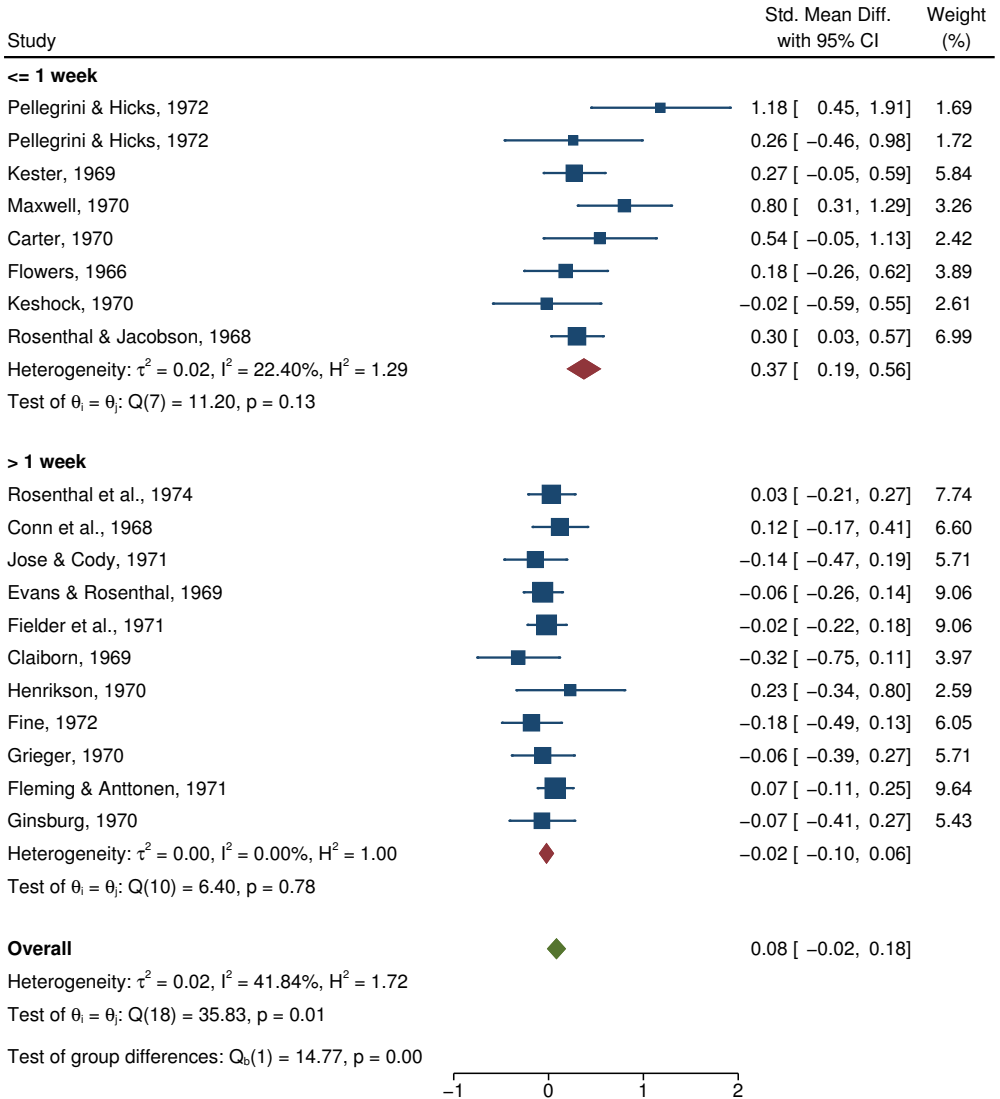
If we look at the heterogeneity summary reported following the main table, we will see that heterogeneity is reduced within each group. It is essentially nonexistent in the high-contact group and is much smaller (for instance, $I^2 = 22\%$ versus the earlier $I^2 = 42\%$) in the low-contact group.

The test of group differences (with $Q_b = 14.77$ and the corresponding p -value of 0.000) reported at the bottom of the output also indicates that the group-specific overall effect sizes are statistically different.

We can also present the results of our subgroup analysis graphically by using the `subgroup()` option with `meta forest`:

```
. meta forestplot, subgroup(week1)

Effect-size label: Std. Mean Diff.
Effect size: stdmdiff
Std. Err.: se
Study label: studylbl
```



Random-effects REML model

It appears that stratifying our meta-analysis on the amount of prior contact between students and teachers explained most of the variability in the magnitudes of the effect sizes, at least in the high-contact group.

When interpreting results from subgroup analysis, we should be mindful that the results are based on fewer studies and thus may not be as precise, in general.

See [META] [meta summarize](#) and [META] [meta forestplot](#).

Cumulative meta-analysis

Cumulative meta-analysis performs multiple meta-analyses by accumulating studies one at a time after ordering them with respect to the variable of interest. This analysis is useful to monitor the trend in the estimates of the overall effect sizes with respect to some factor. For instance, it may be used to detect the time when the effect size of interest became significant.

► Example 5: Computing log odds-ratios using meta esize

Consider the streptokinase dataset described in [Effect of streptokinase after a myocardial infarction \(strepto.dta\)](#).

```
. use https://www.stata-press.com/data/r16/strepto, clear
(Effect of streptokinase after a myocardial infarction)
. describe
Contains data from https://www.stata-press.com/data/r16/strepto.dta
  obs:                33                Effect of streptokinase after a
                                         myocardial infarction
  vars:                7                14 May 2019 18:24
                                         (_dta has notes)
```

variable name	storage type	display format	value label	variable label
study	str12	%12s		Study name
year	int	%10.0g		Publication year
ndeadt	int	%10.0g		Number of deaths in treatment group
nsurvt	int	%9.0g		Number of survivors in treatment group
ndeadc	int	%10.0g		Number of deaths in control group
nsurvc	int	%9.0g		Number of survivors in control group
studyplus	str13	%13s		Study label for cumulative MA

Sorted by:

As in [example 1](#), first we prepare our data for use with `meta` commands. Our dataset contains the summary data that represent the study-specific 2×2 tables. The variables `ndeadt`, `nsurvt`, `ndeadc`, and `nsurvc` record the numbers of deaths and survivors in the treatment and control groups.

[Lau et al. \(1992\)](#) considered an odds ratio as the effect size of interest for these data. For odds ratios, meta-analysis is performed in the log metric. We can use `meta esize` to compute study-specific log odds-ratios and their corresponding standard errors and declare them for the subsequent meta-analysis. To compute log odds-ratios, we specify the four variables containing table cell counts with `meta esize`. As with `meta set` in [example 1](#), we specify the study labels in the `studylabel()` option with `meta esize`.

```

. meta esize ndeadt nsurvt ndeadc nsurvc, studylabel(studyplus) common
Meta-analysis setting information
Study information
  No. of studies: 33
  Study label: studyplus
  Study size: _meta_studysize
  Summary data: ndeadt nsurvt ndeadc nsurvc
  Effect size
    Type: lnratio
    Label: Log Odds-Ratio
    Variable: _meta_es
  Zero-cells adj.: 0.5, only0
  Precision
    Std. Err.: _meta_se
    CI: [_meta_cil, _meta_ciu]
    CI level: 95%
Model and method
  Model: Common-effect
  Method: Mantel-Haenszel

```

`meta esize` reports that there are 33 trials and that the computed effect size is log odds-ratio. This is the default effect size with binary outcomes. You can specify other effect sizes in the `esize()` option, which include a log risk-ratio, risk difference, and log Peto's odds-ratio. (After the declaration, you can use `meta update` to change the effect size more easily without having to respecify your summary data variables; see [META] [meta update](#).)

Lau et al. (1992) used a common-effect model with the Mantel–Haenszel method to perform their cumulative meta-analysis. We will follow their approach. Thus, we also specified the `common` option with `meta esize`. The command reported that the assumed meta-analysis model is a common-effect model. The Mantel–Haenszel estimation method is the default method for log odds-ratios under a common-effect model.

◀

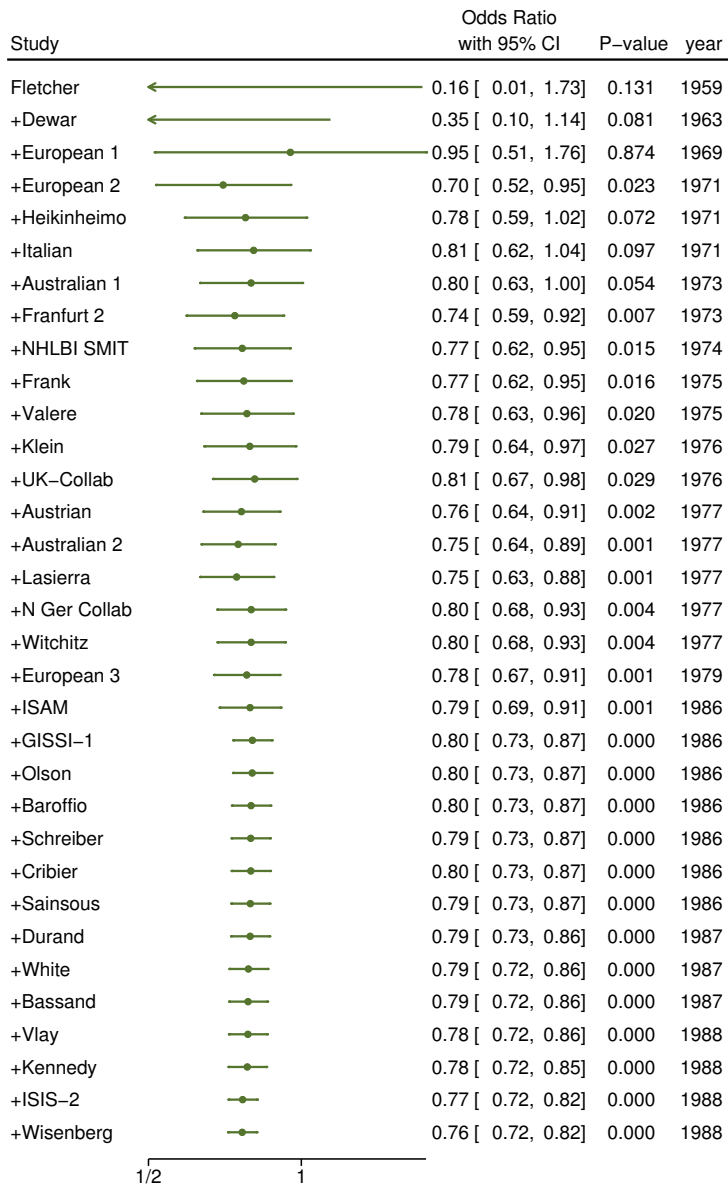
► Example 6: Cumulative meta-analysis

After the data declaration in [example 5](#), we are ready to perform the cumulative meta-analysis. Lau et al. (1992) used cumulative meta-analysis to investigate the trends in the effect of the streptokinase drug used to prevent death after a myocardial infarction. We replicate their analysis below by producing a cumulative meta-analysis plot over the years for these data. Also see Borenstein, Hedges, Higgins, and Rothstein (2009) for the analysis of these data.

We use the `meta forestplot` command with the `cumulative()` option. We use the `or` option to display odds ratios instead of the default log odds-ratios. To match figure 1 in Lau et al. (1992) more closely, we also specify the `crop(0.5 .)` option to crop the lower CI limits and log odds-ratios estimates that are smaller than 0.5.

```
. meta forestplot, cumulative(year) or crop(0.5 .)
```

```
Effect-size label: Log Odds-Ratio
Effect size: _meta_es
Std. Err.: _meta_se
Study label: studyplus
```



Common-effect Mantel-Haenszel model

The cumulative meta-analysis forest plot displays the overall effect-size estimates and the corresponding CIs computed for the first study, for the first two studies, for the first three studies, and so on. The point estimates are represented by green circles, and the CIs are represented by the CI lines. The change in style and color of the plotted markers emphasizes that the (cumulative) overall effect sizes and not the study-specific effect sizes are being plotted.

The “+” sign in front of the study label we used for this analysis (variable `studyplus`) indicates that each subsequent study is being added to the previous ones for each analysis. In addition to the ordered values of the specified variable of interest (`year` in our example), the plot also displays the p -values corresponding to the tests of significance of the computed overall effect sizes.

For example, the cumulative odds ratio in the fourth row marked as `+European 2` is 0.70 with a 95% CI of $[0.52, 0.95]$ and a p -value of 0.023. So, based on the first four trials, the overall odds of death is roughly 30% less in the treatment group (treated with streptokinase) compared with the placebo group.

Notice that the first two odds-ratio estimates (and their lower CI limits) are smaller than 0.5. Because we used the `crop(0.5 .)` option, their values are not displayed on the graph. Instead, the arrowheads are displayed at the lower ends of the CI lines to indicate that the lower limits and the effect-size estimates are smaller than 0.5.

Borenstein, Hedges, Higgins, and Rothstein (2009) states that with the inclusion of additional trials in the cumulative meta-analysis, the overall effect sizes become more uniform because the chance of any new trial reporting a drastically different overall effect size is low. Also, the CIs become more narrow because the precision increases as more data become available.

If we look back at the plot, we will notice that starting from 1977, the overall effect size becomes (and stays) highly significant over the next decade of additional trials. Lau et al. (1992) and Borenstein et al. (2009, chap. 42) noted that if cumulative meta-analysis was used at that time to monitor the accumulated evidence from the trials, perhaps, the benefits from streptokinase could have been adopted in practice as early as 1977.

We can also obtain the same results as above but in a table by using `meta summarize`.

```

. meta summarize, cumulative(year) or
  Effect-size label: Log Odds-Ratio
    Effect size:   _meta_es
    Std. Err.:   _meta_se
    Study label:  studyplus

Cumulative meta-analysis summary          Number of studies =    33
Common-effect model
Method: Mantel-Haenszel
Order variable: year

```

Study	Odds Ratio	[95% Conf. Interval]	P-value	year	
Fletcher	0.159	0.015	1.732	0.131	1959
+Dewar	0.345	0.104	1.141	0.081	1963
+European 1	0.952	0.514	1.760	0.874	1969
+European 2	0.702	0.517	0.951	0.023	1971
+Heikinheimo	0.776	0.589	1.023	0.072	1971
+Italian	0.806	0.624	1.040	0.097	1971
+Australian 1	0.796	0.632	1.004	0.054	1973
+Frankfurt 2	0.740	0.594	0.921	0.007	1973
+NHLBI SMIT	0.765	0.616	0.950	0.015	1974
+Frank	0.770	0.623	0.953	0.016	1975
+Valere	0.781	0.635	0.962	0.020	1975
+Klein	0.792	0.644	0.974	0.027	1976
+UK-Collab	0.809	0.670	0.979	0.029	1976
+Austrian	0.762	0.641	0.906	0.002	1977
+Australian 2	0.751	0.636	0.887	0.001	1977
+Lasierra	0.746	0.632	0.881	0.001	1977
+N Ger Collab	0.797	0.683	0.930	0.004	1977
+Witchitz	0.797	0.683	0.928	0.004	1977
+European 3	0.781	0.673	0.906	0.001	1979
+ISAM	0.793	0.690	0.910	0.001	1986
+GISSI-1	0.801	0.734	0.874	0.000	1986
+Olson	0.800	0.733	0.873	0.000	1986
+Baroffio	0.796	0.730	0.869	0.000	1986
+Schreiber	0.795	0.729	0.867	0.000	1986
+Cribier	0.795	0.729	0.868	0.000	1986
+Sainsous	0.794	0.728	0.866	0.000	1986
+Durand	0.793	0.727	0.865	0.000	1987
+White	0.787	0.721	0.858	0.000	1987
+Bassand	0.785	0.721	0.856	0.000	1987
+Vlay	0.785	0.720	0.856	0.000	1988
+Kennedy	0.783	0.718	0.853	0.000	1988
+ISIS-2	0.766	0.718	0.817	0.000	1988
+Wisenberg	0.765	0.717	0.816	0.000	1988

See [META] [meta summarize](#) and [META] [meta forestplot](#).

4

Heterogeneity: Meta-regression and bubble plot

As we discussed in [Subgroup meta-analysis](#), when effect sizes vary greatly between different subgroups, one can perform separate meta-analysis on each subgroup to account for the between-study heterogeneity. But what if there is an association between the effect sizes and other study-level covariates or moderators that may be continuous? Meta-regression addresses this problem.

Meta-regression performs a weighted linear regression of effect sizes on moderators. Its goal is to investigate whether the differences between the effect sizes can be explained by one or more moderators.

See *Introduction* of [META] `meta regress`.

► Example 7: Computing log risk-ratios using meta esize

Consider the BCG dataset described in *Efficacy of BCG vaccine against tuberculosis (bcg.dta)*.

```
. use https://www.stata-press.com/data/r16/bcg, clear
(Efficacy of BCG vaccine against tuberculosis)
. describe studylbl npost nnegt npsc nnegc latitude
```

variable name	storage type	display format	value label	variable label
studylbl	str27	%27s		Study label
npost	int	%9.0g		Number of TB positive cases in treated group
nnegt	long	%9.0g		Number of TB negative cases in treated group
npsc	int	%9.0g		Number of TB positive cases in control group
nnegc	long	%9.0g		Number of TB negative cases in control group
latitude	byte	%9.0g		Absolute latitude of the study location (in degrees)

As in [example 5](#), this dataset also records summary data for binary outcomes, so we will again use `meta esize` to compute our effect sizes.

In this example, our effect size of interest is a risk ratio. Just like with odds ratios, the meta-analysis of risk ratios is performed in the log metric, so we will be computing log risk-ratios.

```
. meta esize npost nnegt npsc nnegc, esize(lnrratio) studylabel(studylbl)
Meta-analysis setting information
Study information
  No. of studies: 13
  Study label: studylbl
  Study size: _meta_studysize
  Summary data: npost nnegt npsc nnegc
Effect size
  Type: lnrratio
  Label: Log Risk-Ratio
  Variable: _meta_es
Zero-cells adj.: None; no zero cells
Precision
  Std. Err.: _meta_se
  CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML
```

Our specification of `meta esize` is similar to that from [example 5](#), except here we specify the `esize(lnrratio)` option to compute log risk-ratios instead of the default log odds-ratios.

The output indicates that there are $K = 13$ studies in the meta-analysis and the default random-effects meta-analysis model (with the REML estimation method) will be used.



Now that we declared our data as `meta data`, we can proceed with meta-regression.

▷ Example 8: Meta-regression

The efficacy of the BCG vaccine against TB may depend on many factors such as the presence of environmental mycobacteria that provides some immunity to TB. [Berkey et al. \(1995\)](#) suggested that the distance of a study from the equator (the absolute latitude) may be used as a proxy for the presence of environmental mycobacteria and perhaps explain the lower efficacy of the BCG vaccine against TB in some studies. [Borenstein et al. \(2009\)](#) also commented that, in hotter climates, the vaccine may lose potency and certain bacteria necessary for the vaccine to work well are less likely to survive with more exposure to sunlight.

Following [Berkey et al. \(1995\)](#), we will explore these observations by using `meta regress` with the centered latitude as the moderator.

First, we generate a new variable, `latitude_c`, that is the mean-centered version of `latitude`. The mean value of `latitude`, 33.46, can be thought of as the latitude of the city of Atlanta in the United States or the city of Beirut in Lebanon.

```
. summarize latitude, meanonly
. generate double latitude_c = latitude - r(mean)
. label variable latitude_c "Mean-centered latitude"
```

We then fit meta-regression with `latitude_c` as the moderator.

```
. meta regress latitude_c
Effect-size label:  Log Risk-Ratio
Effect size:      _meta_es
Std. Err.:       _meta_se

Random-effects meta-regression
Method: REML

Number of obs =      13
Residual heterogeneity:
    tau2 =    .07635
    I2 (%) =   68.39
    H2 =      3.16
    R-squared (%) = 75.63
Wald chi2(1) =    16.36
Prob > chi2 =    0.0001
```

<code>_meta_es</code>	Coef.	Std. Err.	z	P> z	[95% Conf. Interval]	
<code>latitude_c</code>	-.0291017	.0071953	-4.04	0.000	-.0432043	-.0149991
<code>_cons</code>	-.7223204	.1076535	-6.71	0.000	-.9333174	-.5113234

```
Test of residual homogeneity: Q_res = chi2(11) = 30.73  Prob > Q_res = 0.0012
```

The regression coefficient for `latitude_c` is -0.0291 , which means that every one degree of latitude corresponds to a decrease of 0.0291 units in the log risk-ratio. In other words, the vaccine appears to work better in colder climates.

The proportion of between-study variance explained by the covariates can be assessed via the R^2 statistic. Here roughly 76% of the between-study variance is explained by the covariate `latitude_c`. From the value of I^2 in the output, roughly 68% of the residual variation is due to heterogeneity, which may potentially be explained by other covariates, with the other 32% due to the within-study sampling variability.

The test statistic for residual homogeneity, Q_{res} , is 30.73 with a p -value of 0.0012, so the null hypothesis of no residual heterogeneity is rejected, which is consistent with the reported residual heterogeneity summaries.

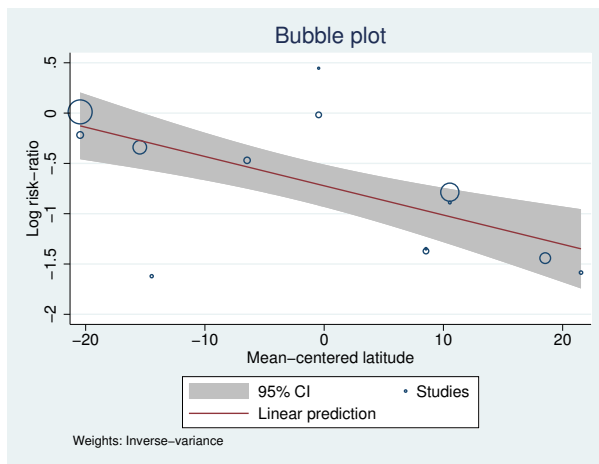
See [META] [meta regress](#) for more examples.

◀

► Example 9: Bubble plot

Whenever there is one continuous covariate in the meta-regression, we may explore the relationship between the effect sizes and that covariate via a bubble plot using the `estat bubbleplot` command. Continuing with [example 8](#), we explore the relationship between the log risk-ratios and `latitude_c`.

```
. estat bubbleplot
```



The bubble plot is a scatterplot of effect sizes and covariate values. Each study is represented by a circle with the size of the circle proportional to the effect-size precision, $1/\hat{\sigma}_j^2$. The fitted line (predicted log risk-ratios) is also plotted on the graph.

The log risk-ratio for the BCG vaccine decreases as the distance from the equator increases. The plot also reveals a few outlying studies that require more thorough investigation. We continue exploring this model in [META] [meta regress postestimation](#).

See [META] [estat bubbleplot](#).

◀

Funnel plots for exploring small-study effects

A funnel plot (Light and Pillemer 1984) plots study-specific effect sizes against measures of study precision such as standard errors. This plot is commonly used to explore publication bias or, more precisely, small-study effects. Small-study effects (Sterne, Gavaghan, and Egger 2000) arise when smaller studies tend to report different results such as larger effect-size estimates than larger studies. In the absence of small-study effects, the shape of the plot should resemble a symmetric inverted funnel.

Publication bias arises when smaller studies with nonsignificant findings are being suppressed from publication. It is one of the more common reasons for the presence of small-study effects, which leads to the asymmetry of the funnel plot. Another common reason for the asymmetry in the funnel plot is the presence of between-study heterogeneity.

See [Introduction](#) in [META] [meta funnelplot](#) for details.

► Example 10: Funnel plot

Let's explore the funnel-plot asymmetry for the NSAIDS dataset described in [Effectiveness of nonsteroidal anti-inflammatory drugs \(nsaids.dta\)](#).

```
. use https://www.stata-press.com/data/r16/nsaids, clear
(Effectiveness of nonsteroidal anti-inflammatory drugs)
. describe
Contains data from https://www.stata-press.com/data/r16/nsaids.dta
  obs:          37          Effectiveness of nonsteroidal
                             anti-inflammatory drugs
  vars:          5          24 Apr 2019 17:09
                             (_dta has notes)
```

variable name	storage type	display format	value label	variable label
study	byte	%8.0g		Study ID
nstreat	byte	%8.0g		Number of successes in the treatment arm
nftreat	byte	%9.0g		Number of failures in the treatment arm
nscontrol	byte	%8.0g		Number of successes in the control arm
nfcontrol	byte	%9.0g		Number of failures in the control arm

Sorted by:

As before, our first step is to declare our data. `nsaids.dta` records summary data for binary outcomes, so we will again use `meta esize` to compute our effect sizes as in [example 5](#) and [example 7](#).

Our effect size of interest is an odds ratio, so we can use the default specification of `meta esize`.

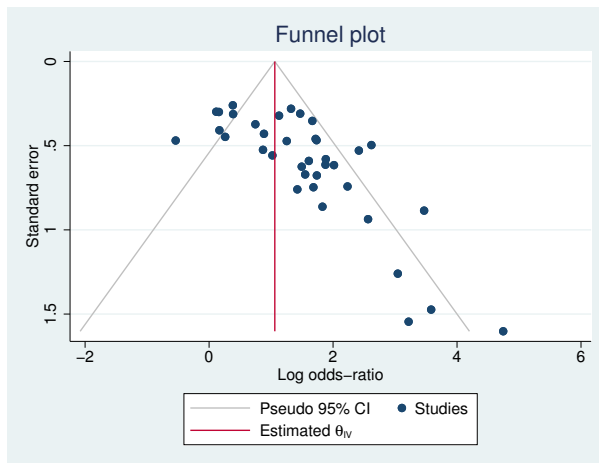
```
. meta esize nstreat-nfcontrol
Meta-analysis setting information
Study information
  No. of studies: 37
  Study label: Generic
  Study size: _meta_studysize
  Summary data: nstreat nftreat nscontrol nfcontrol
  Effect size
    Type: lnoratio
    Label: Log Odds-Ratio
    Variable: _meta_es
  Zero-cells adj.: 0.5, only0
  Precision
    Std. Err.: _meta_se
    CI: [_meta_cil, _meta_ciu]
    CI level: 95%
Model and method
  Model: Random-effects
  Method: REML
```

In the above, instead of listing all four variables with `meta esize` as we did in previous examples, we use one of the varlist shortcuts (see [U] 11.4 varname and varlists) to include all variables between `nstreat` and `nfcontrol`. We could do this because our variables appear in the dataset in the same order they need to be listed with `meta esize`: numbers of successes and failures in the treatment group followed by those in the control group.

There are $K = 37$ trials in this dataset. We will continue using the default random-effects meta-analysis model with the REML estimation method.

We use `meta funnelplot` to produce a funnel plot for the NSAIDS data.

```
. meta funnelplot
Effect-size label: Log Odds-Ratio
Effect size:      _meta_es
Std. Err.:       _meta_se
Model:           Common-effect
Method:          Inverse-variance
```



On a funnel plot, the more precise trials (with smaller standard errors) are displayed at the top of the funnel, and the less precise ones (with larger standard errors) are displayed at the bottom. The red reference line is plotted at the estimate of the overall effect size, the overall log odds-ratio in our example. In the absence of small-study effects, we would expect the points to be scattered around the reference line with the effect sizes from smaller studies varying more around the line than those from larger studies, forming the shape of an inverted funnel.

In our plot, there is an empty space in the bottom left corner. This suggests that the smaller trials with log odds-ratio estimates close to zero may be missing from the meta-analysis.

See [META] `meta funnelplot` for more examples.

◀

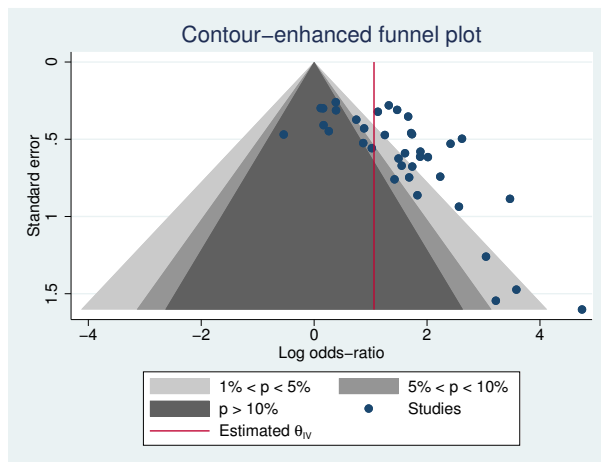
► Example 11: Contour-enhanced funnel plot

The asymmetry is evident in the funnel plot from [example 10](#), but we do not know the cause for this asymmetry. The asymmetry can be the result of publication bias or may be because of other reasons. The so-called contour-enhanced funnel plots can help determine whether the asymmetry of the funnel plot is because of publication bias. The contour lines that correspond to certain levels of

statistical significance (1%, 5%, and 10%) of tests of individual effects are overlaid on the funnel plot. Generally, publication bias is suspect when smaller studies are missing in the nonsignificant regions.

Let's add the 1%, 5%, and 10% significance contours to our funnel plot by specifying them in the `contours()` option.

```
. meta funnelplot, contours(1 5 10)
Effect-size label: Log Odds-Ratio
Effect size:      _meta_es
Std. Err.:       _meta_se
Model:           Common-effect
Method:          Inverse-variance
```



From this plot, we can see that the reported effects of almost all smaller trials (those at the bottom of the funnel) are statistically significant at a 5% level and less. On the other hand, a fair number of the larger trials (at the top of the funnel) reported nonsignificant results. For the funnel plot to look symmetric with respect to the reference line, we should have observed some trials in the middle and the bottom of the darkest region (with p -values larger than 10%). This suggests that we are missing some of the smaller trials with nonsignificant results, which would be consistent with the presence of publication bias.

There is also a chance that the funnel-plot asymmetry is induced by the between-study heterogeneity. Using a random-effects model and investigating the study-level covariates that may account for the heterogeneity should also be considered when exploring the funnel-plot asymmetry.

Also see [example 5](#) of [META] [meta funnelplot](#) for more details about this example.

◀

Testing for small-study effects

We can test for the presence of small-study effects or, technically, the asymmetry in the funnel plot more formally by using, for example, one of the regression-based tests. The main idea behind these tests is to determine whether there is a statistically significant association between the effect sizes and their measures of precision such as effect-size standard errors.

See [Introduction](#) in [META] [meta bias](#) for details.

▷ Example 12: Harbord's regression-based test

In [example 10](#), we investigated the funnel-plot asymmetry visually. Let's check for it more formally by using the `meta bias` command. We will use the Harbord regression-based test ([Harbord, Egger, and Sterne 2006](#)), which is often recommended when the effect size of interest is an odds ratio (or log odds-ratio).

To perform this test, we specify the `harbord` option with `meta bias`.

```
. meta bias, harbord
      Effect-size label:  Log Odds-Ratio
      Effect size:      _meta_es
      Std. Err.:       _meta_se

Regression-based Harbord test for small-study effects
Random-effects model
Method: REML

H0: beta1 = 0; no small-study effects
      beta1 =          3.03
SE of beta1 =         0.741
      z =            4.09
      Prob > |z| =     0.0000
```

The test uses a type of weighted regression that explores the relationship between the effect sizes and their precision. The slope in that regression, labeled as `beta1` in the output, describes the asymmetry of the funnel plot and represents the magnitude of the small-study effects. The further it is from zero, the more asymmetry is present in the funnel plot.

`meta bias` reports the z -test statistic of 4.09 with a p -value less than 0.0000 for the test of $H_0: \text{beta1}=0$ assuming a random-effects model with the REML estimation method. We have statistically significant evidence to reject the null hypothesis of the funnel-plot symmetry.

See [\[META\] meta bias](#).

◀

Trim-and-fill analysis for addressing publication bias

When the presence of publication bias is suspected, it is important to explore its impact on the final meta-analysis results. The trim-and-fill method of [Duval and Tweedie \(2000a, 2000b\)](#) provides a way to evaluate the impact of publication bias on the results. The idea of the method is to estimate the number of studies potentially missing because of publication bias, impute these studies, and use the observed and imputed studies to obtain the overall estimate of the effect size. This estimate can then be compared with the estimate obtained using only the observed studies. For details, see [Introduction](#) in [\[META\] meta trimfill](#).

▷ Example 13: Trim-and-fill analysis

From [example 11](#) and [example 12](#), we suspect the presence of publication bias in the meta-analysis of the NSAIDS data. Let's use the trim-and-fill method to investigate the impact of potentially missing studies on the estimate of the overall log odds-ratio.

We use the `meta trimfill` command. We specify the `eform` option (synonym for `or` when the computed effect sizes are log odds-ratios) to report the results as odds ratios instead of the default log odds-ratios. We also draw a contour-enhanced funnel plot that contains both the observed and imputed studies.

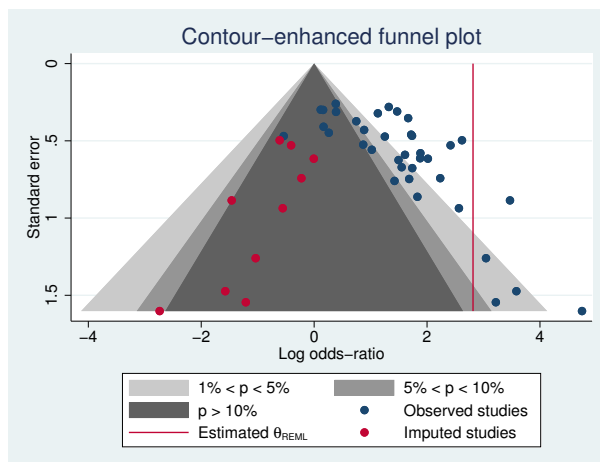
```
. meta trimfill, eform funnel(contours(1 5 10))
Effect-size label: Log Odds-Ratio
Effect size: _meta_es
Std. Err.: _meta_se

Nonparametric trim-and-fill analysis of publication bias
Linear estimator, imputing on the left

Iteration                                Number of studies =    47
Model: Random-effects                    observed =            37
Method: REML                             imputed =            10

Pooling
Model: Random-effects
Method: REML
```

Studies	Odds Ratio	[95% Conf. Interval]	
Observed	3.752	2.805	5.018
Observed + Imputed	2.815	2.067	3.832



`meta trimfill` reports that 10 hypothetical studies are estimated to be missing. When 10 studies are imputed and added to the meta-analysis, the overall odds ratio reduces from 3.752 (based on 37 observed studies) to 2.815 (based on 47 observed and imputed studies). This suggests that the treatment benefit as reported in the literature may be larger than it would be in the absence of publication bias.

From the funnel plot, almost all the imputed studies fall in the darkest-gray region corresponding to a p -value of more than 10%. This further supports the conclusion that the small-study effect is most likely because of publication bias.

See [META] [meta trimfill](#).

◀

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Also see

[META] [Intro](#) — Introduction to meta-analysis

[META] [Glossary](#)

Title

meta data — Declare meta-analysis data

[Description](#)

[Remarks and examples](#)

[References](#)

[Also see](#)

Description

This entry describes how to prepare your data for meta-analysis using the `meta` commands.

In a nutshell, do the following:

1. If you have access to [summary data](#), use `meta esize` to compute and declare effect sizes such as an odds ratio or a Hedges's g .
2. Alternatively, if you have only precomputed (generic) effect sizes, use `meta set`.
3. To update some of your meta-analysis settings after the declaration, use `meta update`.
4. To check whether your data are already meta set or to see the current meta settings, use `meta query`.

Remarks and examples

Remarks are presented under the following headings:

Overview

Declaring meta-analysis information

Declaring effect sizes and their precision

Declaring a meta-analysis model

Declaring a meta-analysis estimation method

Default meta-analysis model and method

Declaring a confidence level for meta-analysis

Declaring display settings for meta-analysis

Modifying default meta settings

Meta-analysis information

Meta settings with meta set

Meta settings with meta esize

System variables

Examples of data declaration for meta-analysis

Declaring precomputed effect sizes using meta set

Computing and declaring effect sizes using meta esize

Displaying and updating meta settings

Overview

The declaration of your data to be `meta` data is the first step of your meta-analysis in Stata. `meta` data are your original data that also store key variables and characteristics about your specifications, which will be used by all `meta` commands during your meta-analysis session. The declaration step helps minimize mistakes and saves you time—you only need to specify the necessary information once.

You can use `meta set` or `meta esize` to declare your data to be `meta` data. If you have access only to precomputed effect sizes and their standard errors, use `meta set`. If you have access to [summary data](#) such as means and standard deviations from individual studies, use `meta esize` to compute the effect sizes and their standard errors and declare them. The latter is preferable because it provides access to more features such as the Mantel–Haenszel estimation method with binary data, which needs access to the actual 2×2 tables and not only the effect sizes for the computations.

For example, suppose that you have variables `es` and `se`, which contain the effect sizes and the corresponding standard errors. You can use

```
. meta set es se
```

to declare your data, and all subsequent `meta` commands will automatically use these variables in the meta-analysis analysis.

To review the current meta settings or to check whether the data are `meta set`, you can use `meta query`; see [META] [meta update](#). After your data are declared, you can update some of the meta-analysis specifications by using `meta update`. If you wish to clear the meta settings after your meta-analysis analysis, you can use `meta clear`; see [META] [meta update](#).

Declaring meta-analysis information

Two main components of meta-analysis are study-specific effect sizes and their precision. You must specify them during declaration. Other important components include the underlying meta-analysis model and an estimation method. You can specify them during declaration or later during analysis or use Stata's defaults. You can also specify options that affect the output of the `meta` commands. Below, we describe how you can declare various meta-analysis information.

Declaring effect sizes and their precision

As we mentioned above, you must declare study-specific effect sizes and their precision. This is done differently for `meta set` and `meta esize`.

`meta esize` computes effect sizes and their standard errors from summary data and then declares them. `meta set` declares already precomputed effect sizes and their standard errors. Thus, to use `meta set`, you do not need summary data from each study, but you need them for `meta esize`. Some analysis may not be available after `meta set` such as the Mantel–Haenszel estimation method and Harbord's test for the funnel-plot asymmetry because they require access to summary data.

Effect sizes and their precision using `meta set`. To use `meta set`, you must specify variables containing study-specific effect sizes and their precision. There are two ways to specify the precision of the effect sizes. You can either specify a variable containing the standard errors,

```
. meta set es se
```

Or, instead of the standard errors, specify the confidence intervals, and `meta set` will compute the corresponding standard errors based on them:

```
. meta set es cil ciu
```

In the above, the specified CI variables will be assumed to correspond to the 95% CIs. You can change this by specifying the `civarlevel()` option:

```
. meta set es cil ciu, civarlevel(90)
```

But do not confuse `civarlevel()` with `level()`. The former affects the confidence level only for the specified CI variables. The latter specifies the confidence level for the meta-analysis.

Effect sizes and their precision using `meta esize`. To use `meta esize`, you must specify summary data for each study. The type of summary data you specify depends on the effect size you wish to compute and consequently on the outcome of interest in the original studies.

`meta esize` computes and declares various effect sizes for two-group comparison of continuous outcomes and of binary outcomes. For continuous outcomes, you must specify the number of observations, means, and standard deviations for each treatment group (group 1) and control group (group 2).

```
. meta esize n1 m1 sd1 n2 m2 sd2
```

To compute effect sizes and their standard errors, `meta esize` also needs to know the type of the effect size. The above assumes Hedges's g standardized mean difference, but you can specify others in the `esize()` option; see effect sizes for continuous outcomes in *Syntax* of [META] `meta esize`.

For binary outcomes, you must specify 2×2 contingency tables for each study. You specify them as follows. Each of the four cells is represented by a variable such that each row represents a 2×2 table from a specific study. For instance,

```
. meta esize n11 n12 n21 n22
```

The order in which you specify the four variables is important: the top-left cell first, the top-right cell next, followed by the bottom-left cell, and finally the bottom-right cell. The above computes the log odds-ratio as an effect size, but you can select a different effect size; see effect sizes for binary outcomes in *Syntax* of [META] `meta esize`.

Options affecting effect-size and precision computations with `meta esize`. Depending on the chosen effect size, `meta esize` provides alternative ways of computing effect sizes and their standard errors.

For the Hedges's g effect size, there are two ways to compute the bias-correction factor used in its formula. For consistency with meta-analysis literature, `meta esize` uses an approximation, but you can specify the `exact` option within `esize()` to use the exact computation:

```
. meta esize n1 m1 sd1 n2 m2 sd2, esize(hedgesg, exact)
```

Note that the `esize` command uses the exact computation.

Both Hedges's g and Cohen's d effect sizes support standard-error adjustment of [Hedges and Olkin \(1985\)](#) with `esize()`'s option `holkinse`:

```
. meta esize n1 m1 sd1 n2 m2 sd2, esize(cohend, holkinse)
```

For the (unstandardized) mean difference, you can choose to compute standard errors assuming unequal variance between the two groups:

```
. meta esize n1 m1 sd1 n2 m2 sd2, esize(mdiff, unequal)
```

For binary outcomes with log odds-ratios or log risk-ratios as effect sizes, `meta esize` automatically adjusts for zero cells when computing effect sizes. By default, it adds 0.5 to all cells of the 2×2 tables that contain at least one zero cell. You can specify other adjustments in the `zero cells(zcspec)` option. For example, with log odds-ratios, you can specify the treatment-arm continuity correction of [Sweeting, Sutton, and Lambert \(2004\)](#) as `zero cells(tacc)`, or you can request no zero-cell adjustment:

```
. meta esize n11 n12 n21 n22, zero cells(none)
```

See *Options* in [META] `meta esize`.

Declaring a meta-analysis model

Before you proceed with performing meta-analysis, we want you to think about the model underlying your meta-analysis. This decision is important because the selected meta-analysis model will determine the availability of some of the meta-analysis methods and, more importantly, how you interpret the obtained results; see *Comparison between the models and interpretation of the results* in [META] **Intro**. Also, most likely, you will want to use the chosen model during your entire meta-analysis session. Thus, we made the choices for the meta-analysis model and, consequently, the meta-analysis estimation method be part of the initial declaration step. But fear not! If desired, you can easily switch to a different meta-analysis model or method for the rest of your meta-analysis session or reset it temporarily for a particular analysis; see *Modifying default meta settings*.

We discuss the available models and the differences between them in detail in *Meta-analysis models* in [META] **Intro**.

Briefly, there are three models to choose from: a common-effect, fixed-effects, or random-effects model. They can be requested by specifying options `common`, `fixed`, or `random`. If you omit all of these options, the random-effects model will be assumed.

A common-effect model makes a strong assumption about the underlying true effect size being the same across (common to) all studies. When this assumption is true, this model is a reasonable choice. Most likely, you will want to verify the plausibility of this assumption for your data. So a model that allows the study effect sizes to be different may be a better choice during the initial analysis.

A fixed-effects model allows the effect sizes to be different across studies and assumes that they are fixed. You may ask: What does “fixed” mean? Different disciplines may have different definitions of a fixed effect. In the context of meta-analysis, you can think of fixed effects as effects of particular interest. In other words, your research questions and final inference are focused only on the specific studies that were selected in the meta-analysis.

Conversely, a random-effects model assumes that the study effect sizes are random, meaning that they represent a random sample from a larger population of similar studies. The results obtained from a random-effects model can be extended to the entire population of similar studies and not just the ones that were selected in the meta-analysis. The meta-analysis literature recommends to start with a random-effects model, which is also Stata’s default for most `meta` commands.

So, which model should you choose? Our recommendation is to start with a random-effects model and explore the heterogeneity, publication bias, and other aspects of your meta-analysis data. If you are interested only in the inference about the particular studies in your data, a fixed-effects model may be a reasonable alternative. We suggest that you avoid using, or at least starting with, a common-effect model unless you verified that the underlying assumption of the common study effects is plausible for your data.

As we described in *Comparison between the models and interpretation of their results* in [META] **Intro**, a fixed-effects model and a common-effect model produce the same results in a meta-analysis. Although the final estimates are the same, their interpretation is different! In a common-effect model, the estimate of the overall effect size is an estimate of the true common effect size, whereas in a fixed-effects model, it is an estimate of the average of true, different study-specific effect sizes. Thus, the `meta` suite provides the two options `common` and `fixed` to emphasize the conceptual differences between the two models. Additionally, when you assume a common-effect model, you essentially imply that certain issues such as study heterogeneity are of no concern in your data. Therefore, when you specify the `common` option, certain commands such as `meta regression` will not be available. This is again our way of reminding you of the underlying assumption of a common-effect model. For other `meta` commands, specifying `common` versus `fixed` will merely change the reported title from, say, “Common-effect meta-analysis” to “Fixed-effects meta-analysis”. Nevertheless, the title change is important because it encourages proper interpretation of the results.

Declaring a meta-analysis estimation method

Depending on a chosen meta-analysis model and effect size, there are a number of methods available to estimate the overall effect size. For a common-effect model and a fixed-effects model, the inverse-variance method, `common(invvariance)` and `fixed(invvariance)`, is used with generic effect sizes, which are declared by `meta set`, and with effect sizes for continuous data, which are declared by `meta esize`. With effect sizes for binary data (except Peto's log odds-ratio), which are also declared by `meta esize`, the Mantel–Haenszel method, `common(mhaenszel)` or `fixed(mhaenszel)`, is also available.

For a random-effects model, there are several different methods to estimate the between-study variance, which contributes to the weights used to estimate the overall effect size. The default method is REML, `random(reml)`, but other methods such as ML, `random(ml)`, and DerSimonian–Laird, `random(dlaird)`, are also available. See *Syntax* in [META] `meta set` for a full list.

When you specify `random`, the REML method is assumed. When you specify `common` or `fixed`, the inverse-variance method is assumed for all effect sizes except log odds-ratios, log risk-ratios, and risk differences, as specified with `meta esize`. For these effect sizes, the Mantel–Haenszel method is the default method.

See *Meta-analysis estimation methods* in [META] **Intro** for detailed descriptions of the methods.

Default meta-analysis model and method

During declaration, `meta set` and `meta esize` assume a random-effects model unless you specify one of options `fixed` or `common`. It also assumes the REML estimation method unless you specify some other method in option `random()`; see *Declaring a meta-analysis estimation method*.

The declared model will be used by all `meta` commands except `meta funnelplot` and `meta labbepplot`, which, for historical reasons, assume a common-effect model with the inverse-variance estimation method. But you can change the assumed model and method by specifying the corresponding options such as `random(dlaird)` with a `meta` command.

Also see *Modifying default meta settings* for details.

Declaring a confidence level for meta-analysis

By default, `meta set` and `meta esize` assume the 95% confidence level (or as set by `set level`) for the entire meta-analysis analysis. You can change this by specifying the `level()` option with these commands. You can also modify the confidence level after the declaration as we describe in *Modifying default meta settings*.

Declaring display settings for meta-analysis

`meta set` and `meta esize` also provide options to control the output of `meta` commands.

The `studylabel(varname)` option specifies a string variable that will be used by `meta` commands such as `meta summarize` and `meta forestplot` to label the studies in the output. By default, the generic labels—Study 1, Study 2, and so on—will be used.

The `eslabel(string)` option specifies a string that will be used by `meta` commands such as `meta summarize` and `meta forestplot` to label effect sizes in the output. The default label with `meta set` is Effect Size. The default label with `meta esize` is specific to the chosen effect size. For instance, it is Log Odds-Ratio for log odds-ratios.

By default, all `meta` commands display a short summary about the declared meta settings such as the variables containing effect sizes and their standard errors. After the declaration, the `meta` commands do not require you to specify the effect-size and standard-error variables again. They simply use the corresponding system variables (see [System variables](#)) created during declaration. The reported summary reminds you that those variables are part of your meta-analysis. You can suppress this summary from all `meta` commands by specifying the `nometashow` option with `meta set` or `meta esize`. You can also suppress this summary for a particular `meta` command by specifying the option with that command; see [Modifying default meta settings](#).

Modifying default meta settings

You can modify the default meta settings both during and after the declaration. Some of the settings may even be modified (temporarily) for a particular `meta` command.

You can modify the default settings during the declaration by simply specifying the corresponding options with `meta set` or `meta esize`. For example, when we type

```
. meta set ...
```

a random-effects model with the REML estimation method is assumed. We can specify another estimation method, for example, ML, by using `random(ml)`:

```
. meta set ... , random(ml)
```

Or we can specify a different meta-analysis model, for example, a fixed-effects model:

```
. meta set ... , fixed
```

After the declaration, you can use `meta update` to modify the current settings. For example, we can switch to a common-effect model for the rest of our meta-analysis by typing

```
. meta update, common
```

Now all subsequent `meta` commands will automatically assume a common-effect model.

In the above examples, we used `meta set`, but you can use the same specifications with `meta esize`. We also demonstrated only a few options, but the same principles apply to the other options supported by `meta set` and `meta esize`.

For options `random()`, `common()` (and `common`), `fixed()` (and `fixed`), `level()`, and `nometashow`, we can also modify the current setting temporarily while running a particular `meta` command. For example, suppose that we want to obtain the results assuming a 90% confidence level with `meta summarize`. We can type

```
. meta summarize, level(90)
```

If we wanted all relevant `meta` commands to use the 90% confidence level, we would have typed

```
. meta update, level(90)
```

Meta-analysis information

When you use `meta set` or `meta esize`, they record information about your study, effect sizes and their precision, and meta-analysis model and meta-analysis estimation method, among other things. This information will be used by subsequent `meta` commands. The summary information is mostly the same between the two commands, but `meta esize` records several additional settings.

Let's get familiar with the meta setting information by looking at examples.

Meta settings with meta set

Consider a fictional dataset, `metaset.dta`, containing generic effect sizes and their standard errors stored in the corresponding variables `es` and `se`.

```
. use https://www.stata-press.com/data/r16/metaset
(Generic effect sizes; fictional data)

. describe es se
```

variable name	storage type	display format	value label	variable label
<code>es</code>	double	%10.0g		Effect sizes
<code>se</code>	double	%10.0g		Std. Err. for effect sizes

At the minimum, with `meta set`, we must specify the variables containing effect sizes and their standard errors. (For other uses of `meta set`, see [Remarks and examples](#) in [\[META\] meta set](#).)

```
. meta set es se
Meta-analysis setting information
Study information
  No. of studies: 10
  Study label: Generic
  Study size: N/A
Effect size
  Type: Generic
  Label: Effect Size
  Variable: es
Precision
  Std. Err.: se
  CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML
```

The summary is divided into four categories: information about the study, the specified effect sizes, their precision, and meta-analysis model and method. Below, we describe in detail each output category.

```
Study information
  No. of studies: 10
  Study label: Generic
  Study size: N/A
```

The study information consists of the number of studies (10 in our example), a study label (`Generic`), and a study size (`N/A`). If the `studylabel(varname)` option is specified, the `Study label:` will contain the name of the specified variable. Otherwise, a generic study label—`Study 1`, `Study 2`, and so on—will be used in the output of `meta` commands. If the `studysize(varname)` option is specified with `meta set`, the `Study size:` will contain the name of the specified variable.

```
Effect size
  Type: Generic
  Label: Effect Size
  Variable: es
```

The effect-size information consists of the type of the effect size, its label, and the variable containing study-specific effect sizes. The effect-size `Type:` is always `Generic` with `meta set`. The effect-size `Label:` is either a generic `Effect Size` or as specified in the `eslabel(string)` option. This label

will be used to label the effect sizes in the output of all `meta` commands. The effect-size `Variable:` displays the name of the declared variable containing effect sizes. After the declaration, both commands store study-specific effect sizes in the system variable `_meta_es` (see *System variables*). `meta set` simply copies them from the declared effect-size variable. Thus, `Variable:` will contain the name of the *esvar* variable, `es` in our example, with `meta set`.

```
Precision
Std. Err.: se
          CI: [_meta_cil, _meta_ciu]
CI level: 95%
```

The precision information consists of variables containing effect-size standard errors, confidence intervals, and the declared confidence level. As with the effect sizes, `meta set` uses the standard errors specified in the *sevar* variable (variable `se` here). The corresponding confidence intervals are computed using the effect sizes and their standard errors and stored in the system variables `_meta_cil` and `_meta_ciu`. With `meta set`, you can specify confidence intervals instead of the standard errors, in which case the standard errors will be computed from the effect sizes and confidence intervals and stored in `_meta_se`, in which case `Std. Err.:` will contain `_meta_se`; see *Syntax* in **[META] meta set**. `CI:` always contains `_meta_cil` and `_meta_ciu`. The specified CI variables will be reported in `User CI:` with their corresponding confidence level reported in `User CI level:`, which is controlled by the `civarlevel()` option. The declared CI variables and the system CI variables will be the same only when `civarlevel()` is the same as `level()`, and the system variables are the ones that are used in the meta-analysis analysis.

`CI level:` reports the confidence level, controlled by the `level()` option, that will be used by all `meta` commands when computing confidence intervals for various meta-analyses such as the CIs of the overall effect size, regression coefficients, and so on. The default confidence level is 95% or as set by `set level`.

```
Model and method
      Model: Random-effects
      Method: REML
```

As we pointed out in *Declaring a meta-analysis model*, the meta-analysis model and, consequently, the meta-analysis estimation method are important aspects of your meta-analysis. As such, we made them be part of your declaration step too. By default, a random-effects model with the REML estimation method is assumed for most `meta` commands; see *Default meta-analysis model and method*. You can change the defaults as we describe in *Modifying default meta settings*.

Meta settings with meta esize

Consider `metaescnt.dta`, containing fictional study-specific summary data for continuous outcomes for group 1 and group 2.

```
. use https://www.stata-press.com/data/r16/metaescnt, clear
(Fictional summary data for continuous outcomes)
```

```
. describe n1 m1 sd1 n2 m2 sd2
```

variable name	storage type	display format	value label	variable label
n1	byte	%9.0g		Study sizes of group 1
m1	float	%9.0g		Means of group 1
sd1	float	%9.0g		Std. Dev. of group 1
n2	byte	%9.0g		Study sizes of group 2
m2	float	%9.0g		Means of group 2
sd2	float	%9.0g		Std. Dev. of group 2

With `meta esize`, we must specify the summary data to compute an effect size. Let's focus on the studies comparing the mean differences between the two groups. Our summary data include the numbers of observations and the estimates of means and standard deviations for each group. We specify the variables containing these summaries following the command name.

```
. meta esize n1 m1 sd1 n2 m2 sd2
```

```
Meta-analysis setting information
```

```
Study information
```

```
  No. of studies: 10
  Study label: Generic
  Study size: _meta_studysize
  Summary data: n1 m1 sd1 n2 m2 sd2
```

```
Effect size
```

```
  Type: hedgesg
  Label: Hedges's g
  Variable: _meta_es
```

```
Bias correction: Approximate
```

```
Precision
```

```
  Std. Err.: _meta_se
  Std. Err. adj.: None
  CI: [_meta_cil, _meta_ciu]
  CI level: 95%
```

```
Model and method
```

```
  Model: Random-effects
  Method: REML
```

The meta setting information from `meta esize` is almost the same as the one produced by `meta set`, which we described in [Meta settings with meta set](#), but has several additional settings. The summary-data variables are listed under `Summary data:`. As we mentioned earlier, `meta esize` computes the effect sizes and their standard errors from the specified summary data, so `effect-size Variable:` and `Std. Err.:` contain the names of the corresponding system variables, `_meta_es` and `_meta_se`. The summary data also include the information about the study size, so `Study size:` displays the name of the system variable, `_meta_studysize`, that contains study size, which is equal to the sum of `n1` and `n2` in our example.

By default, `meta esize` computes the Hedges's *g* effect size for the two-group mean comparison. You can specify the `esize(esspec)` option to select a different effect size. For the Hedges's *g* effect size, there are two methods to compute the underlying bias-correction term: approximate or exact. For consistency with the meta-analysis literature, `meta esize`, by default, uses an approximation, as indicated in `Bias correction:` under `Effect size`. But you can change this by specifying the `exact` option within `esize()`.

Another additional setting describes the type of adjustment applied when computing the standard errors of the effect sizes; see `Std. Err. adj.:` under `Precision`. This adjustment is applicable only with the Hedges's g or Cohen's d effect size. No adjustment is made by default, but you can use the `holkinse` option within `esize()` to specify the adjustment of Hedges and Olkin (1985). For the mean-difference effect size, you can request the adjustment for unequal group variances by specifying `esize()`'s option `unequal`.

Finally, for log odds-ratios or log risk-ratios, `meta esize` additionally reports the type of adjustment made to the zero cells of contingency tables, which represent the summary data for binary outcomes. For these effect sizes, the type of adjustment will be listed in `Zero-cells adj.:` under `Effect size` (not applicable in our example). By default, 0.5 is added to each zero cell, but you can specify the `zerocells()` option with `meta esize` to apply a different adjustment or none.

System variables

`meta set` and `meta esize` store information about the meta-analysis settings in data characteristics ([P] `char`) and system variables.

`meta` system variables are the variables that begin with `_meta_`. There are four main variables that are stored by the two commands.

`_meta_es` stores study-specific effect sizes.

`_meta_se` stores the standard errors of study-specific effect sizes.

`_meta_cil` and `_meta_ciu` store the lower and upper limits of the CIs for study-specific effect sizes. These variables correspond to the confidence level declared for the meta-analysis, the value of which is stored in the data characteristic `_meta_level`.

Other system variables include integer study identifiers stored in `_meta_id`, study labels stored in a string variable `_meta_studylabel`, and study sizes stored in `_meta_studysize`. `_meta_studysize` is always stored with `meta esize`. With `meta set`, it is stored only when the variable containing study sizes is specified in the `studysize()` option.

Also see *Stored results* in [META] `meta set` and *Stored results* in [META] `meta esize`.

Examples of data declaration for meta-analysis

In this section, we demonstrate how to prepare data for meta-analysis in Stata for several case studies.

Declaring precomputed effect sizes using `meta set`

We will demonstrate how to use `meta set` to declare generic effect sizes.

► Example 1: Precomputed log hazard-ratios using `meta set`

We demonstrate how to declare the meta-analysis data from Steurer et al. (2006), who studied the effect of purine analogues for the treatment of chronic lymphocytic leukemia. Variables `loghr` and `seloghr` contain the log hazard-ratios and their standard errors.

```

. use https://www.stata-press.com/data/r16/leukemia2, clear
( Single-agent purine analogue treatment for leukemia )
. describe
Contains data from https://www.stata-press.com/data/r16/leukemia2.dta
  obs:           4                      Single-agent purine analogue
                                         treatment for leukemia
  vars:          6                      25 Apr 2019 12:09
                                         ( _dta has notes )

```

variable name	storage type	display format	value label	variable label
author	str14	%14s		* Author
year	int	%8.0g		Publication year
ntreat	int	%8.0g		Treatment-group sample size
ncontrol	int	%8.0g		Control-group sample size
loghr	float	%9.0g		Log hazard-ratio
seloghr	float	%9.0g		Standard error for loghr
* indicated variables have notes				

Sorted by:

We use the `meta set` command to declare the effect sizes (log hazard-ratios) and their standard errors.

```

. meta set loghr seloghr
Meta-analysis setting information
Study information
  No. of studies: 4
  Study label: Generic
  Study size: N/A
Effect size
  Type: Generic
  Label: Effect Size
  Variable: loghr
Precision
  Std. Err.: seloghr
  CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML

```

`meta set` reports that there are 4 studies in this dataset. The type of effect size is `Generic` because we used the precalculated effect size. The default label `Effect Size` will be used in the output. The command also reports the variables that were used to declare the effect sizes, `loghr`, and their standard errors, `seloghr`. The other settings are as we described in [Meta settings with meta set](#).

As we described in [System variables](#), meta set created several system variables that will be used by other meta commands in the computations:

```
. describe _meta*
      storage   display   value
variable name  type     format   label    variable label
-----
 _meta_id      byte     %9.0g   Study ID
 _meta_studylabel~1 str7     %9s     Study label
 _meta_es      float    %9.0g   Generic ES
 _meta_se      float    %9.0g   Std. Err. for ES
 _meta_cil     double  %10.0g  95% lower CI limit for ES
 _meta_ciu     double  %10.0g  95% upper CI limit for ES

. list _meta*

```

	_meta_id	_meta~el	_meta_es	_meta_se	_meta_cil	_meta_ciu
1.	1	Study 1	-.592	.345	-1.2681876	.08418756
2.	2	Study 2	-.0791	.0787	-.23334916	.07514916
3.	3	Study 3	-.237	.144	-.51923481	.0452348
4.	4	Study 4	.163	.312	-.44850877	.77450878

`_meta_id` contains integers identifying the studies, and `_meta_studylabel` contains the study labels. `_meta_es` and `_meta_se` contain log hazard-ratios and their standard errors, and `_meta_cil` and `_meta_ciu` contain the corresponding lower and upper bounds of the 95% CIs for log hazard-ratios.

We did not specify the `studylabel()` option in this example, so generic labels will be used in the output of other meta commands such as [meta summarize](#):

```
. meta summarize
      Effect-size label: Effect Size
      Effect size: loghr
      Std. Err.: seloghr

Meta-analysis summary
Random-effects model
Method: REML
Number of studies = 4
Heterogeneity:
      tau2 = 0.0000
      I2 (%) = 0.00
      H2 = 1.00

```

Study	Effect Size	[95% Conf. Interval]		% Weight
Study 1	-0.592	-1.268	0.084	3.68
Study 2	-0.079	-0.233	0.075	70.70
Study 3	-0.237	-0.519	0.045	21.12
Study 4	0.163	-0.449	0.775	4.50
theta	-0.120	-0.250	0.009	

```
Test of theta = 0: z = -1.82          Prob > |z| = 0.0688
Test of homogeneity: Q = chi2(3) = 3.62      Prob > Q = 0.3049

```

Generic labels Study 1, Study 2, Study 3, and Study 4 are used to label the studies. Also, the generic label Effect Size is used to label the log hazard-ratios. See [\[META\] meta summarize](#) for details about `meta summarize`.

We can provide more descriptive labels for the studies and the effect sizes by specifying options `studylabel()` and `eslabel()`.

```

. generate studylbl = author + " (" + string(year) + ")"
. meta set loghr seloghr, studylabel(studylbl) eslabel("Ln(HR)")
Meta-analysis setting information
Study information
  No. of studies: 4
  Study label: studylbl
  Study size: N/A
  Effect size
    Type: Generic
    Label: Ln(HR)
    Variable: loghr
  Precision
  Std. Err.: seloghr
    CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML

```

We created a new variable, `studylbl`, that combines the author and year information of the published studies to use as our study labels. `meta set` reported that `studylbl` will be used to label the studies and `Ln(HR)` to label the effect sizes.

If we now rerun `meta summarize` (suppressing the table header), we see the new labels in the output.

```

. meta summarize, noheader
Effect-size label: Ln(HR)
Effect size: loghr
Std. Err.: seloghr
Study label: studylbl

```

Study	Ln(HR)	[95% Conf. Interval]		% Weight
Johnson et al. (1996)	-0.592	-1.268	0.084	3.68
Leporrier (2001)	-0.079	-0.233	0.075	70.70
Rai (2000)	-0.237	-0.519	0.045	21.12
Robak (2000)	0.163	-0.449	0.775	4.50
theta	-0.120	-0.250	0.009	

```

Test of theta = 0: z = -1.82                      Prob > |z| = 0.0688
Test of homogeneity: Q = chi2(3) = 3.62          Prob > Q = 0.3049

```

After the original declaration, we can use `meta update` to update the meta settings instead of repeating `meta set`; see [example 4](#).

◀

Also see [Remarks and examples](#) in `[META] meta set` for more examples of using `meta set`.

Computing and declaring effect sizes using `meta esize`

We demonstrate how to use `meta esize` to compute and declare effect sizes for continuous and binary outcomes.

► Example 2: Mean differences for continuous data using meta esize

Consider the study of [Gibson et al. \(2002\)](#), who compared the performance of asthma-management programs for adults with asthma.

The `asthma` dataset contains the following summary-data variables:

```
use https://www.stata-press.com/data/r16/asthma, clear
(Education and medical review for asthma patients)
```

```
. describe ni meani sdi nc meanc sdc
```

variable name	storage type	display format	value label	variable label
<code>ni</code>	int	%9.0g		Intervention-group sample size
<code>meani</code>	double	%9.0g		Average days off work/school for intervention group
<code>sdi</code>	double	%9.0g		Std. Dev. of days off work/school for intervention group
<code>nc</code>	int	%9.0g		Control-group sample size
<code>meanc</code>	double	%9.0g		Average days off work/school for control group
<code>sdc</code>	double	%9.0g		Std. Dev. of days off work/school for control group

Variables `ni`, `meani`, and `sdi` record the study-specific sample sizes, mean numbers of days off work/school, and standard deviations in the intervention group, and variables `nc`, `meanc`, and `sdc` record those items in the control group.

To illustrate, we will compute and declare a couple of effect sizes using `meta esize`. We will start with the default effect size—Hedges’s g standardized mean. We use `meta esize` to compute this effect size for each study from the summary variables and declare them for further meta-analysis.

```
. meta esize ni meani sdi nc meanc sdc
(2 missing values generated)

Meta-analysis setting information

Study information
  No. of studies: 13
  Study label: Generic
  Study size: _meta_studysize
  Summary data: ni meani sdi nc meanc sdc

Effect size
  Type: hedgesg
  Label: Hedges's g
  Variable: _meta_es

Bias correction: Approximate

Precision
  Std. Err.: _meta_se
  Std. Err. adj.: None
  CI: [_meta_cil, _meta_ciu]
  CI level: 95%

Model and method
  Model: Random-effects
  Method: REML
```

There are missing values in the summary variables, so some of the generated system variables will also contain missing values as reported by the note.

`meta esize` reports that the computed effect size is Hedges’s g . See [Meta settings with meta esize](#) for the explanation of other settings.

The `above command` is equivalent to

```
. meta esize ni meani sdi nc meanc sdc, esize(hedgesg)
(output omitted)
```

With this effect size, we can specify that the adjustment of [Hedges and Olkin \(1985\)](#) be applied to the standard errors.

```
. meta esize ni meani sdi nc meanc sdc, esize(hedgesg, holkinse)
(2 missing values generated)
```

```
Meta-analysis setting information
Study information
  No. of studies: 13
  Study label: Generic
  Study size: _meta_studysize
  Summary data: ni meani sdi nc meanc sdc
  Effect size
    Type: hedgesg
    Label: Hedges's g
    Variable: _meta_es
  Bias correction: Approximate
  Precision
    Std. Err.: _meta_se
  Std. Err. adj.: Hedges-Olkin
    CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML
```

`meta esize` updates the adjustment in `Std. Err. adj.:` under `Precision` to `Hedges-Olkin`.

Because all studies measured our outcome of interest on the same scale (number of days off work or school), we may consider the raw (unstandardized) mean difference as our effect size. We can compute it by specifying the `esize(mdif)` option.

```
. meta esize ni meani sdi nc meanc sdc, esize(mdif)
(2 missing values generated)
```

```
Meta-analysis setting information
Study information
  No. of studies: 13
  Study label: Generic
  Study size: _meta_studysize
  Summary data: ni meani sdi nc meanc sdc
  Effect size
    Type: mdif
    Label: Mean Diff.
    Variable: _meta_es
  Precision
    Std. Err.: _meta_se
  Std. Err. adj.: None
    CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML
```

The information about the type of the effect size and its label is updated to correspond to the mean differences.

As with `meta set`, we could have used `meta update` to update the meta settings after the initial declaration instead of using `meta esize`; see [example 4](#).

◀

▶ Example 3: Log odds-ratios and log risk-ratios for binary data

Let's revisit the declaration we used in [example 1](#) in [\[META\] meta](#) for the `bcg` dataset from the BCG vaccine study ([Colditz et al. 1994](#)). The summary data (contingency tables) are recorded in the following variables:

```
use https://www.stata-press.com/data/r16/bcg, clear
(Efficacy of BCG vaccine against tuberculosis)
```

```
. describe npost nnegt npsc nnegc
```

variable name	storage type	display format	value label	variable label
<code>npost</code>	int	%9.0g		Number of TB positive cases in treated group
<code>nnegt</code>	long	%9.0g		Number of TB negative cases in treated group
<code>npsc</code>	int	%9.0g		Number of TB positive cases in control group
<code>nnegc</code>	long	%9.0g		Number of TB negative cases in control group

The summary variables represent the cells of the 2×2 tables for each study.

As with continuous data, we specify the summary variables for binary data following `meta esize`:

```
. meta esize npost nnegt npsc nnegc
Meta-analysis setting information
Study information
  No. of studies: 13
  Study label: Generic
  Study size: _meta_studysize
  Summary data: npost nnegt npsc nnegc
Effect size
  Type: lnoratio
  Label: Log Odds-Ratio
  Variable: _meta_es
Zero-cells adj.: None; no zero cells
Precision
  Std. Err.: _meta_se
  CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML
```

The computed default effect sizes are log odds-ratios, whereas the effect of interest in this study is the risk ratio or, equivalently, the log risk-ratio.

To compute log risk-ratios, we specify `esize(lnrratio)`. We also specify the variable `studylbl` containing study labels in the `studylabel()` option.

```
. meta esize npost nnegt npsc nnegc, esize(lnrratio) studylabel(studylbl)
Meta-analysis setting information
Study information
  No. of studies: 13
  Study label: studylbl
  Study size: _meta_studysize
  Summary data: npost nnegt npsc nnegc
  Effect size
    Type: lnrratio
    Label: Log Risk-Ratio
    Variable: _meta_es
  Zero-cells adj.: None; no zero cells
  Precision
    Std. Err.: _meta_se
    CI: [_meta_cil, _meta_ciu]
    CI level: 95%
  Model and method
    Model: Random-effects
    Method: REML
```

Notice that there are no zero cells in our data, so there is no zero-cells adjustment (see `Zero-cells adj.:` under `Effect size`).

◀

Also see [example 4](#) for how to update the above meta settings without having to respecify the summary variables.

Displaying and updating meta settings

We show examples of how to display the current meta settings by using `meta query` and update them by using `meta update`.

► Example 4: Commands meta query and meta update

Recall [example 3](#). Let's reload the dataset and use `meta query` to check whether the dataset is meta set.

```
. use https://www.stata-press.com/data/r16/bcg, clear
(Efficacy of BCG vaccine against tuberculosis)
. meta query
(data not meta set; use meta set or meta esize to declare as meta data)
```

The data are not meta set.

Let's again use `meta esize` to declare the data (quietly) and use `meta query` to display the current settings.

```
. quietly meta esize npost nnegt npsc nnegc
. meta query
-> meta esize npost nnegt npsc nnegc
Meta-analysis setting information from meta esize
Study information
  No. of studies: 13
  Study label: Generic
  Study size: _meta_studysize
  Summary data: npost nnegt npsc nnegc
  Effect size
    Type: lnoratio
    Label: Log Odds-Ratio
    Variable: _meta_es
  Zero-cells adj.: None; no zero cells
  Precision
  Std. Err.: _meta_se
    CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML
```

In [example 3](#), we redeclared the data to use the log risk-ratios as effect sizes. After the initial declaration, it is more convenient to use `meta update` to update the meta settings because we do not need to respecify the summary variables with `meta update`.

```
. meta update, esize(lnrratio) studylabel(study1bl)
-> meta esize npost nnegt npsc nnegc , esize(lnrratio) studylabel(study1bl)
Meta-analysis setting information from meta esize
Study information
  No. of studies: 13
  Study label: study1bl
  Study size: _meta_studysize
  Summary data: npost nnegt npsc nnegc
  Effect size
    Type: lnrratio
    Label: Log Risk-Ratio
    Variable: _meta_es
  Zero-cells adj.: None; no zero cells
  Precision
  Std. Err.: _meta_se
    CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML
```

If your summary variables change, however, you must specify them with `meta esize`.

You can use `meta update` after either `meta esize` or `meta set`. `meta update` will respect the options of `meta esize` and `meta set`.

For example, recall the meta set declaration from [example 1](#):

```
. use https://www.stata-press.com/data/r16/leukemia2, clear
(Single-agent purine analogue treatment for leukemia)
. quietly meta set loghr seloghr
```

Let's update the meta settings to include the variable containing study sizes.

```
. generate ssize = ntreat + ncontrol
. meta update, studysize(ssize)
-> random(reml) studysize(ssize)
```

Meta-analysis setting information from meta set

```
Study information
  No. of studies: 4
  Study label: Generic
  Study size: ssize
  Effect size
    Type: Generic
    Label: Effect Size
    Variable: loghr
  Precision
  Std. Err.: seloghr
    CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML
```

The `studysize()` option is supported only with `meta set`. If we tried to specify this option with `meta update` after `meta esize`, we would have received an error message.

◀

References

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Also see

[META] **meta esize** — Compute effect sizes and declare meta-analysis data

[META] **meta set** — Declare meta-analysis data using generic effect sizes

[META] **meta update** — Update, describe, and clear meta-analysis settings

[META] **meta** — Introduction to meta

[META] **Glossary**

[META] **Intro** — Introduction to meta-analysis

Title

meta esize — Compute effect sizes and declare meta-analysis data

Description	Quick start	Menu	Syntax
Options	Remarks and examples	Stored results	Methods and formulas
References	Also see		

Description

`meta esize` computes effect sizes from study summary data and uses the results to declare the data in memory to be `meta` data, informing Stata of key variables and their roles in a meta-analysis. It computes various effect sizes and their respective standard errors for two-group comparisons of continuous and binary outcomes. It then uses the computed effect sizes and standard errors to declare the data in memory to be `meta` data. If you do not have the summary data from individual studies and, instead, you have precalculated effect sizes, you can use `meta set` to declare your meta-analysis data. You must use `meta esize` or `meta set` to perform meta-analysis using the `meta` command; see [\[META\] meta data](#).

If you need to update some of the meta settings after the data declaration, see [\[META\] meta update](#). To display current meta settings, use `meta query`; see [\[META\] meta update](#).

Quick start

Compute Hedges's g standardized mean differences and their standard errors from variables `nt` (sample size in treatment group), `meant` (mean of treatment group), `sdt` (standard deviation in treatment group), and their counterparts in the control group: `nc`, `meanc`, and `sd`

```
meta esize nt meant sdt nc meanc sdc
```

As above, but compute Cohen's d instead of the default Hedges's g , and use the DerSimonian–Laird estimation method instead of the default REML method

```
meta esize nt meant sdt nc meanc sdc, esize(cohend) random(dlaird)
```

Compute log odds-ratios and their standard errors from variables `nst` (number of successes in treatment group), `nft` (number of failures in treatment group), and their respective counterparts in control group: `nsc` and `nfc`

```
meta esize nst nft nsc nfc
```

As above, but compute the log risk-ratios instead of the default log odds-ratios

```
meta esize nst nft nsc nfc, esize(lnrratio)
```

As above, but request a common-effect meta-analysis

```
meta esize nst nft nsc nfc, esize(lnrratio) common
```

Menu

Statistics > Meta-analysis

Syntax

Compute and declare effect sizes for two-group comparison of continuous outcomes

```
meta esize n1 mean1 sd1 n2 mean2 sd2 [if] [in] [, options_continuous options]
```

Compute and declare effect sizes for two-group comparison of binary outcomes

```
meta esize n11 n12 n21 n22 [if] [in] [, options_binary options]
```

Variables *n1*, *mean1*, and *sd1* contain sample sizes, means, and standard deviations from individual studies for group 1 (treatment), and variables *n2*, *mean2*, and *sd2* contain the respective summaries for group 2 (control).

Variables *n11* and *n12* contain numbers of successes and numbers of failures from individual studies for group 1 (treatment), and variables *n21* and *n22* contain the respective numbers for group 2 (control). A single observation defined by variables *n11*, *n12*, *n21*, and *n22* represents a 2×2 table from an individual study. Therefore, variables *n11*, *n12*, *n21*, and *n22* represent a sample of 2×2 tables from all studies. We will thus refer to observations on these variables as 2×2 tables and to values of these variables as cells.

options_continuous Description

Main

esize(*esspecct*) specify effect size for continuous outcome to be used in the meta-analysis

Model

random[(*remethod*)] random-effects meta-analysis; default is **random**(*rem1*)

common common-effect meta-analysis; implies inverse-variance method

fixed fixed-effects meta-analysis; implies inverse-variance method

options_binary Description

Main

esize(*estypebin*) specify effect size for binary outcome to be used in the meta-analysis

Model

random[(*remethod*)] random-effects meta-analysis; default is **random**(*rem1*)

common[(*cefemethod*)] common-effect meta-analysis

fixed[(*cefemethod*)] fixed-effects meta-analysis

Options

zerocells(*zcspec*) adjust for zero cells during computation; default is to add 0.5 to all cells of those 2×2 tables that contain zero cells

<i>options</i>	Description
<u>studylabel</u> (<i>varname</i>)	variable to be used to label studies in all meta-analysis output
<u>eslabel</u> (<i>string</i>)	effect-size label to be used in all meta-analysis output; default is <code>eslabel(Effect Size)</code>
<u>level</u> (#)	confidence level for all subsequent meta-analysis commands
[<u>no</u>] <u>metashow</u>	display or suppress meta settings with other <code>meta</code> commands

The syntax of *esspecnt* is

estypecnt [, *esopts*]

<i>estypecnt</i>	Description
<u>hedgesg</u>	Hedges's <i>g</i> standardized mean difference; the default
<u>cohend</u>	Cohen's <i>d</i> standardized mean difference
<u>glassdelta2</u>	Glass's Δ mean difference standardized by group 2 (control) standard deviation; more common than <code>glassdelta1</code>
<u>glassdelta1</u>	Glass's Δ mean difference standardized by group 1 (treatment) standard deviation
<u>mdiff</u>	(unstandardized) mean difference

<i>estypebin</i>	Description
<u>lnoratio</u>	log odds-ratio; the default
<u>lnrratio</u>	log risk-ratio (also known as log rate-ratio and log relative-risk)
<u>rdiff</u>	risk difference
<u>lnorpeto</u>	Peto's log odds-ratio

<i>remethod</i>	Description
<u>reml</u>	restricted maximum likelihood; the default
<u>mle</u>	maximum likelihood
<u>ebayes</u>	empirical Bayes
<u>dlaird</u>	DerSimonian–Laird
<u>sjonkman</u>	Sidik–Jonkman
<u>hedges</u>	Hedges
<u>hschmidt</u>	Hunter–Schmidt

<i>cefemethod</i>	Description
<u>mhaenszel</u>	Mantel–Haenszel
<u>invvariance</u>	inverse variance
<u>ivariance</u>	synonym for <code>invvariance</code>

Options

Main

`esize(esspec)` specifies the effect size to be used in the meta-analysis. For continuous outcomes, *esspec* is *estypecnt* [, *esopts*]. For binary outcomes, *esspec* is *estypebin*.

For continuous outcomes, *estypecnt* is one of the following: `hedgesg`, `cohend`, `glassdelta2`, `glassdelta1`, or `mdiff`. Below, we describe each type with its specific options, *esopts*.

`hedgesg` [, `exact` `holkinse`] computes the effect size as the Hedges's *g* (1981) standardized mean difference. This is the default for continuous outcomes. For consistency with meta-analysis literature, `hedgesg` uses an approximation to compute *g* rather than the exact computation (see *Methods and formulas*), as provided by `esize`'s option `hedgesg`. You can use the `exact` suboption to match the results from `esize` (see [R] `esize`).

`cohend` [, `holkinse`] computes the effect size as the Cohen's *d* (1969, 1988) standardized mean difference.

`glassdelta2` computes the effect size as the Glass's Δ standardized mean difference, where the standardization uses the standard deviation of the group 2 (control group). `glassdelta2` is more common in practice than `glassdelta1`.

`glassdelta1` computes the effect size as the Glass's Δ standardized mean difference, where the standardization uses the standard deviation of the group 1 (treatment group). `glassdelta2` is more common in practice than `glassdelta1`.

`mdiff` [, `unequal`] computes the effect size as the unstandardized or raw mean difference.

esopts are `exact`, `holkinse`, and `unequal`.

`exact` specifies that the exact computation be used for the bias-correction factor in Hedges's *g* instead of an approximation used by default.

`holkinse` specifies that the standard error of Hedges's *g* and Cohen's *d* be computed as described in Hedges and Olkin (1985). This is another approximation to the standard error of these effect sizes sometimes used in practice.

`unequal` specifies that the computation of the standard error of the mean difference (`esize(mdiff)`) assume unequal group variances.

For binary outcomes, *estypebin* is one of the following: `lnoratio`, `lnrratio`, `rdiff`, or `lnorpeto`.

`lnoratio` specifies that the effect size be the log odds-ratio. This is the default for binary outcomes.

`lnrratio` specifies that the effect size be the log risk-ratio, also known as a log relative-risk and a log risk-rate.

`rdiff` specifies that the effect size be the risk difference.

`lnorpeto` specifies that the effect size be the log odds-ratio as defined by Peto et al. (1977). This effect size is preferable with rare events.

For effect sizes in the log metric such as log odds-ratios, the results by default are displayed in the log metric. You can use *eform_option* to obtain exponentiated results such as odds ratios.

Model

Options `random()`, `common()`, and `fixed()` declare the meta-analysis model globally throughout the entire meta-analysis; see *Declaring a meta-analysis model* in [META] `meta data`. In other words, once you set your meta-analysis model using `meta esize`, all subsequent `meta` commands will assume

that same model. You can update the declared model by using `meta update` or change it temporarily by specifying the corresponding option with the `meta` commands. Options `random()`, `common()`, and `fixed()` may not be combined. If these options are omitted, `random(reml)` is assumed; see *Default meta-analysis model and method* in [META] **meta data**. Also see *Meta-analysis models* in [META] **Intro**.

`random` and `random(remethod)` specify that a random-effects model be assumed for meta-analysis; see *Random-effects model* in [META] **Intro**.

remethod specifies the type of estimator for the between-study variance τ^2 . *remethod* is one of `reml`, `mle`, `ebayes`, `dlaird`, `sjonkman`, `hedges`, or `hschmidt`. `random` is a synonym for `random(reml)`. Below, we provide a short description for each method based on Veroniki et al. (2016). Also see *Declaring a meta-analysis estimation method* in [META] **meta data**.

`reml`, the default, specifies that the REML method (Raudenbush 2009) be used to estimate τ^2 . This method produces an unbiased, nonnegative estimate of the between-study variance and is commonly used in practice. Method `reml` requires iteration.

`mle` specifies that the ML method (Hardy and Thompson 1996) be used to estimate τ^2 . It produces a nonnegative estimate of the between-study variance. With a few studies or small studies, this method may produce biased estimates. With many studies, the ML method is more efficient than the REML method. Method `mle` requires iteration.

`ebayes` specifies that the empirical Bayes estimator (Berkey et al. 1995), also known as the Paule–Mandel estimator (Paule and Mandel 1982), be used to estimate τ^2 . From simulations, this method, in general, tends to be less biased than other random-effects methods, but it is also less efficient than `reml` or `dlaird`. Method `ebayes` produces a nonnegative estimate of τ^2 and requires iteration.

`dlaird` specifies that the DerSimonian–Laird method (DerSimonian and Laird 1986) be used to estimate τ^2 . This method, historically, is one of the most popular estimation methods because it does not make any assumptions about the distribution of random effects and does not require iteration. But it may underestimate the true between-study variance, especially when the variability is large and the number of studies is small. This method may produce a negative value of τ^2 and is thus truncated at zero in that case.

`sjonkman` specifies that the Sidik–Jonkman method (Sidik and Jonkman 2005) be used to estimate τ^2 . This method always produces a nonnegative estimate of the between-study variance and thus does not need truncating at 0, unlike the other noniterative methods. Method `sjonkman` does not require iteration.

`hedges` specifies that the Hedges method (Hedges 1983) be used to estimate τ^2 . When the sampling variances of effect-size estimates can be estimated without bias, this estimator is exactly unbiased (before truncation), but it is not widely used in practice (Veroniki et al. 2016). Method `hedges` does not require iteration.

`hschmidt` specifies that the Hunter–Schmidt method (Schmidt and Hunter 2015) be used to estimate τ^2 . Although this estimator achieves a lower MSE than other methods, except ML, it is known to be negatively biased. Method `hschmidt` does not require iteration.

`common` and `common(cefemethod)` specify that a common-effect model be assumed for meta-analysis; see *Common-effect (“fixed-effect”) model* in [META] **Intro**. Also see the discussion in [META] **meta data** about common-effect versus fixed-effects models.

`common` implies `common(mhaenszel)` for effect sizes `lnoratio`, `lnrratio`, and `rdiff` and `common(invvariance)` for all other effect sizes.

cefemethod is one of *mhaenszel* or *invvariance* (synonym *ivariance*). Below, we provide a short description for each method. Also see [Declaring a meta-analysis estimation method in \[META\] meta data](#).

mhaenszel is available only for binary outcomes. It specifies a meta-analysis using the Mantel–Haenszel method to estimate the overall effect size for binary outcomes. This method is the default for effect sizes *lnratio*, *lnrratio*, and *rdiff* but is not available for effect size *lnorpeto*.

invvariance specifies a meta-analysis using the inverse-variance method to estimate the overall effect size. This method is available for all types of outcomes and effect sizes and is the default for continuous outcomes and for binary outcomes using effect size *lnorpeto*.

ivariance is a synonym for *invvariance*.

fixed and *fixed(cefemethod)* specify that a fixed-effects model be assumed for meta-analysis; see [Fixed-effects model](#) in [META] **Intro**. Also see the [discussion](#) in [META] **meta data** about fixed-effects versus common-effect models.

fixed implies *fixed(mhaenszel)* for effect sizes *lnratio*, *lnrratio*, and *rdiff* and *fixed(invvariance)* for all other effect sizes.

cefemethod is one of *mhaenszel* or *invvariance* (synonym *ivariance*); see descriptions [above](#).

Options

zerocells(zcspec) is for use with binary outcomes when the effect size is either *lnratio* or *lnrratio*. It specifies the adjustment to be used for the cells, the values of variables *n11*, *n12*, *n21*, and *n22*, when there are zero cells. The adjustment is applied during computation—the original data are not modified. The default is *zerocells(0.5, only0)*; it adds 0.5 to all cells of 2×2 tables that contain at least one zero cell. To request no adjustment, specify *zerocells(none)*. More generally, the syntax of *zcspec* is

```
# [ , zcadj ]
```

where *#* is the adjustment value, also known as the continuity-correction value in the meta-analysis literature, and *zcadj* is *only0* or *allif0*.

only0 specifies that *#* be added to all cells of only those 2×2 tables that contain at least one zero cell. That is, during computation, *#* is added to each observation defined by variables *n11*, *n12*, *n21*, and *n22* if that observation contains a value of zero in any of those variables.

allif0 specifies that *#* be added to all cells of all 2×2 tables but only if there is at least one 2×2 table with a zero cell. That is, during computation, *#* is added to all values of variables *n11*, *n12*, *n21*, and *n22* but only if there is a zero value in one of the four variables.

For the effect size *lnratio*, *zcspec* may also be *tacc*, which implements the treatment-arm continuity correction of [Sweeting, Sutton, and Lambert \(2004\)](#). This method estimates the group-specific adjustment values from the data to minimize the bias of the overall odds-ratio estimator in the presence of zero cells. This method is recommended when the groups are unbalanced.

studylabel(varname) specifies a string variable containing labels for the individual studies to be used in all applicable meta-analysis output. The default study labels are *Study 1*, *Study 2*, ..., *Study K*, where *K* is the total number of studies in the meta-analysis.

eslabel(string) specifies that *string* be used as the effect-size label in all relevant meta-analysis output. The default label is *Effect Size*.

level(#) specifies the confidence level, as a percentage, for confidence intervals. It will be used by all subsequent meta-analysis commands when computing confidence intervals. The default is

level(95) or as set by set level; see [R] [level](#). After the declaration, you can specify level() with meta update to update the confidence level to be used throughout the rest of the meta-analysis session. You can also specify level() directly with the meta commands to modify the confidence level, temporarily, during the execution of the command.

metashow and nometashow display or suppress the meta setting information in the output of other meta commands. By default, this information is displayed at the top of their output. You can also specify nometashow with meta update to suppress the meta setting output for the entire meta-analysis session after the declaration.

Remarks and examples

Remarks are presented under the following headings:

Meta-analysis of binary data

Meta-analysis of continuous data

meta esize computes various effect sizes, their standard errors, and CIs for two-group comparisons of continuous and binary outcomes from the summary data provided for each study. It then declares the computed effect-size data as the meta data; see [META] [meta data](#). Different types of effect sizes may be specified in the esize() option. They depend on the data type, so we describe them separately for binary and continuous data below, together with other data-specific options. Also see *Declaring meta-analysis information* in [META] [meta data](#).

Meta-analysis of binary data

Meta-analysis primarily compares two groups. The first group is commonly referred to as the experimental or treatment group. The second group is commonly referred to as the control group.

For a binary outcome, each study typically reports cell counts from the following 2×2 table.

group	success	failure	size
treatment	n_{11}	n_{12}	$n_1 = n_{11} + n_{12}$
control	n_{21}	n_{22}	$n_2 = n_{21} + n_{22}$

The cells of the table are composed of the numbers of “successes” and “failures” within each of the comparison groups. If a subject experiences an event of interest, it is a success; otherwise, it is a failure. Thus, the summary data for binary outcomes include the above 2×2 table for each study.

For binary data, meta esize requires that four variables be specified containing the numbers of successes and failures in the treatment and control groups.

The goal of each study is to compare the probabilities of a success between the two groups. Various effect-size measures can be used for the comparison. For binary outcomes, meta esize provides the following effect sizes: log odds-ratios (including Peto’s log odds-ratios), the default; log risk-ratios; and risk differences. These are specified as lnoratio, lnorpeto, lnrratio, and rdiff in the esize() option.

As described in *Declaring a meta-analysis model* in [META] [meta data](#), you can choose between a random-effects, a fixed-effects, or a common-effect model. You can also choose from a number of estimation methods that are specific to the chosen model. For fixed-effects and common-effect models, in addition to the inverse-variance method, the Mantel–Haenszel method is available (and is the default) with effect sizes lnoratio, lnrratio, and rdiff; see *Declaring a meta-analysis estimation method* in [META] [meta data](#) and *Meta-analysis estimation methods* in [META] [Intro](#) for details.

Zero cell counts are known to create computational difficulties for odds ratios and risk ratios. A common solution is to add a small number, say, 0.5, to all cells of tables containing zero cells. This and other zero-cells adjustments are available in the `zeros` option.

Let's now look at several examples. Consider the following fictional meta-analysis dataset:

```
. use https://www.stata-press.com/data/r16/metaesbin
(Fictional data for binary outcomes)
. describe
Contains data from https://www.stata-press.com/data/r16/metaesbin.dta
  obs:          4          Fictional data for binary
                        outcomes
  vars:          5          23 Apr 2019 12:14
```

variable name	storage type	display format	value label	variable label
study	str7	%9s		Study label
tdead	byte	%9.0g		Deaths in treatment group
tsurv	int	%9.0g		Survivors in treatment group
cdead	byte	%9.0g		Deaths in control group
csurv	int	%9.0g		Survivors in control group

Sorted by:

We will use this dataset to demonstrate how to compute effect sizes, specify different meta-analysis models, and adjust for zero cells with binary outcome meta-analysis data.

► Example 1: A simple case

When working with meta-analysis data that do not have precomputed effect sizes, we can choose to compute effect sizes in a few different ways such as odds ratios and risk ratios. Using the simplest syntactical specification, we can compute the effect sizes, their standard errors, and the corresponding confidence intervals by specifying the number of successes and failures for one group, as well as the successes and failures for the second group, in that order.

```
. meta esize tdead tsurv cdead csurv
Meta-analysis setting information
Study information
  No. of studies: 4
  Study label: Generic
  Study size: _meta_studysize
  Summary data: tdead tsurv cdead csurv
Effect size
  Type: lnoratio
  Label: Log Odds-Ratio
  Variable: _meta_es
Zero-cells adj.: 0.5, only0
Precision
  Std. Err.: _meta_se
  CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML
```

The output indicates that there are 4 studies in the meta-analysis and, by default, a random-effects meta-analysis is to be assumed, where the heterogeneity parameter τ^2 is estimated via the REML

method. The default computed effect size is the log odds-ratio. `meta esize` creates multiple system variables (see *System variables* in [META] **meta data**) that store the effect-size values, their standard errors, and the upper and lower limits of the CIs for the effect sizes.

We can now use, for example, `meta summarize` to list the individual log odds-ratios and the overall log odds-ratio, which is denoted as `theta`.

```
. meta summarize
      Effect-size label:  Log Odds-Ratio
            Effect size:  _meta_es
            Std. Err.:   _meta_se

Meta-analysis summary          Number of studies =      4
Random-effects model          Heterogeneity:
Method: REML                   tau2 = 1.4417
                                I2 (%) = 69.33
                                H2 = 3.26
```

Study	Log Odds-Ratio	[95% Conf. Interval]		% Weight
Study 1	-0.600	-2.079	0.879	27.80
Study 2	0.351	-2.510	3.212	15.65
Study 3	0.778	-0.031	1.586	34.69
Study 4	-2.567	-4.638	-0.495	21.85
theta	-0.403	-1.869	1.063	

```
Test of theta = 0: z = -0.54          Prob > |z| = 0.5899
Test of homogeneity: Q = chi2(3) = 9.93    Prob > Q = 0.0192
```

See [META] **meta summarize** for details.

If we have a variable that stores the labels for each study, perhaps noting the study authors or journal, we can specify it in the `studylabel()` option with `meta esize`. Because we do not have such a variable in this dataset, each study is denoted generically by `Study #`. See [example 4](#) in [META] **meta set** for an example of how to specify the study label and effect-size label. ◀

► Example 2: Specify the effect size

The default is to compute the log odds-ratio for the effect size. To specify another metric, we can use the `esize()` option. For example, below we use the risk ratio (on the log scale) as our effect size by specifying `esize(lnrratio)`:

```
. meta esize tdead tsurv cdead csurv, esize(lnrratio)
Meta-analysis setting information
Study information
  No. of studies: 4
  Study label: Generic
  Study size: _meta_studysize
  Summary data: tdead tsurv cdead csurv
  Effect size
    Type: lnrratio
    Label: Log Risk-Ratio
    Variable: _meta_es
  Zero-cells adj.: 0.5, only0
  Precision
    Std. Err.: _meta_se
    CI: [_meta_cil, _meta_ciu]
    CI level: 95%
Model and method
  Model: Random-effects
  Method: REML
```

◀

► Example 3: Sparse data and adjustments for zero cells

Note that when we list the data, one of the studies has zero deaths.

```
. list tdead tsurv cdead csurv
```

	tdead	tsurv	cdead	csurv
1.	2	116	17	541
2.	0	15	15	682
3.	8	61	37	614
4.	1	421	9	291

By default, `meta esize` adds a constant value of 0.5 (that is, option `zerocells(0.5, only0)` is assumed) to each cell of a study that has a zero cell; see `Zero-cells adj.:` in the output of `meta set` in [example 1](#). We can modify this adjustment by specifying a different constant factor. For example, we might add 0.003 to each zero cell:

```
. meta esize tdead tsurv cdead csurv, zerocells(.003)
Meta-analysis setting information
Study information
  No. of studies: 4
  Study label: Generic
  Study size: _meta_studysize
  Summary data: tdead tsurv cdead csurv
  Effect size
    Type: lnoratio
    Label: Log Odds-Ratio
    Variable: _meta_es
  Zero-cells adj.: .003, only0
  Precision
    Std. Err.: _meta_se
    CI: [_meta_cil, _meta_ciu]
    CI level: 95%
Model and method
  Model: Random-effects
  Method: REML
```

Or we may instead choose a different type of continuity correction, for example, the treatment-arm continuity correction (TACC), which we specify as `zerocells(tacc)`:

```
. meta esize tdead tsurv cdead csurv, zerocells(tacc)
Meta-analysis setting information
Study information
  No. of studies: 4
  Study label: Generic
  Study size: _meta_studysize
  Summary data: tdead tsurv cdead csurv
  Effect size
    Type: lnoratio
    Label: Log Odds-Ratio
    Variable: _meta_es
  Zero-cells adj.: tacc
  Precision
    Std. Err.: _meta_se
    CI: [_meta_cil, _meta_ciu]
    CI level: 95%
Model and method
  Model: Random-effects
  Method: REML
```

Note that this option can be specified only when using the log odds-ratio as the effect size.

◀

► Example 4: Specify the meta-analysis model

In the examples above, we have been using the default random-effects model, but we could specify a different model. For example, we can use a common-effect model using the Mantel–Haenszel method to estimate the overall effect size:

```
. meta esize tdead tsurv cdead csurv, common(mhaenszel)
Meta-analysis setting information
Study information
  No. of studies: 4
  Study label: Generic
  Study size: _meta_studysize
  Summary data: tdead tsurv cdead csurv
  Effect size
    Type: lnoratio
    Label: Log Odds-Ratio
    Variable: _meta_es
  Zero-cells adj.: 0.5, only0
  Precision
    Std. Err.: _meta_se
    CI: [_meta_cil, _meta_ciu]
    CI level: 95%
Model and method
  Model: Common-effect
  Method: Mantel-Haenszel
```

In the above, we could have specified simply `common` because the Mantel–Haenszel method is the default for a common-effect model with log odds-ratios.

◀

Meta-analysis of continuous data

As with binary outcomes, meta-analysis of continuous outcomes also compares two groups. As before, the first group is commonly referred to as the experimental or treatment group, and the second group is commonly referred to as the control group.

For a continuous outcome, each study often reports the numbers of observations, means, and standard deviations in the two groups. Various effect sizes are then computed from these summary data for each study. Thus, to compute effect sizes for continuous data, `meta esize` requires that six variables be specified containing the numbers of observations, means, and standard deviations of the treatment and control groups. The supported effect sizes are the raw mean difference, `esize(mdif)`, and standardized mean differences: Hedges's g , `esize(hedgesg)` (the default); Cohen's d , `esize(cohend)`; and Glass's Δ s, `esize(glassdelta2)` and `esize(glassdelta1)`; see [Methods and formulas](#) for their definitions.

As described in [Declaring a meta-analysis model](#) in [META] [meta data](#), you can choose between a random-effects, a fixed-effects, or a common-effect model. You can also choose from several [estimation methods](#) for random-effects models. Fixed-effects and common-effect models assume the inverse-variance estimation method. Also see [Declaring a meta-analysis estimation method](#) in [META] [meta data](#) and [Meta-analysis estimation methods](#) in [META] [Intro](#) for details.

Let's now demonstrate several usages of `meta esize` for continuous outcomes. Consider the following fictional meta-analysis dataset:

```
. use https://www.stata-press.com/data/r16/metaescnt, clear
(Fictional summary data for continuous outcomes)
. describe
Contains data from https://www.stata-press.com/data/r16/metaescnt.dta
  obs:                10                Fictional summary data for
                                         continuous outcomes
  vars:                6                19 Apr 2019 14:00
```

variable name	storage type	display format	value label	variable label
n1	byte	%9.0g		Study sizes of group 1
m1	float	%9.0g		Means of group 1
sd1	float	%9.0g		Std. Dev. of group 1
n2	byte	%9.0g		Study sizes of group 2
m2	float	%9.0g		Means of group 2
sd2	float	%9.0g		Std. Dev. of group 2

Sorted by:

We will use this dataset to demonstrate different usages of the `meta esize` command with continuous-outcomes meta-analysis data.

▷ Example 5: The assumed model

In the simplest specification, `meta esize` requires that we specify the sample sizes, means, and standard deviations for each group in the meta-analysis.

```
. meta esize n1 m1 sd1 n2 m2 sd2
Meta-analysis setting information
Study information
  No. of studies: 10
  Study label: Generic
  Study size: _meta_studysize
  Summary data: n1 m1 sd1 n2 m2 sd2
  Effect size
    Type: hedgesg
    Label: Hedges's g
    Variable: _meta_es
  Bias correction: Approximate
  Precision
    Std. Err.: _meta_se
  Std. Err. adj.: None
    CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML
```

We see from the output that the Hedges's g standardized mean difference is used for the effect size, and, as for binary outcomes, a random-effects REML model is assumed. See [Meta settings with meta esize](#) in [META] **meta data** for a detailed description of all settings for this dataset.

◀

▷ Example 6: Selecting an effect size

If we do not feel the need to standardize the mean differences, we could instead use the raw mean difference as the effect size by specifying `esize(mdif)`.

```
. meta esize n1 m1 sd1 n2 m2 sd2, esize(mdif)
Meta-analysis setting information
Study information
  No. of studies: 10
  Study label: Generic
  Study size: _meta_studysize
  Summary data: n1 m1 sd1 n2 m2 sd2
  Effect size
    Type: mdif
    Label: Mean Diff.
    Variable: _meta_es
  Precision
    Std. Err.: _meta_se
  Std. Err. adj.: None
    CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML
```

◀

► Example 7: Specifying different meta-analysis models and methods

Rather than using the default REML estimation method, we may want to use a different method, such as the DerSimonian–Laird method. We can specify this method in the `random()` option.

```
. meta esize n1 m1 sd1 n2 m2 sd2, random(dlaird)
Meta-analysis setting information
Study information
  No. of studies: 10
  Study label: Generic
  Study size: _meta_studysize
  Summary data: n1 m1 sd1 n2 m2 sd2
  Effect size
    Type: hedgesg
    Label: Hedges's g
    Variable: _meta_es
  Bias correction: Approximate
  Precision
    Std. Err.: _meta_se
  Std. Err. adj.: None
    CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: DerSimonian-Laird
```

Or, instead of the random-effects model, we may specify a fixed-effects model, which implies the inverse-variance estimation method.

```
. meta esize n1 m1 sd1 n2 m2 sd2, fixed
Meta-analysis setting information
Study information
  No. of studies: 10
  Study label: Generic
  Study size: _meta_studysize
  Summary data: n1 m1 sd1 n2 m2 sd2
  Effect size
    Type: hedgesg
    Label: Hedges's g
    Variable: _meta_es
  Bias correction: Approximate
  Precision
    Std. Err.: _meta_se
  Std. Err. adj.: None
    CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Fixed-effects
  Method: Inverse-variance
```

Stored results

meta esize stores the following characteristics and system variables:

Characteristics

<code>_dta[_meta_marker]</code>	“_meta_ds_1”
<code>_dta[_meta_K]</code>	number of studies in the meta-analysis
<code>_dta[_meta_studylabel]</code>	name of string variable containing study labels or Generic
<code>_dta[_meta_estype]</code>	type of effect size; varies
<code>_dta[_meta_eslabelopt]</code>	eslabel(<i>eslab</i>), if specified
<code>_dta[_meta_eslabel]</code>	effect-size label from eslabel(); default varies
<code>_dta[_meta_eslabeldb]</code>	effect-size label for dialog box
<code>_dta[_meta_esvardb]</code>	_meta_es
<code>_dta[_meta_level]</code>	default confidence level for meta-analysis
<code>_dta[_meta_esizeopt]</code>	esize(<i>estype</i>), if specified
<code>_dta[_meta_esopt_exact]</code>	exact, if esize(, exact) is specified
<code>_dta[_meta_esopt_holkinse]</code>	holkinse, if esize(, holkinse) is specified
<code>_dta[_meta_esopt_unequal]</code>	unequal, if esize(, unequal) is specified
<code>_dta[_meta_modellabel]</code>	meta-analysis model label: Random-effects, Common-effect, or Fixed-effects
<code>_dta[_meta_model]</code>	meta-analysis model: random, common, or fixed
<code>_dta[_meta_methodlabel]</code>	meta-analysis method label; varies by meta-analysis model
<code>_dta[_meta_method]</code>	meta-analysis method; varies by meta-analysis model
<code>_dta[_meta_randomopt]</code>	random(<i>remethod</i>), if specified
<code>_dta[_meta_zcopt]</code>	zerocells(<i>zcspec</i>), if specified
<code>_dta[_meta_zcadj]</code>	type of adjustment for zero cells, if zerocells() specified
<code>_dta[_meta_zcvalue]</code>	value added to cells to adjust for zero cells, if specified
<code>_dta[_meta_show]</code>	empty or nometashow
<code>_dta[_meta_n1var]</code>	name of group 1 sample-size variable; for continuous data
<code>_dta[_meta_mean1var]</code>	name of group 1 mean variable; for continuous data
<code>_dta[_meta_sd1var]</code>	name of group 1 std. dev. variable; for continuous data
<code>_dta[_meta_n2var]</code>	name of group 2 sample-size variable; for continuous data
<code>_dta[_meta_mean2var]</code>	name of group 2 mean variable; for continuous data
<code>_dta[_meta_sd2var]</code>	name of group 2 std. dev. variable; for continuous data
<code>_dta[_meta_n11var]</code>	name of <i>n11</i> variable; for binary data
<code>_dta[_meta_n12var]</code>	name of <i>n12</i> variable; for binary data
<code>_dta[_meta_n21var]</code>	name of <i>n21</i> variable; for binary data
<code>_dta[_meta_n22var]</code>	name of <i>n22</i> variable; for binary data
<code>_dta[_meta_datatype]</code>	data type; continuous or binary
<code>_dta[_meta_datavars]</code>	variables specified with meta esize
<code>_dta[_meta_setcmdline]</code>	meta esize command line
<code>_dta[_meta_ifexp]</code>	<i>if</i> specification
<code>_dta[_meta_inexp]</code>	<i>in</i> specification

System variables

<code>_meta_id</code>	study ID variable
<code>_meta_es</code>	variable containing effect sizes
<code>_meta_se</code>	variable containing effect-size standard errors
<code>_meta_cil</code>	variable containing lower bounds of CIs for effect sizes
<code>_meta_ciu</code>	variable containing upper bounds of CIs for effect sizes
<code>_meta_studylabel</code>	string variable containing study labels
<code>_meta_studysize</code>	variable containing total sample size per study

Methods and formulas

Methods and formulas are presented under the following headings:

- Effect sizes for continuous outcomes*
 - Unstandardized mean difference*
 - Standardized mean difference*
- Effect sizes for binary outcomes*
 - Odds ratio*
 - Risk ratio (rate ratio)*
 - Risk difference*
 - Zero-cells adjustments*
- Confidence intervals for effect sizes*

Effect sizes for continuous outcomes

As we described in *Meta-analysis of continuous data*, meta-analysis often compares two groups: experimental (or treated) group and control group.

When the response (measurement) is continuous, studies typically report a mean and standard deviation for each group. For a given study, the following table denotes the underlying population parameters and the reported summary statistics (data) for each group.

group	population		sample		
	mean	sd	mean	sd	size
treatment	μ_1	σ_1	\bar{x}_1	s_1	n_1
control	μ_2	σ_2	\bar{x}_2	s_2	n_2

The majority of this section is based on [Borenstein \(2009\)](#).

Unstandardized mean difference

Consider the population mean difference

$$\theta = \mu_1 - \mu_2$$

For each study in the meta-analysis, `meta esize` with option `esize(mdif)` estimates θ using the difference in sample means,

$$D = \bar{x}_1 - \bar{x}_2$$

The variance of D , assuming that the two population standard deviations are equal, is estimated by

$$\widehat{\text{Var}}(D) = \left(\frac{1}{n_1} + \frac{1}{n_2} \right) s^2$$

where s is the pooled sample standard deviation

$$s = \sqrt{\frac{(n_1 - 1) s_1^2 + (n_2 - 1) s_2^2}{n_1 + n_2 - 2}}$$

For unequal population standard deviations, use option `esize(mdif, unequal)`; then the variance of D is estimated by

$$\widehat{\text{Var}}(D) = \frac{s_1^2}{n_1} + \frac{s_2^2}{n_2}$$

Unstandardized (raw) mean differences are not comparable across studies if the underlying means are measured on different scales.

Standardized mean difference

The standardized mean difference is

$$\theta = \frac{\mu_1 - \mu_2}{\sigma}$$

Note that θ does not depend on the scale of measurement. The definition of the standardized mean difference implicitly assumes that the population standard deviations, σ_1 and σ_2 , are the same: $\sigma_1 = \sigma_2 = \sigma$.

`meta esize` with option `esize(cohend)` estimates θ using Cohen's d statistic (Cohen 1969, 1988),

$$d = \frac{\bar{x}_1 - \bar{x}_2}{s}$$

The estimated variance of d is given by

$$\widehat{\text{Var}}(d) = \frac{n_1 + n_2}{n_1 n_2} + \frac{d^2}{2(n_1 + n_2)}$$

Hedges (1981) introduced an adjustment to Cohen's d for small samples that accounts for a small upward bias in the absolute value of θ . `meta esize` with option `esize(hedgesg, exact)` computes Hedges's g as

$$g = c(m) \times d$$

where $m = n_1 + n_2 - 2$ is the degrees of freedom used to estimate s and

$$c(m) = \frac{\Gamma\left(\frac{m}{2}\right)}{\sqrt{\frac{m}{2}} \Gamma\left(\frac{m-1}{2}\right)}$$

The adjustment $c(m)$ is less than 1 and approaches 1 as m gets large. The variance estimate of Hedges's g is

$$\widehat{\text{Var}}(g) = c(m)^2 \times \widehat{\text{Var}}(d)$$

Hedges (1981) also introduced an accurate approximation for $c(m)$ that has been traditionally used in meta-analysis. The approximation for $c(m)$ is

$$J = 1 - \frac{3}{4m - 1}$$

`meta esize` with option `esize(hedgesg)` computes Hedges's g using J for $c(m)$; thus,

$$g = J \times d$$

and

$$\widehat{\text{Var}}(g) = J^2 \times \widehat{\text{Var}}(d)$$

`meta esize` with option `esize(glassdelta2)` estimates θ using Glass's Δ (Smith and Glass 1977),

$$\Delta = \frac{\bar{x}_1 - \bar{x}_2}{s_2}$$

Notice that the standard deviation in the denominator is s_2 , the sample standard deviation from the control group, which is considered to be a more reliable estimate of the common variance. The estimated variance of Δ is given by

$$\widehat{\text{Var}}(\Delta) = \frac{n_1 + n_2}{n_1 n_2} + \frac{\Delta^2}{2(n_2 - 1)}$$

In the absence of the control group, such as in observational studies, [Kline \(2013\)](#), among others, suggests providing statistics standardized by the standard deviation of each group. Glass's Δ where standardization is based on the treatment group may be computed via option `esize(glassdelta1)`.

Alternative standard-error estimators are available for Hedges's g and Cohen's d effect sizes.

[Hedges and Olkin \(1985, eq. 8, 80\)](#) provide another commonly used estimator for the variance of Hedges's g .

$$\widehat{\text{Var}}(g) = \frac{n_1 + n_2}{n_1 n_2} + \frac{g^2}{2(m - 1.94)}$$

`meta esize` uses this formula when option `esize(hedgesg, holkinse)` is specified.

The alternative variance estimator of d is given by

$$\widehat{\text{Var}}(d) = \frac{n_1 + n_2}{n_1 n_2} + \frac{d^2}{2(n_1 + n_2 - 2)}$$

This variance estimator may be requested via option `esize(cohend, holkinse)`.

Effect sizes for binary outcomes

As we described in [Meta-analysis of binary data](#), meta-analysis often compares two groups: experimental (or treated) group and control group. When the response (measurement) is binary, each study typically reports cell counts from the following 2×2 table.

group	success	failure	size
treatment	a	b	$n_1 = a + b$
control	c	d	$n_2 = c + d$

Here, for simplicity, we use a different notation for the cell counts (a , b , c , and d) compared with the similar table in [Meta-analysis of binary data](#).

For the treatment group, n_1 is assumed fixed, $a \sim \text{binomial}(n_1, \pi_1)$, and π_1 is the probability of a success. For the control group, n_2 is assumed fixed, $c \sim \text{binomial}(n_2, \pi_2)$, and π_2 is the probability of a success. The goal of each study is to compare the two success probabilities, π_1 and π_2 .

Estimates of the success probabilities are $\hat{\pi}_1 = a/n_1$ for the treatment group and $\hat{\pi}_2 = c/n_2$ for the control group.

Odds ratio

`meta esize` with option `esize(lnoratio)` computes estimates of the log odds-ratios. Odds ratio is the ratio of the odds of a success in the treatment group over the odds of a success in the control group.

$$\text{OR} = \frac{\pi_1 / (1 - \pi_1)}{\pi_2 / (1 - \pi_2)}$$

The odds ratio is estimated by

$$\widehat{\text{OR}} = \frac{ad}{bc}$$

The distribution of $\widehat{\text{OR}}$ is typically skewed, but the natural logarithm of $\widehat{\text{OR}}$, $\ln(\widehat{\text{OR}})$, is asymptotically normally distributed. The estimate of the variance of $\ln(\widehat{\text{OR}})$ is

$$\widehat{\text{Var}} \{ \ln(\widehat{\text{OR}}) \} = \frac{1}{a} + \frac{1}{b} + \frac{1}{c} + \frac{1}{d}$$

`meta esize` with option `esize(lnorpeto)` computes estimates of effect size using Peto's log odds-ratio (Peto et al. 1977; Yusuf et al. 1985). Peto's odds ratio and log odds-ratio are

$$\widehat{\text{OR}}^{\text{Peto}} = \exp \left\{ \frac{a - E(a)}{\text{Var}(a)} \right\}$$

$$\ln \left(\widehat{\text{OR}}^{\text{Peto}} \right) = \frac{a - E(a)}{\text{Var}(a)}$$

where the expectation and variance of a are estimated assuming a hypergeometric distribution:

$$E(a) = \frac{(a+c)n_1}{n}$$

$$\text{Var}(a) = \frac{n_1 n_2 (a+c)(b+d)}{n^2 (n-1)}$$

The variance estimate of $\ln \left(\widehat{\text{OR}}^{\text{Peto}} \right)$ is

$$\widehat{\text{Var}} \left\{ \ln \left(\widehat{\text{OR}}^{\text{Peto}} \right) \right\} = \frac{1}{\text{Var}(a)}$$

See, for instance, Fleiss 1993, Fleiss, Levin, and Paik 2003, and Bradburn et al. (2007) for a discussion of potential bias of Peto's odds ratio and its performance in sparse data.

Risk ratio (rate ratio)

`meta esize` with option `esize(lnrratio)` computes estimates of the log risk-ratios. The risk ratio (RR), also known as the rate ratio or relative risk in the health sciences, is

$$\text{RR} = \frac{\pi_1}{\pi_2}$$

RR is estimated by

$$\widehat{\text{RR}} = \frac{a/n_1}{c/n_2}$$

Similarly to odds ratios, $\widehat{\text{RR}}$ typically has a skewed distribution, but the natural logarithm of $\widehat{\text{RR}}$, $\ln(\widehat{\text{RR}})$, is asymptotically normally distributed. The estimate of the variance of $\ln(\widehat{\text{RR}})$ is

$$\widehat{\text{Var}} \{ \ln(\widehat{\text{RR}}) \} = \frac{1}{a} + \frac{1}{c} - \frac{1}{a+b} - \frac{1}{c+d}$$

Risk difference

`meta esize` with option `esize(rdif)` computes estimates of the risk differences. The risk difference is

$$RD = \pi_1 - \pi_2$$

and is estimated by

$$\widehat{RD} = \frac{a}{n_1} - \frac{c}{n_2}$$

\widehat{RD} is asymptotically normally distributed and is thus typically used without a transformation in meta-analysis.

The estimated variance of \widehat{RD} is

$$\widehat{\text{Var}}(\widehat{RD}) = \frac{ab}{n_1^3} + \frac{cd}{n_2^3}$$

Zero-cells adjustments

The variance estimates of $\ln(\widehat{OR})$ and $\ln(\widehat{RR})$ are not defined if there are any empty (zero count) cells in a 2×2 table. In this case, it is customary to add a small value, often referred to as “continuity correction”, to each cell prior to computing the log odds- or risk-ratio.

By default, `meta esize` adds 0.5 to each cell of 2×2 tables containing empty cells (Gart and Zweifel 1967 and Gart, Pettigrew, and Thomas 1985). Alternatively, you can add a different number or add a number to each cell of all 2×2 tables, as long as there is at least one 2×2 table with zero cells; see option `zerocells()`.

For odds ratios, Sweeting, Sutton, and Lambert (2004) proposed the treatment-arm continuity correction (TACC) method, which estimates the continuity-correction values from the data separately for each group; see `zerocells(tacc)`.

Confidence intervals for effect sizes

For the j th study in a given meta-analysis, let $\widehat{\theta}_j$ be one of the effect-size estimators described above; then the asymptotic $100(1 - \alpha)\%$ confidence interval computed by `meta esize` is

$$\widehat{\theta}_j \pm z_{1-\alpha/2} \sqrt{\widehat{\text{Var}}(\widehat{\theta}_j)}$$

where $z_{1-\alpha/2}$ is the usual critical value from the standard normal distribution.

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Also see

[META] **meta data** — Declare meta-analysis data

[META] **meta set** — Declare meta-analysis data using generic effect sizes

[META] **meta update** — Update, describe, and clear meta-analysis settings

[META] **meta** — Introduction to meta

[META] **Glossary**

[META] **Intro** — Introduction to meta-analysis

[R] **esize** — Effect size based on mean comparison

Title

meta set — Declare meta-analysis data using generic effect sizes

Description	Quick start	Menu	Syntax
Options	Remarks and examples	Stored results	References
Also see			

Description

`meta set` declares the data in memory to be meta data, informing Stata of key variables and their roles in a meta-analysis. It is used with generic (precomputed) effect sizes; see [\[META\] meta esize](#) if you need to compute and declare effect sizes. You must use `meta set` or `meta esize` to perform meta-analysis using the `meta` command; see [\[META\] meta data](#).

If you need to update some of the meta settings after the data declaration, see [\[META\] meta update](#). To display current meta settings, use `meta query`; see [\[META\] meta update](#).

Quick start

Declare generic effect sizes and their standard errors from individual studies stored in variables `es` and `se`

```
meta set es se
```

As above, but request a random-effects meta-analysis where between-study heterogeneity is estimated using the DerSimonian–Laird method instead of the default REML method

```
meta set es se, random(dlaird)
```

Specify a common-effect meta-analysis, study labels stored in a string variable `studylab`, and label effect sizes as `log(HR)` in the output

```
meta set es se, common studylabel(studylab) eslabel("log(HR)")
```

Use 90% confidence level, and suppress the display of meta settings for all subsequent meta-analysis commands

```
meta set es se, level(90) nometashow
```

Specify study sizes stored in variable `ssize`

```
meta set es se, studysize(ssize)
```

Declare generic effect sizes, and compute their standard errors based on the specified 90% CI variables, `cil` and `ciu`

```
meta set es cil ciu, civarlevel(90)
```

Menu

Statistics > Meta-analysis

Syntax

Specify generic effect sizes and their standard errors

```
meta set esvar sevar [if] [in] [, options]
```

Specify generic effect sizes and their confidence intervals

```
meta set esvar cilvar ciuvar [if] [in] [, civarlevel(#) options]
```

esvar specifies a variable containing the effect sizes, *sevar* specifies a variable containing standard errors of the effect sizes, and *cilvar* and *ciuvar* specify variables containing the respective lower and upper bounds of confidence intervals for the effect sizes.

<i>options</i>	Description
Model	
<code>random</code> [<i>(remethod)</i>]	random-effects meta-analysis; default is <code>random(rem1)</code>
<code>common</code>	common-effect meta-analysis; implies inverse-variance method
<code>fixed</code>	fixed-effects meta-analysis; implies inverse-variance method
Options	
<code>studylabel</code> (<i>varname</i>)	variable to be used to label studies in all meta-analysis output
<code>studysize</code> (<i>varname</i>)	total sample size per study
<code>eslabel</code> (<i>string</i>)	effect-size label to be used in all meta-analysis output; default is <code>eslabel(Effect Size)</code>
<code>level</code> (#)	confidence level for all subsequent meta-analysis commands
<code>[no]metashow</code>	display or suppress meta settings with other <code>meta</code> commands

<i>remethod</i>	Description
<code>reml</code>	restricted maximum likelihood; the default
<code>mle</code>	maximum likelihood
<code>ebayes</code>	empirical Bayes
<code>dlaird</code>	DerSimonian–Laird
<code>sjonkman</code>	Sidik–Jonkman
<code>hedges</code>	Hedges
<code>hschmidt</code>	Hunter–Schmidt

Options

Main

`civarlevel`(#) is relevant only when you specify CI variables *cilvar* and *ciuvar* with `meta set`. It specifies the confidence level corresponding to these variables. The default is `civarlevel(95)`. This option affects the computation of the effect-size standard errors stored in the `system variable` `_meta_se`.

Do not confuse `civarlevel()` with `level()`. The former affects the confidence level only for the specified CI variables. The latter specifies the confidence level for the meta-analysis.

Model

Options `random()`, `common`, and `fixed` declare the meta-analysis model globally throughout the entire meta-analysis; see *Declaring a meta-analysis model* in [META] **meta data**. In other words, once you set your meta-analysis model using `meta set`, all subsequent `meta` commands will assume that same model. You can update the declared model by using `meta update` or change it temporarily by specifying the corresponding option with the `meta` commands. Options `random()`, `common`, and `fixed` may not be combined. If these options are omitted, `random(rem1)` is assumed; see *Default meta-analysis model and method* in [META] **meta data**. Also see *Meta-analysis models* in [META] **Intro**.

`random` and `random(remethod)` specify that a random-effects model be assumed for meta-analysis; see *Random-effects model* in [META] **Intro**.

`remethod` specifies the type of estimator for the between-study variance τ^2 . `remethod` is one of `rem1`, `mle`, `ebayes`, `dlaird`, `sjonkman`, `hedges`, or `hschmidt`. `random` is a synonym for `random(rem1)`. Below, we provide a short description for each method based on Veroniki et al. (2016). Also see *Declaring a meta-analysis estimation method* in [META] **meta data**.

`rem1`, the default, specifies that the REML method (Raudenbush 2009) be used to estimate τ^2 . This method produces an unbiased, nonnegative estimate of the between-study variance and is commonly used in practice. Method `rem1` requires iteration.

`mle` specifies that the ML method (Hardy and Thompson 1996) be used to estimate τ^2 . It produces a nonnegative estimate of the between-study variance. With a few studies or small studies, this method may produce biased estimates. With many studies, the ML method is more efficient than the REML method. Method `mle` requires iteration.

`ebayes` specifies that the empirical Bayes estimator (Berkey et al. 1995), also known as the Paule–Mandel estimator (Paule and Mandel 1982), be used to estimate τ^2 . From simulations, this method, in general, tends to be less biased than other random-effects methods, but it is also less efficient than `rem1` or `dlaird`. Method `ebayes` produces a nonnegative estimate of τ^2 and requires iteration.

`dlaird` specifies that the DerSimonian–Laird method (DerSimonian and Laird 1986) be used to estimate τ^2 . This method, historically, is one of the most popular estimation methods because it does not make any assumptions about the distribution of random effects and does not require iteration. But it may underestimate the true between-study variance, especially when the variability is large and the number of studies is small. This method may produce a negative value of τ^2 and is thus truncated at zero in that case.

`sjonkman` specifies that the Sidik–Jonkman method (Sidik and Jonkman 2005) be used to estimate τ^2 . This method always produces a nonnegative estimate of the between-study variance and thus does not need truncating at 0, unlike the other noniterative methods. Method `sjonkman` does not require iteration.

`hedges` specifies that the Hedges method (Hedges 1983) be used to estimate τ^2 . When the sampling variances of effect-size estimates can be estimated without bias, this estimator is exactly unbiased (before truncation), but it is not widely used in practice (Veroniki et al. 2016). Method `hedges` does not require iteration.

`hschmidt` specifies that the Hunter–Schmidt method (Schmidt and Hunter 2015) be used to estimate τ^2 . Although this estimator achieves a lower MSE than other methods, except ML, it is known to be negatively biased. Method `hschmidt` does not require iteration.

`common` specifies that a common-effect model be assumed for meta-analysis; see *Common-effect (“fixed-effect”) model* in [META] **Intro**. It uses the inverse-variance estimation method; see *Meta-analysis estimation methods* in [META] **Intro**. Also see the *discussion* in [META] **meta data** about common-effect versus fixed-effects models.

`fixed` specifies that a fixed-effects model be assumed for meta-analysis; see *Fixed-effects model* in [META] **Intro**. It uses the inverse-variance estimation method; see *Meta-analysis estimation methods* in [META] **Intro**. Also see the *discussion* in [META] **meta data** about fixed-effects versus common-effect models.

Options

`studylabel(varname)` specifies a string variable containing labels for the individual studies to be used in all applicable meta-analysis output. The default study labels are `Study 1`, `Study 2`, `...`, `Study K`, where `K` is the total number of studies in the meta-analysis.

`studysize(varname)` specifies the variable that contains the total sample size for each study. This option is useful for subsequent meta commands that use this information in computations such as `meta funnelplot` using the sample-size metric.

`eslabel(string)` specifies that *string* be used as the effect-size label in all relevant meta-analysis output. The default label is `Effect Size`.

`level(#)` specifies the confidence level, as a percentage, for confidence intervals. It will be used by all subsequent meta-analysis commands when computing confidence intervals. The default is `level(95)` or as set by `set level`; see [R] **level**. After the declaration, you can specify `level()` with `meta update` to update the confidence level to be used throughout the rest of the meta-analysis session. You can also specify `level()` directly with the meta commands to modify the confidence level, temporarily, during the execution of the command.

`metashow` and `nometashow` display or suppress the meta setting information in the output of other meta commands. By default, this information is displayed at the top of their output. You can also specify `nometashow` with `meta update` to suppress the meta setting output for the entire meta-analysis session after the declaration.

Remarks and examples

Remarks are presented under the following headings:

[Overview](#)
[Using meta set](#)

Overview

When you perform meta-analysis, it is common for studies included in the meta-analysis to contain precalculated effect sizes, which we refer to as generic effect sizes, such as mean differences, odds ratios, correlations, and hazard ratios. You can use `meta set` to declare the generic effect sizes. (If you have summary data from which effect sizes can be computed, use [META] **meta esize** instead.)

In addition to effect sizes, their standard errors must be available for meta-analysis. Sometimes, the standard errors are not available, but the confidence intervals (CIs) are. In that case, the standard errors can be computed from the effect-size estimates and CIs. `meta set` supports both cases. You can supply the variables containing effect sizes and their standard errors, or, instead of the standard errors, you can specify the variables containing the CIs.

When you specify the CI variables, you can specify their corresponding confidence level in the `civarlevel()` option. (Do not confuse this option with the `level()` option. The former corresponds to the specified CI variables, whereas the latter specifies the confidence level for the entire meta-analysis.)

Meta-analysis uses effect sizes in a metric that makes them approximately normally distributed such as log odds-ratios instead of odds ratios and log hazard-ratios instead of hazard ratios. The corresponding standard errors or CIs should be provided in the same metric as effect sizes. For example, if you are working with log hazard-ratios, the provided CIs should be for the log hazard-ratios and not the hazard ratios.

See [\[META\] meta data](#) for more details.

Using meta set

Consider the following fictional meta-analysis dataset:

```
. use https://www.stata-press.com/data/r16/metaset
(Generic effect sizes; fictional data)
. describe
Contains data from https://www.stata-press.com/data/r16/metaset.dta
  obs:           10              Generic effect sizes; fictional
                                data
  vars:           9              19 Apr 2019 01:28
```

variable name	storage type	display format	value label	variable label
study	byte	%9.0g		Study ID
es	double	%10.0g		Effect sizes
se	double	%10.0g		Std. Err. for effect sizes
cil	double	%10.0g		95% lower CI limit
ciu	double	%10.0g		95% upper CI limit
cil90	double	%10.0g		90% lower CI limit
ciu90	double	%10.0g		90% upper CI limit
studylab	str23	%23s		Study label
ssize	byte	%9.0g		Study size

Sorted by:

We will use it to describe various usages of the `meta set` command. For examples of declarations of real datasets, see [\[META\] meta data](#).

► Example 1: Declaring effect sizes and standard errors

Meta-analysis datasets often contain precomputed effect sizes and their standard errors. To declare them for meta-analysis using the `meta` commands, we specify the corresponding variables with `meta set`.


```

. meta set es se
Meta-analysis setting information
Study information
  No. of studies: 10
  Study label: Generic
  Study size: N/A
  Effect size
    Type: Generic
    Label: Effect Size
    Variable: es
  Precision
  Std. Err.: se
    CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML

```

Briefly, `meta set` reports that there are 10 studies, that `es` and `se` are the variables used to declare effect sizes and their standard errors, that the default confidence level is 95%, and more. See [Meta settings with meta set](#) in [\[META\] meta data](#) for a detailed description of all settings for this dataset.

We can now use, for example, `meta summarize` to compute the overall effect size (labeled as `theta` in the output below).

```

. meta summarize
  Effect-size label: Effect Size
  Effect size: es
  Std. Err.: se

Meta-analysis summary
Random-effects model
Method: REML
Number of studies = 10
Heterogeneity:
  tau2 = 0.0157
  I2 (%) = 5.30
  H2 = 1.06

```

Study	Effect Size	[95% Conf. Interval]		% Weight
Study 1	1.480	-0.352	3.311	2.30
Study 2	0.999	-0.933	2.931	2.07
Study 3	1.272	0.427	2.117	10.15
Study 4	1.001	0.750	1.252	63.77
Study 5	1.179	-0.527	2.884	2.65
Study 6	1.939	0.427	3.452	3.35
Study 7	2.377	1.005	3.750	4.05
Study 8	0.694	-0.569	1.956	4.75
Study 9	1.099	-0.147	2.345	4.88
Study 10	1.805	-0.151	3.761	2.02
theta	1.138	0.857	1.418	

```

Test of theta = 0: z = 7.95
Test of homogeneity: Q = chi2(9) = 6.34
Prob > |z| = 0.0000
Prob > Q = 0.7054

```

See [\[META\] meta summarize](#) for details about this command.

► Example 2: Declaring effect sizes and confidence intervals

Continuing with [example 1](#), we find that some meta-analysis datasets contain confidence intervals associated with effect sizes instead of standard errors. In that case, you can specify confidence intervals with `meta set` instead of the standard errors. For example, variables `cil` and `ciu` contain the 95% lower and upper CI limits for the effect sizes stored in variable `es`. We can declare them as follows.

```
. meta set es cil ciu
Meta-analysis setting information
Study information
  No. of studies: 10
  Study label: Generic
  Study size: N/A
  Effect size
    Type: Generic
    Label: Effect Size
    Variable: es
  Precision
  Std. Err.: _meta_se
             CI: [_meta_cil, _meta_ciu]
  CI level: 95%, controlled by level()
  User CI: [cil, ciu]
  User CI level: 95%, controlled by civarlevel()
Model and method
  Model: Random-effects
  Method: REML
```

Compared with `Std. Err.:` in [example 1](#), `Std. Err.:` under `Precision` now contains the system variable `_meta_se`; see [System variables](#) in [\[META\] meta data](#). The standard errors are computed from `cil` and `ciu` and stored in this system variable. The CI values are stored in the corresponding system variables `_meta_cil` and `_meta_ciu`.

The output additionally reports the user-specified CI variables, `cil` and `ciu`, under `User CI:` and their corresponding confidence level, 95%, under `User CI level:`. As we will see later, `User CI level`, controlled by the `civarlevel()` option, and `CI level`, controlled by the `level()` option, may be different.

Let's now check that we obtain the same results as before using the equivalent CI declaration.

```
. meta summarize
  Effect-size label: Effect Size
    Effect size: es
    Std. Err.: _meta_se

Meta-analysis summary
Random-effects model
Method: REML

Number of studies = 10
Heterogeneity:
  tau2 = 0.0157
  I2 (%) = 5.30
  H2 = 1.06
```

Study	Effect Size	[95% Conf. Interval]		% Weight
Study 1	1.480	-0.352	3.311	2.30
Study 2	0.999	-0.933	2.931	2.07
Study 3	1.272	0.427	2.117	10.15
Study 4	1.001	0.750	1.252	63.77
Study 5	1.179	-0.527	2.884	2.65
Study 6	1.939	0.427	3.452	3.35
Study 7	2.377	1.005	3.750	4.05
Study 8	0.694	-0.569	1.956	4.75
Study 9	1.099	-0.147	2.345	4.88
Study 10	1.805	-0.151	3.761	2.02
theta	1.138	0.857	1.418	

```
Test of theta = 0: z = 7.95
Test of homogeneity: Q = chi2(9) = 6.34

Prob > |z| = 0.0000
Prob > Q = 0.7054
```

In the [earlier meta set](#), we assumed that the `cil` and `ciu` variables correspond to the 95% CIs. Although typical, this may not always be the case. You can use the `civarlevel()` option to specify the confidence level of the CI variables. We have variables `cil90` and `ciu90` in our dataset, which contain the 90% CIs for `es`. We can use them in the declaration as long as we also specify the `civarlevel(90)` option.

```
. meta set es cil90 ciu90, civarlevel(90)

Meta-analysis setting information

Study information
  No. of studies: 10
  Study label: Generic
  Study size: N/A

Effect size
  Type: Generic
  Label: Effect Size
  Variable: es

Precision
  Std. Err.: _meta_se
  CI: [_meta_cil, _meta_ciu]
  CI level: 95%, controlled by level()
  User CI: [cil90, ciu90]
  User CI level: 90%, controlled by civarlevel()

Model and method
  Model: Random-effects
  Method: REML
```

The `User CI level` now contains 90%. Do not confuse the `civarlevel()` option, whose value is reported in `User CI level`, with the `level()` option, whose value is reported in `CI level`. The former specifies the confidence level corresponding to the declared CI variables. The latter specifies the

confidence level that will be used to compute various confidence intervals during your meta-analysis session. Note that the system CI variables, `_meta_cil` and `_meta_ciu`, always correspond to the confidence level controlled by `level()`.

```
. meta summarize
      Effect-size label: Effect Size
      Effect size:      es
      Std. Err.:       _meta_se

Meta-analysis summary
Random-effects model
Method: REML

Number of studies =    10
Heterogeneity:
  tau2 = 0.0157
  I2 (%) = 5.30
  H2 = 1.06
```

Study	Effect Size	[95% Conf. Interval]		% Weight
Study 1	1.480	-0.352	3.311	2.30
Study 2	0.999	-0.933	2.931	2.07
Study 3	1.272	0.427	2.117	10.15
Study 4	1.001	0.750	1.252	63.77
Study 5	1.179	-0.527	2.884	2.65
Study 6	1.939	0.427	3.452	3.35
Study 7	2.377	1.005	3.750	4.05
Study 8	0.694	-0.569	1.956	4.75
Study 9	1.099	-0.147	2.345	4.88
Study 10	1.805	-0.151	3.761	2.02
theta	1.138	0.857	1.418	

```
Test of theta = 0: z = 7.95          Prob > |z| = 0.0000
Test of homogeneity: Q = chi2(9) = 6.34    Prob > Q = 0.7054
```

Although the specified CI variables corresponded to the 90% confidence level, the CIs reported by `meta summarize` are the 95% CIs because the default confidence level is 95%, `level(95)`.

◀

► Example 3: Declaring meta-analysis methods and models

In *Declaring a meta-analysis model* in [META] **meta data**, we describe the importance of choosing the appropriate meta-analysis model and method for the analysis. Here we demonstrate how to specify different meta-analysis models and methods.

From [example 1](#) and as described in *Default meta-analysis model and method* in [META] **meta data**, the default meta-analysis model and estimation method are random-effects and REML. We can specify a different [random-effects method](#) in the `random()` option. For example, let's use the DerSimonian–Laird estimation method.

```
. meta set es se, random(dlaird)
Meta-analysis setting information
Study information
  No. of studies: 10
  Study label: Generic
  Study size: N/A
  Effect size
    Type: Generic
    Label: Effect Size
    Variable: es
  Precision
  Std. Err.: se
    CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: DerSimonian-Laird
```

meta set reports in Method: that the current method is now DerSimonian–Laird.

We can also choose a different meta-analysis model. For example, we can specify a fixed-effects model by using the fixed option.

```
. meta set es se, fixed
Meta-analysis setting information
Study information
  No. of studies: 10
  Study label: Generic
  Study size: N/A
  Effect size
    Type: Generic
    Label: Effect Size
    Variable: es
  Precision
  Std. Err.: se
    CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Fixed-effects
  Method: Inverse-variance
```

The inverse-variance estimation method is assumed for the fixed-effects model.

We can also specify a common-effect model, although the literature does not recommend starting your meta-analysis with this model.

```
. meta set es se, common
Meta-analysis setting information
Study information
  No. of studies: 10
  Study label: Generic
  Study size: N/A
Effect size
  Type: Generic
  Label: Effect Size
  Variable: es
Precision
  Std. Err.: se
  CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Common-effect
  Method: Inverse-variance
```

The inverse-variance estimation method is assumed for the common-effect model.

As we describe in *Declaring a meta-analysis model* in [META] **meta data**, some of the meta-analysis will not be available for common-effect models. For example, because a common-effect model implies no heterogeneity, you will not be able to perform tests of small-study effects using meta bias in the presence of moderators.

```
. meta bias x, egger
meta bias with moderators not supported with a common-effect model
The declared model is a common-effect model, which assumes no
heterogeneity. Specifying moderators that account for potential
heterogeneity is not valid in this case. You may override this
assumption by specifying one of options fixed or random(remethod).
r(498);
```

See [META] **meta bias**.

◀

► Example 4: Specifying study and effect-size labels, confidence intervals, and more

In *Declaring display settings for meta-analysis* of [META] **meta data**, we describe the options to control the display from the meta commands. Below, we use `studylabel()` and `eslabel()` to specify our own study and effect-size labels, `level(90)` to report the 90% CIs, and `nometashow` to suppress the information about the effect-size and standard-error variables in the output of all meta commands.

```

. meta set es se, studylabel(studylab) eslabel("Mean Diff.") level(90)
> nometashow
Meta-analysis setting information

Study information
  No. of studies: 10
  Study label: studylab
  Study size: N/A
  Effect size
    Type: Generic
    Label: Mean Diff.
    Variable: es
  Precision
  Std. Err.: se
    CI: [_meta_cil, _meta_ciu]
  CI level: 90%

Model and method
  Model: Random-effects
  Method: REML

```

If we now run `meta summarize`, we will see the new labels for the studies in the Study column, the effect-size column labeled as Mean Diff., the 90% CIs, and no meta setting information above the table header.

```

. meta summarize
Meta-analysis summary
Random-effects model
Method: REML
Number of studies = 10
Heterogeneity:
  tau2 = 0.0157
  I2 (%) = 5.30
  H2 = 1.06

```

Study	Mean Diff.	[90% Conf. Interval]		% Weight
Smith et al. (1984)	1.480	-0.057	3.016	2.30
Jones and Miller (1989)	0.999	-0.622	2.620	2.07
Johnson et al. (1991)	1.272	0.563	1.981	10.15
Brown et al. (1995)	1.001	0.790	1.211	63.77
Clark and Thomas (1998)	1.179	-0.252	2.610	2.65
Williams et al. (2003)	1.939	0.670	3.209	3.35
Davis and Wilson (2010)	2.377	1.226	3.529	4.05
Moore and Parker (2014)	0.694	-0.366	1.753	4.75
Miller et al. (2018)	1.099	0.053	2.144	4.88
Assaad et al. (2019)	1.805	0.164	3.446	2.02
theta	1.138	0.902	1.373	

```

Test of theta = 0: z = 7.95
Test of homogeneity: Q = chi2(9) = 6.34
Prob > |z| = 0.0000
Prob > Q = 0.7054

```

◀

► Example 5: Specifying study size

Some analysis such as a funnel plot with sample-size metrics (see [\[META\] meta funnelplot](#)) requires that you specify the sample size for each study with `meta set`. You can use the `studysize()` option for this.

```

. meta set es se, studysize(ssize)
Meta-analysis setting information
Study information
  No. of studies: 10
  Study label: Generic
  Study size: ssize
  Effect size
    Type: Generic
    Label: Effect Size
    Variable: es
  Precision
  Std. Err.: se
    CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML

```

The name of the study-size variable, `ssize`, is now reported in `Study size:`.

◀

Stored results

`meta set` stores the following characteristics and system variables:

Characteristics

<code>_dta[_meta_marker]</code>	“_meta_ds_1”
<code>_dta[_meta_K]</code>	number of studies in the meta-analysis
<code>_dta[_meta_studylabel]</code>	name of string variable containing study labels or <code>Generic</code>
<code>_dta[_meta_studysize]</code>	name of numeric variable containing study sizes, when <code>studysize()</code> specified
<code>_dta[_meta_estype]</code>	type of effect size; <code>Generic</code>
<code>_dta[_meta_eslabelopt]</code>	<code>eslabel(eslab)</code> , if specified
<code>_dta[_meta_eslabel]</code>	effect-size label from <code>eslabel()</code> ; default is <code>Effect Size</code>
<code>_dta[_meta_eslabeldb]</code>	effect-size label for dialog box
<code>_dta[_meta_esvar]</code>	name of effect-size variable
<code>_dta[_meta_esvardb]</code>	abbreviated name of effect-size variable for dialog box
<code>_dta[_meta_sevar]</code>	name of standard-error variable, if specified, or <code>_meta_se</code>
<code>_dta[_meta_cilvar]</code>	name of variable containing lower CI bounds, if specified, or <code>_meta_cil</code>
<code>_dta[_meta_ciuvar]</code>	name of variable containing upper CI bounds, if specified, or <code>_meta_ciu</code>
<code>_dta[_meta_civarlevel]</code>	confidence level associated with CI variables, if specified
<code>_dta[_meta_level]</code>	default confidence level for meta-analysis
<code>_dta[_meta_modellabel]</code>	meta-analysis model label: <code>Random-effects</code> , <code>Common-effect</code> , or <code>Fixed-effects</code>
<code>_dta[_meta_model]</code>	meta-analysis model: <code>random</code> , <code>common</code> , or <code>fixed</code>
<code>_dta[_meta_methodlabel]</code>	meta-analysis method label; varies by meta-analysis model
<code>_dta[_meta_method]</code>	meta-analysis method; varies by meta-analysis model
<code>_dta[_meta_randomopt]</code>	<code>random(remethod)</code> , if specified
<code>_dta[_meta_show]</code>	empty or <code>nometashow</code>
<code>_dta[_meta_datatype]</code>	data type; <code>Generic</code>
<code>_dta[_meta_datavars]</code>	variables specified with <code>meta set</code>
<code>_dta[_meta_setcmdline]</code>	<code>meta set</code> command line
<code>_dta[_meta_ifexp]</code>	<i>if</i> specification
<code>_dta[_meta_inexp]</code>	<i>in</i> specification

System variables

<code>_meta_id</code>	study ID variable
-----------------------	-------------------

<code>_meta_es</code>	variable containing effect sizes
<code>_meta_se</code>	variable containing effect-size standard errors
<code>_meta_cil</code>	variable containing lower bounds of CIs for effect sizes
<code>_meta_ciu</code>	variable containing upper bounds of CIs for effect sizes
<code>_meta_studylabel</code>	string variable containing study labels
<code>_meta_studysize</code>	variable containing total sample size per study

References

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Also see

- [META] [meta data](#) — Declare meta-analysis data
- [META] [meta esize](#) — Compute effect sizes and declare meta-analysis data
- [META] [meta update](#) — Update, describe, and clear meta-analysis settings
- [META] [meta](#) — Introduction to meta
- [META] [Glossary](#)
- [META] [Intro](#) — Introduction to meta-analysis

Title

meta update — Update, describe, and clear meta-analysis settings

[Description](#)
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Description

`meta update` updates certain components of the meta-analysis after it was declared by `meta set` or `meta esize`. This command is useful for updating some of the meta settings without having to fully respecify your meta-analysis variables. The updated settings will be used throughout the rest of your meta-analysis session.

`meta query` reports whether the data in memory are `meta data` and, if they are, displays the current meta setting information identical to that produced by `meta set` or `meta esize`.

`meta clear` clears meta settings, including meta data characteristics and system variables. The original data remain unchanged. You do not need to use `meta clear` before doing another `meta set` or `meta esize`.

Quick start

Check whether data are declared as meta data, and, if they are, describe their current meta-analysis setting information

```
meta query
```

Keep the same meta-analysis setting (specified earlier using `meta set` or `meta esize`), but use a DerSimonian–Laird random-effects model

```
meta update, random(dlaird)
```

Keep the same meta-analysis setting (specified earlier using `meta esize`), but use the log risk-ratio as the effect size

```
meta update, esize(lnrratio)
```

Clear meta-analysis declaration

```
meta clear
```

Menu

Statistics > Meta-analysis

Syntax

Update meta-analysis settings declared using `meta esize` for continuous outcomes

```
meta update [ , options_continuous options ]
```

Update meta-analysis settings declared using `meta esize` for binary outcomes

```
meta update [ , options_binary options ]
```

Update meta-analysis settings declared using `meta set`

```
meta update [ , options_generic options ]
```

Describe meta data

```
meta query [ , short ]
```

Clear meta data

```
meta clear
```

<i>options_continuous</i>	Description
<code>esize(<i>esspecct</i>)</code>	specify effect size for continuous outcome to be used in the meta-analysis
<code>random[(<i>remethod</i>)]</code>	random-effects meta-analysis
<code>common</code>	common-effect meta-analysis; implies inverse-variance method
<code>fixed</code>	fixed-effects meta-analysis; implies inverse-variance method
<i>options_binary</i>	Description
<code>esize(<i>estypebin</i>)</code>	specify effect size for binary outcome to be used in the meta-analysis
<code>random[(<i>remethod</i>)]</code>	random-effects meta-analysis
<code>common[(<i>cefemethod</i>)]</code>	common-effect meta-analysis
<code>fixed[(<i>cefemethod</i>)]</code>	fixed-effects meta-analysis
<code>zerocells(<i>zcspec</i>)</code>	adjust for zero cells during computation; default is to add 0.5 to all cells of those 2×2 tables that contain zero cells
<i>options_generic</i>	Description
<code>random[(<i>remethod</i>)]</code>	random-effects meta-analysis
<code>common</code>	common-effect meta-analysis; implies inverse-variance method
<code>fixed</code>	fixed-effects meta-analysis; implies inverse-variance method
<code>studysize(<i>varname</i>)</code>	total sample size per study

<i>options</i>	Description
<code>studylabel(<i>varname</i>)</code>	variable to be used to label studies in all meta-analysis output
<code>eslabel(<i>string</i>)</code>	effect-size label to be used in all meta-analysis output; default is <code>eslabel(Effect Size)</code>
<code>level(#)</code>	confidence level for all subsequent meta-analysis commands
<code>[no]metashow</code>	display or suppress meta settings in the output

Options

For meta update options, see *Options* of [META] **meta set** and *Options* of [META] **meta esize**.

`short` is used with `meta query`. It displays a short summary of the meta settings containing the information about the declared type of the effect size, effect-size and standard-error variables, and meta-analysis model and estimation method. This option does not appear on the dialog box.

Remarks and examples

When conducting a meta-analysis, you may wish to explore how your results are affected by modifying certain characteristics of your model. For example, suppose you are using log odds-ratios as your effect sizes and the DerSimonian–Laird random-effects model. You want to investigate how your results would change if you were to use log risk-ratios instead. You could use `meta esize`, but you would need to respecify all four of your summary-data variables.

```
. meta esize summary'data, esize(lnrratio) random(dlaird)
```

Instead, you can use `meta update` to simply update the effect sizes.

```
. meta update, esize(lnrratio)
```

`meta update` will run `meta esize` keeping all the model components unchanged except for those you specified.

You can use `meta query` to describe the current meta-analysis settings. With `meta` data in memory, `meta query` produces the same output as `meta set` and `meta esize`. If the data in memory are not declared to be meta data, `meta query` will report the following:

```
. meta query
(data not meta set; use meta set or meta esize to declare as meta data)
```

To clear meta settings, use `meta clear`.

For more details and examples, see *Modifying default meta settings* and *Displaying and updating meta settings* in [META] **meta data**.

Stored results

`meta update` updates characteristics and contents of system variables described in *Stored results* of [META] **meta set** and *Stored results* of [META] **meta esize**.

Also see

[META] [meta data](#) — Declare meta-analysis data

[META] [meta esize](#) — Compute effect sizes and declare meta-analysis data

[META] [meta set](#) — Declare meta-analysis data using generic effect sizes

[META] [meta](#) — Introduction to meta

[META] [Glossary](#)

[META] [Intro](#) — Introduction to meta-analysis

Title

meta forestplot — Forest plots

Description
Options
Also see

Quick start
Remarks and examples

Menu
Methods and formulas

Syntax
References

Description

`meta forestplot` summarizes [meta data](#) in a graphical format. It reports individual effect sizes and the overall effect size (ES), their confidence intervals (CIs), heterogeneity statistics, and more. `meta forestplot` can perform random-effects (RE), common-effect (CE), and fixed-effects (FE) meta-analyses. It can also perform subgroup, cumulative, and sensitivity meta-analyses. For tabular display of meta-analysis summaries, see [\[META\] meta summarize](#).

Quick start

Default forest plot after data are declared by using either `meta set` or `meta esize`

```
meta forestplot
```

Default forest plot after data are declared with `meta set` but with the columns spelled out

```
meta forestplot _id _plot _esci _weight
```

Default forest plot after data are declared with `meta esize` but with the columns spelled out

```
meta forestplot _id _data _plot _esci _weight
```

As above, but with the weights omitted

```
meta forestplot _id _data _plot _esci
```

Same as above, but the columns are rearranged

```
meta forestplot _id _data _esci _plot
```

Change the format of the `_esci` column

```
meta forestplot, columnopts(_esci, format(%7.4f))
```

Forest plot based on subgroup meta-analysis

```
meta forestplot, subgroup(groupvar)
```

Forest plot based on cumulative meta-analysis

```
meta forestplot, cumulative(ordervar)
```

Menu

Statistics > Meta-analysis

Syntax

```
meta forestplot [column_list] [ , options ]
```

column_list is a list of column names given by *col*. In the *Meta-Analysis Control Panel*, the columns can be specified on the **Forest plot** tab of the Forest plot pane.

<i>options</i>	Description
Main	
<code>random</code> [(<i>remethod</i>)]	random-effects meta-analysis
<code>common</code> [(<i>cefemethod</i>)]	common-effect meta-analysis
<code>fixed</code> [(<i>cefemethod</i>)]	fixed-effects meta-analysis
<i>reopts</i>	random-effects model options
<code>subgroup</code> (<i>varlist</i>)	subgroup meta-analysis for each variable in <i>varlist</i>
<code>cumulative</code> (<i>cumulspec</i>)	cumulative meta-analysis
Options	
<code>level</code> (#)	set confidence level; default is as declared for meta-analysis
<i>eform_option</i>	report exponentiated results
<code>tdistribution</code>	report <i>t</i> test instead of <i>z</i> test
[no]metashow	display or suppress meta settings in the output
Maximization	
<i>maximize_options</i>	control the maximization process; seldom used
Forest plot	
<code>columnopts</code> (<i>col</i> , [<i>colopts</i>])	column options; can be repeated
<code>cibind</code> (<i>bind</i>)	change binding of CIs for columns <code>_esci</code> and <code>_ci</code> ; default is <code>cibind(brackets)</code>
<code>sebind</code> (<i>bind</i>)	change binding of standard errors for column <code>_esse</code> ; default is <code>sebind(parentheses)</code>
<code>nohrule</code>	suppress horizontal rule
<code>hruleopts</code> (<i>hrule_options</i>)	change look of horizontal rule
<i>text_options</i>	change looks of text options such as column titles, supertitles, and more
<i>plot_options</i>	change look or suppress markers, restrict range of CIs, and more
<i>test_options</i>	suppress information about heterogeneity statistics and tests
<i>twoway_options</i>	any options other than <code>by()</code> documented in [G-3] <i>twoway_options</i>

<i>col</i>	Description
Default columns and order	
<code>_id</code>	study label
<code>_data</code>	summary data; <code>_data1</code> and <code>_data2</code> (only after <code>meta esize</code>)
<code>_plot</code>	forest graph
<code>_esci</code>	effect size and its confidence interval
<code>_weight</code>	percentage of total weight given to each study
Summary-data columns and order	
Continuous outcomes	
Treatment group	
<code>_data1</code>	summary data for treatment group; <code>_n1</code> , <code>_mean1</code> , and <code>_sd1</code>
<code>_n1</code>	sample size in the treatment group
<code>_mean1</code>	mean in the treatment group
<code>_sd1</code>	standard deviation in the treatment group
Control group	
<code>_data2</code>	summary data for control group; <code>_n2</code> , <code>_mean2</code> , and <code>_sd2</code>
<code>_n2</code>	sample size in the control group
<code>_mean2</code>	mean in the control group
<code>_sd2</code>	standard deviation in the control group
Dichotomous outcomes	
Treatment group	
<code>_data1</code>	summary data for treatment group; <code>_a</code> and <code>_b</code>
<code>_a</code>	number of successes in the treatment group
<code>_b</code>	number of failures in the treatment group
Control group	
<code>_data2</code>	summary data for control group; <code>_c</code> and <code>_d</code>
<code>_c</code>	number of successes in the control group
<code>_d</code>	number of failures in the control group
Other columns	
<code>_es</code>	effect size
<code>_ci</code>	confidence interval for effect size
<code>_lb</code>	lower confidence limit for effect size
<code>_ub</code>	upper confidence limit for effect size
<code>_se</code>	standard error of effect size
<code>_esse</code>	effect size and its standard error
<code>_pvalue</code>	<i>p</i> -value for significance test with <code>subgroup()</code> or <code>cumulative()</code>
<code>_K</code>	number of studies with <code>subgroup()</code>
<code>_order</code>	order variable for cumulative meta-analysis with <code>cumulative()</code>

Columns `_data`, `_data1`, `_data2`, and the other corresponding data columns are not available after the declaration by using `meta set`.

Columns `_n1`, `_mean1`, `_sd1`, `_n2`, `_mean2`, and `_sd2` are available only after the declaration by using `meta esize` for continuous outcomes.

Columns `_a`, `_b`, `_c`, and `_d` are available only after the declaration by using `meta esize` for binary outcomes.

Column `_pvalue` is available only when option `cumulative()` or `subgroup()` with multiple variables is specified.

Column `_K` is available only when option `subgroup()` with multiple variables is specified.

<i>colopts</i>	Description
<u>supertitle</u> (<i>string</i>)	super title specification
<u>title</u> (<i>string</i>)	title specification
<u>format</u> (<i>%fmt</i>)	numerical format for column items
<u>mask</u> (<i>mask</i>)	string mask for column items
<u>plotregion</u> (<i>region_options</i>)	attributes of plot region
<i>textbox_options</i>	appearance of textboxes

<i>text_options</i>	Description
<u>coltitleopts</u> (<i>textbox_options</i>)	change look of column titles and supertitles
<u>itemopts</u> (<i>textbox_options</i>)	change look of study rows
<u>overallopts</u> (<i>textbox_options</i>)	change look of the overall row
<u>groupopts</u> (<i>textbox_options</i>)	change look of subgroup rows
<u>bodyopts</u> (<i>textbox_options</i>)	change look of study, subgroup, and overall rows
<u>nonotes</u>	suppress notes about the meta-analysis model, method, and more

<i>plot_options</i>	Description
<u>crop</u> (# #)	restrict the range of CI lines
<u>ciopts</u> (<i>ci_options</i>)	change look of CI lines (size, color, etc.)
<u>nomarkers</u>	suppress study markers
<u>markeropts</u> (<i>marker_options</i>)	change look of study markers (size, color, etc.)
<u>nowmarkers</u>	suppress weighting of study markers
<u>noomarker</u>	suppress the overall marker
<u>omarkeropts</u> (<i>marker_options</i>)	change look of the overall marker (size, color, etc.)
<u>nogmarkers</u>	suppress subgroup markers
<u>gmarkeropts</u> (<i>marker_options</i>)	change look of subgroup markers (size, color, etc.)

<i>test_options</i>	Description
<u>noohetstats</u>	suppress overall heterogeneity statistics
<u>noohomtest</u>	suppress overall homogeneity test
<u>noosigtest</u>	suppress test of significance of overall effect size
<u>noghetstats</u>	suppress subgroup heterogeneity statistics
<u>nogwhomtests</u>	suppress within-subgroup homogeneity tests
<u>nogbhomtests</u>	suppress between-subgroup homogeneity tests

Options

Main

`random`[(*remethod*)], `common`[(*cefemethod*)], `fixed`[(*cefemethod*)], `subgroup`(*varlist*), and `cumulative`(*cumulspec*); see [Options](#) in [META] **meta summarize**.

reopts are `tau2`(#), `i2`(#), and `se`(*seadj*). These options are used with random-effects meta-analysis. See [Options](#) in [META] **meta summarize**.

Options

`level(#)`, `eform_option`, `tdistribution`, and `[no]metashow`; see [Options](#) in [\[META\] meta summarize](#).

Maximization

`maximize_options`: `iterate(#)`, `tolerance(#)`, `nrtolerance(#)`, `nonrtolerance`, `from(#)`, and `showtrace`; see [Options](#) in [\[META\] meta summarize](#).

Forest plot

`columnopts(col [, colopts])` changes the look of the column identified by *col*. This option can be repeated.

colopts are the following options:

`supertitle(string)` specifies that the column's supertitle is *string*.

`title(string)` specifies that the column's title is *string*.

`format(%fmt)` specifies the format for the column's numerical values.

`mask(mask)` specifies a string composed of formats for the column's statistics. For example, *mask* for column `_weight` that identifies the column of weight percentages may be specified as `"%6.2f %%"`.

`plotregion(region_options)` modifies attributes for the plot region. You can change the margins, background color, an outline, and so on; see [\[G-3\] region_options](#).

textbox_options affect how the column's items (study and group) are rendered. These options override what is specified in global options `bodyopts()`, `itemopts()`, and `groupopts()`. See [\[G-3\] added_text_options](#).

Options `format()`, `mask()`, and *textbox_options* are ignored by `_plot`.

`cibind(bind)` changes the binding of the CIs for columns `_esci` and `_ci`. *bind* is one of `brackets` or `parentheses`. By default, the CIs are bound by using brackets, `cibind(brackets)`. This option is relevant only when `_esci` or `_ci` appears in the plot.

`sebind(bind)` changes the binding of the standard errors for column `_esse`. *bind* is one of `parentheses` or `brackets`. By default, the standard errors are bound by using parentheses, `cibind(parentheses)`. This option is relevant only when `_esse` appears in the plot.

`nohrule` suppresses the horizontal rule.

`hruleopts(hrule_options)` affects the look of the horizontal rule.

hrule_options are the following options:

`lcolor(colorstyle)` specifies the color of the rule; see [\[G-4\] colorstyle](#).

`lwidth(linewidthstyle)` specifies the width of the rule; see [\[G-4\] linewidthstyle](#).

`lalign(linealignmentstyle)` specifies the alignment of the rule; see [\[G-4\] linealignmentstyle](#).

`lpattern(linepatternstyle)` specifies the line pattern of the rule; see [\[G-4\] linepatternstyle](#).

`lstyle(linestyle)` specifies the overall style of the rule; see [\[G-4\] linestyle](#).

`margin(marginstyle)` specifies the margin of the rule; see [\[G-4\] marginstyle](#).

text_options are the following options:

`coltitleopts(textbox_options)` affects the look of text for column titles and supertitles. See [\[G-3\] textbox_options](#).

`itemopts(textbox_options)` affects the look of text for study rows; see [G-3] [textbox_options](#).

This option is ignored when option `subgroup()` is specified and contains multiple variables or when option `cumulative()` is specified.

`overallopts(textbox_options)` affects the look of text for the overall row.

See [G-3] [textbox_options](#).

`groupopts(textbox_options)` (synonym `subgroupopts()`) affects the look of text for subgroup rows when option `subgroup()` is specified. See [G-3] [textbox_options](#).

`bodyopts(textbox_options)` affects the look of text for study, subgroup, and overall rows. See [G-3] [textbox_options](#).

`nonotes` suppresses the notes displayed on the graph about the specified meta-analysis model and method and the standard-error adjustment.

`plot_options` are the following options:

`crop(#1 #2)` restricts the range of the CI lines to be between $\#_1$ and $\#_2$. A missing value may be specified for any of the two values to indicate that the corresponding limit should not be cropped. Otherwise, lines that extend beyond the specified value range are cropped and adorned with arrows. This option is useful in the presence of small studies with large standard errors, which lead to confidence intervals that are too wide to be displayed nicely on the graph. Option `crop()` may be used to handle this case.

`ciopts(ci_options)` affects the look of the CI lines and, in the presence of cropped CIs (see option `crop()`), arrowheads.

`ci_options` are any options documented in [G-3] [line_options](#) and the following options of [G-2] [graph twoway pcarrow](#): `mstyle()`, `msize()`, `mangle()`, `barbsize()`, `mcolor()`, `mfcolor()`, `mlcolor()`, `mlwidth()`, `mlstyle()`, and `color()`.

`nomarkers` suppresses the study markers.

`markeropts(marker_options)` affects the look of the study markers.

`marker_options`: `msymbol()`, `mcolor()`, `mfcolor()`, `mlcolor()`, `mlwidth()`, `mlalign()`, `mlstyle()`, and `mstyle()`; see [G-3] [marker_options](#).

`nowmarkers` suppresses weighting of the study markers.

`nowmarkers`, `markeropts()`, and `nowmarkers` are ignored when option `subgroup()` is specified and contains multiple variables or when option `cumulative()` is specified.

`nooverallmarker` suppresses the overall marker.

`overallmarkeropts(marker_options)` affects the look of the overall marker.

`marker_options`: `mcolor()`, `mfcolor()`, `mlcolor()`, `mlwidth()`, `mlalign()`, `mlstyle()`, and `mstyle()`; see [G-3] [marker_options](#).

`nosubgroupmarkers` suppresses the subgroup markers.

`subgroupmarkeropts(marker_options)` affects the look of the subgroup markers.

`marker_options`: `mcolor()`, `mfcolor()`, `mlcolor()`, `mlwidth()`, `mlalign()`, `mlstyle()`, and `mstyle()`; see [G-3] [marker_options](#).

`nosubgroupmarkers` and `subgroupmarkeropts()` are ignored when option `subgroup()` is not specified.

`test_options` are defined below. These options are not relevant with cumulative meta-analysis.

`nohetstats` suppresses overall heterogeneity statistics reported under the Overall row heading on the plot.

`noohomtest` suppresses the overall homogeneity test labeled as **Test of $\theta_i=\theta_j$** under the **Overall** row heading on the plot.

`noosigtest` suppresses the test of significance of the overall effect size labeled as **Test of $\theta=0$** under the **Overall** row heading on the plot. This test is not reported with subgroup analyses, and thus this option is implied when option `subgroup()` is specified.

`noghetstats` suppresses subgroup heterogeneity statistics reported when a single subgroup analysis is performed, that is, when option `subgroup()` is specified with one variable. These statistics are reported under the group-specific row headings.

`nogwhomtests` suppresses within-subgroup homogeneity tests. These tests investigate the differences between effect sizes of studies within each subgroup. These tests are reported when a single subgroup analysis is performed, that is, when option `subgroup()` is specified with one variable. The tests are labeled as **Test of $\theta_i=\theta_j$** under the group-specific row headings.

`nogbhomtests` suppresses between-subgroup homogeneity tests reported when any subgroup analysis is performed, that is, when option `subgroup()` is specified. These tests investigate the differences between the subgroup overall effect sizes and are labeled as **Test of group differences** on the plot.

`twoway_options` are any of the options documented in [G-3] *twoway_options*, excluding `by()`. These include options for titling the graph (see [G-3] *title_options*) and for saving the graph to disk (see [G-3] *saving_option*).

Remarks and examples

Remarks are presented under the following headings:

Overview

Using meta forestplot

Plot columns

Examples of using meta forestplot

Overview

Meta-analysis results are often presented using a forest plot (for example, [Lewis and Ellis \[1982\]](#)). A forest plot shows effect-size estimates and their confidence intervals for each study and, usually, the overall effect size from the meta-analysis (for example, [Lewis and Clarke \[2001\]](#); [Harris et al. \[2016\]](#); and [Fisher 2016](#)). Each study is represented by a square with the size of the square being proportional to the study weight; that is, larger squares correspond to larger (more precise) studies. The weights depend on the chosen meta-analysis model and method. Studies' CIs are plotted as whiskers extending from each side of the square and spanning the width of the CI. Heterogeneity measures such as the I^2 and H^2 statistics, homogeneity test, and the significance test of the overall effect sizes are also commonly reported.

A subgroup meta-analysis forest plot also shows group-specific results. Additionally, it reports a test of the between-group differences among the overall effect sizes. A cumulative meta-analysis forest plot shows the overall effect sizes and their CIs by accumulating the results from adding one study at a time to each subsequent analysis. By convention, group-specific and overall effect sizes are represented by diamonds centered on their estimated values with the diamond width corresponding to the CI length.

For more details about forest plots, see, for instance, [Anzures-Cabrera and Higgins \(2010\)](#). Also see [Schriger et al. \(2010\)](#) for an overview of their use in practice.

Using meta forestplot

`meta forestplot` produces meta-analysis forest plots. It provides a graphical representation of the results produced by `meta summarize` and thus supports most of its options such as those specifying a meta-analysis model and estimation method; see [META] [meta summarize](#).

The default look of the forest plot produced by `meta forestplot` depends on the type of analysis. For basic meta-analysis, `meta forestplot` plots the study labels, effect sizes and their confidence intervals, and percentages of total weight given to each study. If `meta esize` was used to declare `meta data`, summary data are also plotted. That is,

```
. meta forestplot
```

is equivalent to typing

```
. meta forestplot _id _plot _esci _weight
```

after declaration by using `meta set` and to typing

```
. meta forestplot _id _data _plot _esci _weight
```

after declaration by using `meta esize`.

If multiple variables are specified in the `subgroup()` option,

```
. meta forestplot, subgroup(varlist)
```

is equivalent to typing

```
. meta forestplot _id _K _plot _esci _pvalue, subgroup(varlist)
```

For cumulative meta-analysis,

```
. meta forestplot, cumulative(varname)
```

is equivalent to typing

```
. meta forestplot _id _plot _esci _pvalue _order, cumulative(varname)
```

You can also specify any of the supported columns with `meta forestplot`. See [Plot columns](#) for details about the supported columns.

The CIs correspond to the confidence level as declared by `meta set` or `meta esize`. You can specify a different level in the `level()` option. Also, by default, the CIs are bound by using brackets. You can specify `cibind(parentheses)` to use parentheses instead.

As we mentioned earlier, you can produce forest plots for subgroup analysis by using the `subgroup()` option and for cumulative meta-analysis by using the `cumulative()` option.

You can modify the default column supertitles, titles, formats, and so on by specifying the `columnopts()` option. You can repeat this option to modify the look of particular columns. If you want to apply the same formatting to multiple columns, you can specify these columns within `columnopts()`. See [Options](#) for the list of supported column options.

Another option you may find useful is `crop(#1 #2)`. Sometimes, some of the smaller studies may have large standard errors that lead to CIs that are too wide to be displayed on the plot. You can “crop” such CIs by restricting their range. The restricted range will be indicated by the arrowheads at the corresponding ends of the CIs. You can crop both limits or only one of the limits. You can modify the default look of the arrowheads or CI lines, in general, by specifying `ciopts()`.

You can specify many more options to customize the look of your forest plot such as modifying the look of text for column titles in `coltitleopts()` or the column format in `format()`; see [Syntax](#) for details.

`meta forestplot` uses the following default convention when displaying the results. The results from individual studies—individual effects sizes—are plotted as navy squares with areas proportional to study weights. The overall effect size is plotted as a green (or, more precisely, forest green using Stata's color convention) diamond with the width corresponding to its CI. The results of a single subgroup analysis—subgroup effect sizes—are plotted as maroon diamonds with the widths determined by the respective CIs. The results of multiple subgroup analyses are plotted as maroon circles with the CI lines. The cumulative meta-analysis results—cumulative overall effect sizes—are displayed as green circles with CI lines.

Options `itemopts()`, `nomarkers`, and `markeropts()` control the look of study rows and markers, which represent individual effect sizes. These options are not relevant when individual studies are not reported such as with multiple subgroup analysis and cumulative meta-analysis.

Options `groupopts()`, `nogmarkers`, and `gmarkeropts()` control the look of subgroup rows and markers and are relevant only when subgroup analysis is performed by specifying the `subgroup()` option.

Options `overallopts()`, `noomarker`, and `omarkeropts()` control the look of overall rows and markers, which represent the overall effect sizes. These options are always applicable because the overall results are always displayed by default. With cumulative meta-analysis, these options affect the displayed cumulative overall effect sizes.

Graphs created by `meta forestplot` cannot be combined with other Stata graphs using `graph combine`.

Plot columns

`meta forestplot` supports many columns that you can include in your forest plot; see the list of [supported columns](#) in *Syntax*. The default columns plotted for various analyses were described in *Using meta forestplot* above. Here we provide more details about some of the supported columns.

`meta forestplot` provides individual columns such as `_es` and `_se` and column shortcuts such as `_esse`. Column shortcuts are typically shortcuts for specifying multiple columns. For instance, column `_data` is a shortcut for columns `_data1` and `_data2`, which themselves are shortcuts to individual summary-data columns. For continuous data, `_data1` is a shortcut for columns `_n1`, `_mean1`, and `_sd1`, and `_data2` is a shortcut for `_n2`, `_mean2`, and `_sd2`. For binary data, `_data1` corresponds to the treatment-group numbers of successes and failures, `_a` and `_b`, and `_data2` to the respective numbers in the control group, `_c` and `_d`. Column `_data` and the corresponding summary-data columns are available only after declaration by using `meta esize`.

The other column shortcuts are `_ci`, `_esci`, and `_esse`. In addition to serving as shortcuts to the respective columns (`_lb` and `_ub`; `_es`, `_lb`, and `_ub`; and `_es` and `_se`), these shortcut columns have additional properties. For instance, when you specify `_ci`, the lower and upper CI bounds are separated with a comma, bounded in brackets, and share a title. That is,

```
. meta forestplot _ci
```

is similar to specifying

```
. meta forestplot _lb _ub,
> columnopts(_lb _ub, title(95% CI))
> columnopts(_lb, mask("[%6.2f,")")
> columnopts(_ub, mask("%6.2f]"))
```

Similarly, `_esci` additionally combines `_es` and `_ci` with the common column title, and `_esse` combines `_es` and `_se` and bounds the standard errors in parentheses. `_ci`, `_esci`, and `_esse` also apply other properties to improve the default look of the specified columns such as modifying the default column margins by specifying `plotregion(margin())`.

If you want to modify the individual columns of the shortcuts, you need to specify the corresponding column names in `columnopts()`. For instance, if we want to display the effect sizes of the `_esci` column with three decimal digits but continue using the default format for CIs, we can type

```
. meta forestplot _esci, columnopts(_es, format(%6.3f))
```

If we specify `_esci` instead of `_es` in `columnopts()`,

```
. meta forestplot _esci, columnopts(_esci, format(%6.3f))
```

both effect sizes and CIs will be displayed with three decimal digits. On the other hand, if we want to change the default title and supertitle for `_esci`, we should specify `_esci` in `columnopts()`,

```
. meta forestplot _esci, columnopts(_esci, supertitle("My ES") title("with my CI"))
```

Also see [example 6](#) and [example 7](#) for more examples of customizing the default look of columns.

Column `_plot` corresponds to the plot region that contains graphical representation of the effect sizes and their confidence intervals. You can modify the default look of the plot by specifying the *plot_options* in *Syntax*.

Column `_es` corresponds to the plotted effect sizes. For basic meta-analysis, this column displays the individual and overall effect sizes. For subgroup meta-analysis, it also displays subgroup-specific overall effect sizes. For cumulative meta-analysis, it displays the overall effect sizes corresponding to the accumulated studies.

Some of the columns such as `_pvalue`, `_K`, and `_order` are available only with specific meta-analyses. `_pvalue` is available only with multiple subgroup analyses or with cumulative analysis; it displays the *p*-values of the significant tests of effect sizes. `_K` is available with multiple subgroup analyses and displays the number of studies within each subgroup. `_order` is available only with cumulative meta-analysis; it displays the values of the specified ordering variable.

Duplicate columns are ignored with `meta forestplot`. Also, column shortcuts take precedence. That is, if you specified both `_es` and `_esci`, the latter will be displayed.

Examples of using meta forestplot

In this section, we demonstrate some of the uses of `meta forestplot`.

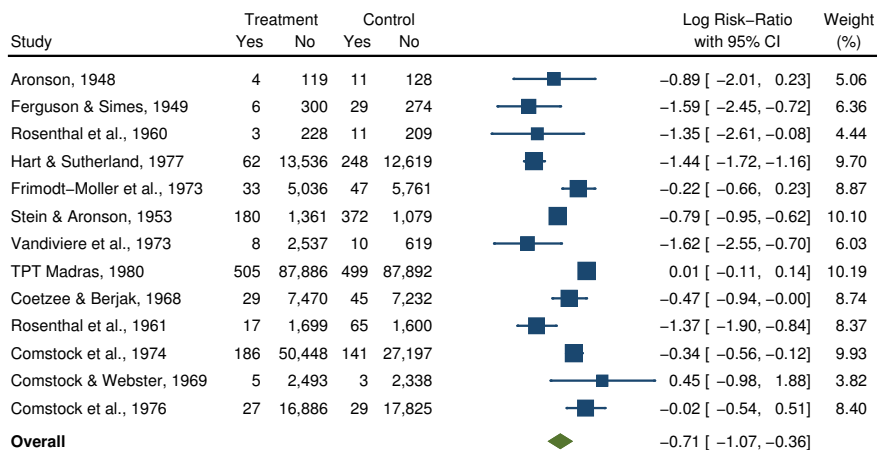
► Example 1: Forest plot for binary data

Consider the dataset from [Colditz et al. \(1994\)](#) of clinical trials that studies the efficacy of a Bacillus Calmette-Guérin (BCG) vaccine in the prevention of tuberculosis (TB). This dataset was introduced in *Efficacy of BCG vaccine against tuberculosis (bcg.dta)* of [META] `meta`. In this section, we use its declared version and focus on the demonstration of various options of `meta forest`.

```
. use https://www.stata-press.com/data/r16/bcgset
(Efficacy of BCG vaccine against tuberculosis; set with -meta esize-)
```

Let's construct a basic forest plot by simply typing

```
. meta forestplot
Effect-size label: Log Risk-Ratio
Effect size: _meta_es
Std. Err.: _meta_se
Study label: studylbl
```



Heterogeneity: $\tau^2 = 0.31$, $I^2 = 92.22\%$, $H^2 = 12.86$
 Test of $\theta_1 = \theta_0$: $Q(12) = 152.23$, $p = 0.00$
 Test of $\theta = 0$: $z = -3.97$, $p = 0.00$



Random-effects REML model

By default, the basic forest plot displays the study labels (column `_id`), the summary data (`_data`), graphical representation of the individual and overall effect sizes and their CIs (`_plot`), the corresponding values of the effect sizes and CIs (`_esci`), and the percentages of total weight for each study (`_weight`). You can also customize the columns on the forest plot; see [example 6](#).

In the graph, each study corresponds to a navy square centered at the point estimate of the effect size with a horizontal line (whiskers) extending on either side of the square. The horizontal line depicts the CI. The area of the square is proportional to the corresponding study weight.

The overall effect size corresponds to the green diamond centered at the estimate of the overall effect size. The width of the diamond corresponds to the width of the overall CI. Note that the height of the diamond is irrelevant. It is customary in meta-analysis forest plots to display an overall effect size as a diamond filled inside with color. This, however, may overemphasize the actual area of the diamond whereas only the width of it matters. If desired, you may suppress the fill color by specifying the `omarkeropts(mfcolor(none))` option.

Under the diamond, three lines are reported. The first line contains heterogeneity measures I^2 , H^2 , and $\hat{\tau}^2$. The second line displays the homogeneity test based on the Q statistic. The third line displays the test of the overall effect size being equal to zero. These lines may be suppressed by specifying options `noohetstats`, `noohomtest`, and `noosigtest`. See [\[META\] meta summarize](#) for a substantive interpretation of these results.

`meta forestplot` provides a quick way to assess between-study heterogeneity visually. In the absence of heterogeneity, we would expect to see that the middle points of the squares are close to the middle of the diamond and the CIs are overlapping. In these data, there is certainly evidence of

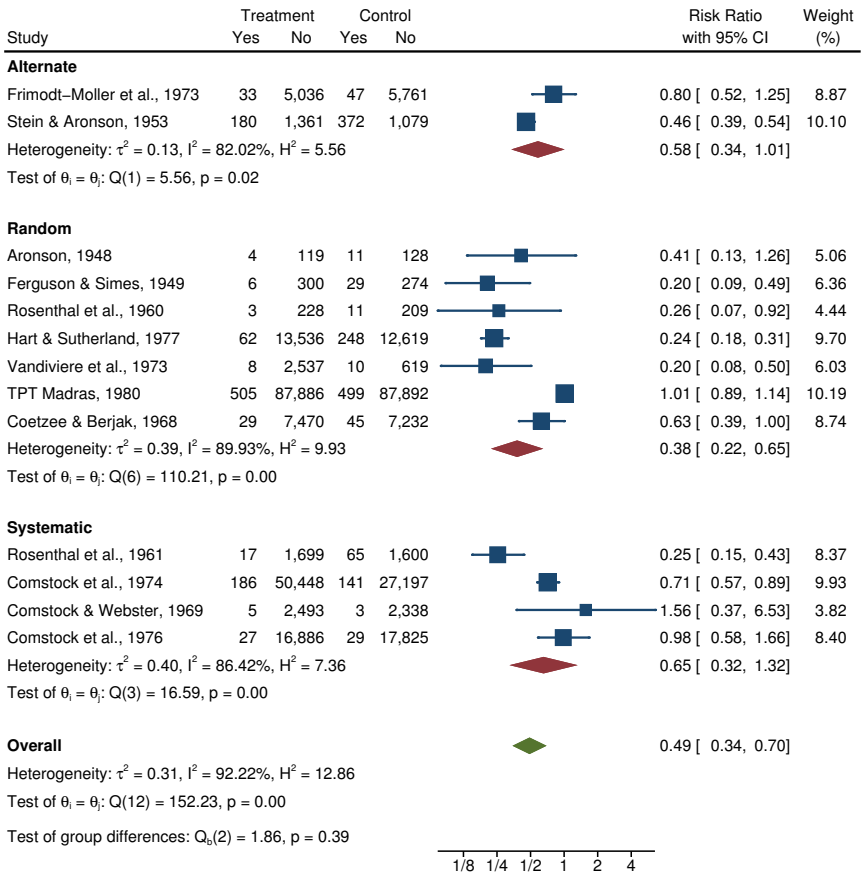
some heterogeneity because the squares for some studies are far away from the diamond and there are studies with nonoverlapping CIs.



► Example 2: Subgroup-analysis forest plot

Continuing with [example 1](#), let's now perform a subgroup meta-analysis based on the method of treatment allocation recorded in variable `alloc`. We specify `subgroup(alloc)` and also use the `eform` option to display exponentiated results, risk ratios instead of log risk-ratios in our example.

```
. meta forestplot, subgroup(alloc) eform
Effect-size label: Log Risk-Ratio
Effect size:      _meta_es
Std. Err.:       _meta_se
Study label:     studylbl
```



Random-effects REML model

In addition to the overall results, the forest plot shows the results of meta-analysis for each of the three groups. With subgroup meta-analysis, each group gets its own maroon diamond marker that represents the group-specific overall effect size. Just like with the overall diamond, only the widths

(not the heights) of the group-specific diamonds are relevant on the plot. Similarly to the overall marker, you can specify the `gmarkeropts(mfcolor(none))` option to suppress the fill color for the group-specific diamonds.

Heterogeneity measures and homogeneity tests are reported at the bottom (below the group-specific diamond marker) within each group. These provide information regarding the heterogeneity among the studies within each group. They may be suppressed with options `noghetstats` and `nogwhomtests`, respectively. Notice that the significance test of the overall effect size equal to zero is no longer reported under Overall summary for subgroup analysis.

A test of between-group differences based on the Q_b statistic is reported at the bottom. This test investigates the difference between the group-specific overall effect sizes. It may be suppressed with `nogbhomtests`.

You may also specify multiple variables in `subgroup()`, in which case a separate subgroup analysis is performed for each variable; see [example 5](#) for details.

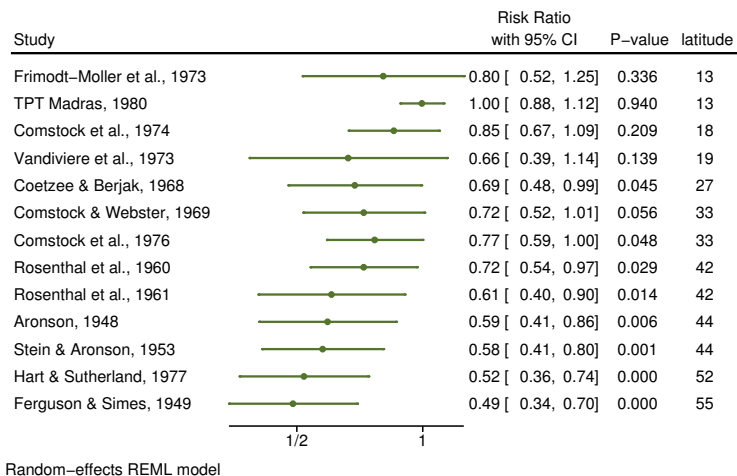


► Example 3: Cumulative forest plot

Continuing with [example 1](#), we now perform a cumulative meta-analysis in the ascending order of variable `latitude`. You can specify suboption `descending` within the `cumulative()` option to request a descending order.

We also specify `rr`, which is a synonym of the `eform` option we used in [example 2](#), to display the risk ratios instead of the default log risk-ratios.

```
. meta forestplot, cumulative(latitude) rr
Effect-size label:  Log Risk-Ratio
Effect size:       _meta_es
Std. Err.:        _meta_se
Study label:      studylbl
```

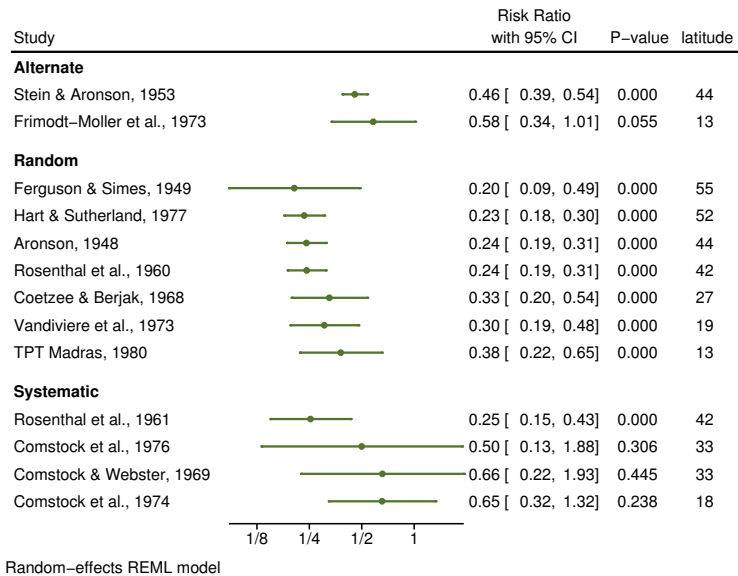


By default, the cumulative meta-analysis forest plot displays the study labels (`_id`), the plot of effect sizes and their CIs (`_plot`), the values of effect sizes and their CIs (`_esci`), the p -values (`_pvalue`) of the corresponding significance tests of the effect sizes, and the values of the order variable (`_order`).

The displayed effect sizes correspond to cumulative overall effect sizes or the overall effect sizes computed for each set of accumulated studies. To distinguish them from study-specific effect sizes, we plot them as unweighted circles using the same color, green, as the overall effect size in a standard meta-analysis forest plot. You can change the default style and color of the markers by specifying the `omarkeropts()` option. The corresponding CIs of the cumulative effect sizes are plotted as CI lines.

We may construct a cumulative forest plot stratified by a covariate by specifying `by()` within `cumulative()`. For example, let's stratify our cumulative analysis by the method of treatment allocation recorded in variable `alloc`.

```
. meta forestplot, cumulative(latitude, by(alloc) descending) rr
Effect-size label: Log Risk-Ratio
Effect size:      _meta_es
Std. Err.:       _meta_se
Study label:      studylbl
```



We specified that the analysis be conducted in the descending order of the `latitude` variable. The stratified forest plot shows the same columns as before but the cumulative analysis is performed separately for each group of `alloc`. A consistent pattern is observed across all three groups—risk ratios tend to increase as latitude decreases.

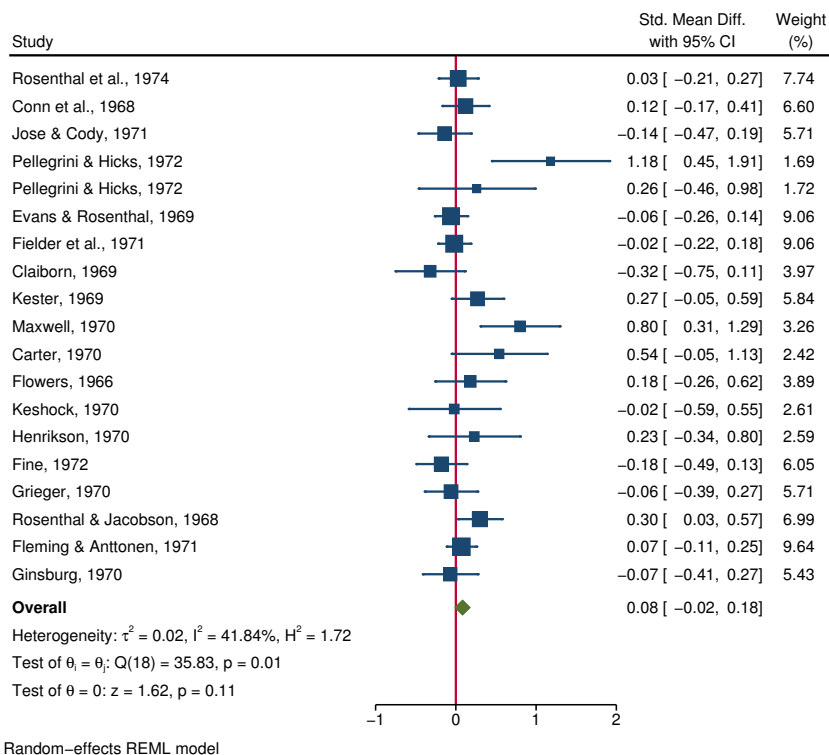
Example 4: Forest plot for precomputed effect sizes

Recall the pupil IQ data (Raudenbush and Bryk 1985; Raudenbush 1984) described in *Effects of teacher expectancy on pupil IQ (pupiliq.dta)* of [META] meta. Here we will use its declared version (declared with meta set) to construct a forest plot of precomputed effect sizes.

```
. use https://www.stata-press.com/data/r16/pupiliqset, clear
(Effects of teacher expectancy on pupil IQ; set with -meta set-)
```

We specify the `xline(0)` option to show the no-effect line at 0. Effect sizes with corresponding CIs that cross this line are not statistically significant at the 5% level.

```
. meta forestplot, xline(0)
Effect-size label: Std. Mean Diff.
Effect size: stdmdiff
Std. Err.: se
Study label: studylbl
```

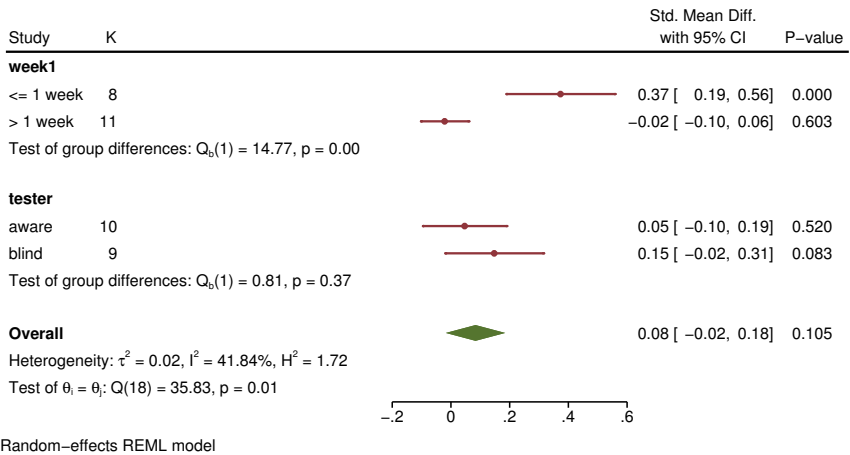


When meta data are declared by using `meta set` (that is, when we are working with precomputed effect sizes), the `_data` column is not available. If desired, you may plot the values of effect sizes and their standard errors by specifying the `_esse` column. Other components of the graph are interpreted as in [example 1](#).

Example 5: Multiple subgroup-analyses forest plot

Continuing with [example 4](#), we will conduct multiple subgroup analyses and summarize their results on a forest plot. We specify variables `tester` and `week1` in `subgroup()` as follows:

```
. meta forestplot, subgroup(week1 tester)
Effect-size label: Std. Mean Diff.
Effect size: stdmdiff
Std. Err.: se
Study label: studylbl
```



By default, the forest plot displays the study labels (`_id`), the number of studies within each group (`_K`), the plot of effect sizes and their CIs (`_plot`), the values of effect sizes and their CIs (`_esci`), and the p -values (`_pvalue`) of the corresponding significance tests.

To keep the output compact, the forest plot does not report individual studies, only the number of studies in each group. The between-group homogeneity test based on the Q_b is reported for each subgroup analysis. For example, for subgroup analysis based on variable `week1`, there are two groups, `<= 1 week` and `> 1 week`. The test investigates whether the overall effect sizes corresponding to these two groups are the same. The results of this test are identical to those we would have obtained if we had specified `subgroup(week1)`.

Just like with cumulative meta-analysis in [example 3](#), `meta forestplot` uses unweighted circles and CI lines to display the overall group-specific effect sizes and their CIs. But here the circles are displayed in maroon—the same color used to display the group-specific diamonds in a single-variable subgroup analysis (see [example 2](#)).

Example 6: Modifying columns' order and cropping confidence intervals

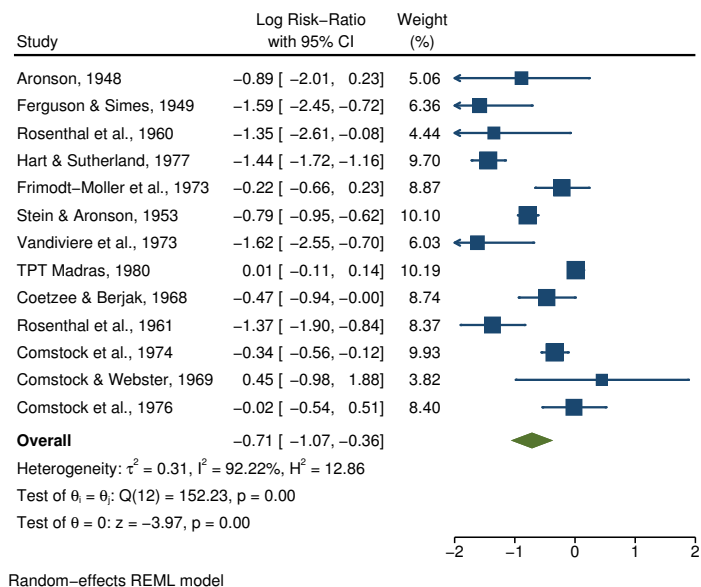
For this and the following examples, let's return to our BCG dataset from [example 1](#).

```
. use https://www.statapress.com/data/r16/bcgset, clear
(Efficacy of BCG vaccine against tuberculosis; set with -meta esize-)
. meta update, nometashow
```

We used `meta update` to suppress the meta setting information displayed by `meta forest` for the rest of our meta-analysis.

We can choose which columns to display and the order in which to display them in the forest plot by specifying the corresponding column names in the desired order. In the code below, we display the study labels first, followed by the effect sizes and their CIs, then weights, and finally the plot. We also use the `crop(-2 .)` option to restrict the range of the CIs at a lower limit of `-2`.

```
. meta forestplot _id _esci _weight _plot, crop(-2 .)
```



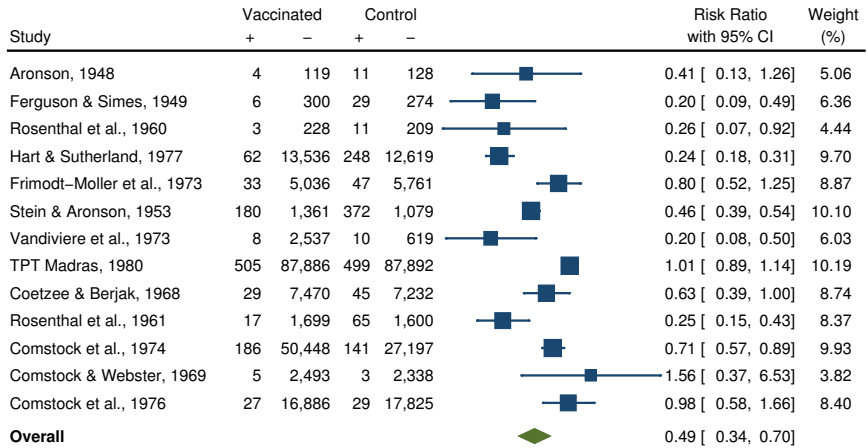
CIs that extend beyond the lower limit of `-2` are identified with an arrow head at the cropped endpoint.

► Example 7: Changing titles and supertitles

Continuing with the BCG dataset from [example 6](#), we demonstrate how to override the default title and supertitle for the `_data` column. The binary data we have correspond to a 2×2 table with cells `_a`, `_b`, `_c`, and `_d`. Cells `_a` and `_b` may be referred to as `_data1`, and cells `_c` and `_d` may be referred to as `_data2`.

We override the supertitle for the `_data1` column to display “Vaccinated” and the titles for each cell to display either a “+” or a “-” as follows.

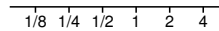
```
. meta forestplot, eform
> columnopts(_data1, supertitle(Vaccinated))
> columnopts(_a _c, title(+)) columnopts(_b _d, title(-))
```



Heterogeneity: $\tau^2 = 0.31$, $I^2 = 92.22\%$, $H^2 = 12.86$

Test of $\theta_1 = \theta_2$: $Q(12) = 152.23$, $p = 0.00$

Test of $\theta = 0$: $z = -3.97$, $p = 0.00$

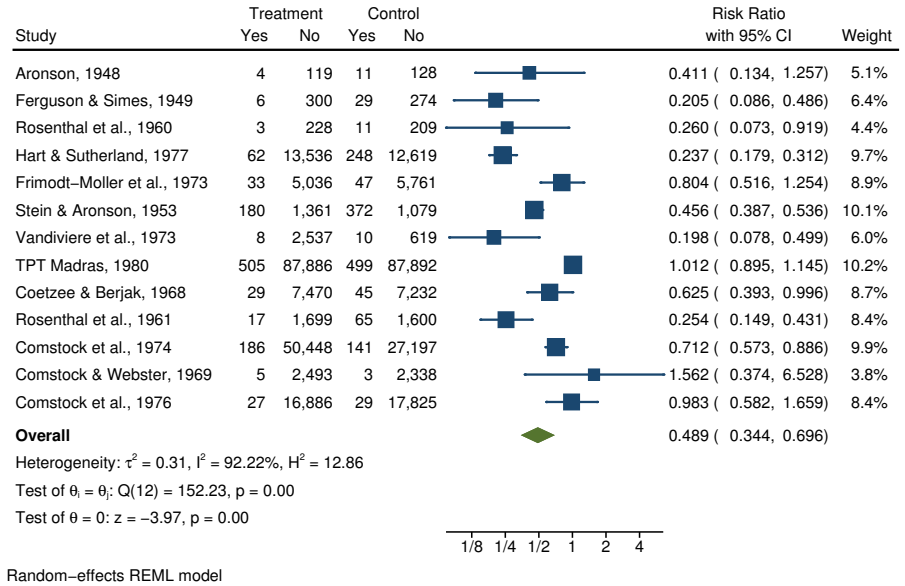


Random-effects REML model

► Example 8: Changing columns' formatting

Following [example 7](#), we now demonstrate how to override the default formats for the `_esci` and `_weight` columns. For the `_esci` column, we also specify that the CIs be displayed inside parentheses instead of the default brackets. For the `_weight` column, we specify that the plotted weights be adorned with a `%` sign and modify the title and supertitle accordingly.

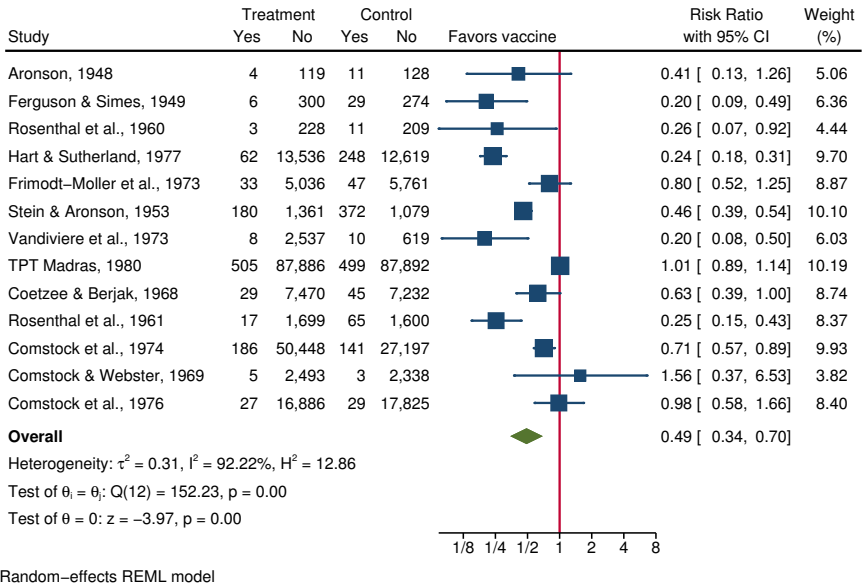
```
. meta forestplot, eform cibind(parentheses)
> columnopts(_esci, format(%6.3f))
> columnopts(_weight, mask(%6.1f%%) title(Weight) supertitle(""))
```



► Example 9: Changing axis range

In this example, we specify the `xscale(range(1/8 8))` and `xlabel(#7)` options to specify that the x -axis range be symmetric (on the risk-ratio scale) about the no-effect value of 1 and that 7 tick marks be shown on the axis.

```
. meta forest, eform xline(1) xscale(range(1/8 8)) xlabel(#7)
> columnopts(_plot, title("Favors vaccine{space 20}"))
```



We also used the `columnopts(_plot, title())` option to specify which side of the plot (with respect to the no-effect line) favors the treatment. In other words, the effect sizes that are falling on that side reported that the treatment (vaccine) reduced the risk of tuberculosis.

◀

Methods and formulas

Methods and formulas for the statistics reported by `meta forestplot` are given in [Methods and formulas](#) of [\[META\] meta summarize](#).

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Also see

- [META] **meta data** — Declare meta-analysis data
- [META] **meta summarize** — Summarize meta-analysis data
- [META] **meta** — Introduction to meta
- [META] **Glossary**
- [META] **Intro** — Introduction to meta-analysis

Title

meta summarize — Summarize meta-analysis data

Description	Quick start	Menu	Syntax
Options	Remarks and examples	Stored results	Methods and formulas
References	Also see		

Description

`meta summarize` summarizes [meta data](#). It reports individual effect sizes and the overall effect size (ES), their confidence intervals (CIs), heterogeneity statistics, and more. `meta summarize` can perform random-effects (RE), common-effect (CE), and fixed-effects (FE) meta-analyses. It can also perform subgroup, cumulative, and sensitivity meta-analyses. For graphical display of meta-analysis summaries, see [\[META\] meta forestplot](#).

Quick start

Perform meta-analysis and summarize meta data, which were declared by either `meta set` or `meta esize`

```
meta summarize
```

As above, but summarize meta-analysis results using the empirical Bayes RE method instead of the declared method

```
meta summarize, random(ebayes)
```

Perform subgroup meta-analysis based on the categorical variable `x1`

```
meta summarize, subgroup(x1)
```

Perform subgroup analysis based on the categorical variables `x1`, `x2`, and `x3`

```
meta summarize, subgroup(x1 x2 x3)
```

Perform cumulative meta-analysis (CMA), where studies are included in the CMA based on the ascending order of observations in variable `x4`

```
meta summarize, cumulative(x4)
```

As above, but stratify the results of the CMA based on groups of the categorical variable `x5`

```
meta summarize, cumulative(x4, by(x5))
```

Perform sensitivity meta-analysis by assuming a fixed value of 0.2 for the between-study heterogeneity parameter τ^2 , assuming that the declared model is RE

```
meta summarize, tau2(.2)
```

Menu

Statistics > Meta-analysis

Syntax

Meta-analysis as declared with meta set or meta esize

```
meta summarize [if] [in] [, options reopts]
```

Random-effects meta-analysis

```
meta summarize [if] [in], random[(remethod)] [options reopts]
```

Common-effect meta-analysis

```
meta summarize [if] [in], common[(cefemethod)] [options]
```

Fixed-effects meta-analysis

```
meta summarize [if] [in], fixed[(cefemethod)] [options]
```

<i>options</i>	Description
----------------	-------------

Main

<code>subgroup(<i>varlist</i>)</code>	subgroup meta-analysis for each variable in <i>varlist</i>
<code>cumulative(<i>cumuluspec</i>)</code>	cumulative meta-analysis

Options

<code>level(#)</code>	set confidence level; default is as declared for meta-analysis
<code>eform_option</code>	report exponentiated results
<code>tdistribution</code>	report <i>t</i> test instead of <i>z</i> test for the overall effect size
<code>nostudies</code>	suppress output for individual studies
<code>noheader</code>	suppress output header
<code>[no]metashow</code>	display or suppress meta settings in the output
<code>display_options</code>	control column formats

Maximization

<code>maximize_options</code>	control the maximization process; seldom used
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<i>remethod</i>	Description
-----------------	-------------

<code>reml</code>	restricted maximum likelihood; the default
<code>mle</code>	maximum likelihood
<code>ebayes</code>	empirical Bayes
<code>dlaird</code>	DerSimonian–Laird
<code>sjonkman</code>	Sidik–Jonkman
<code>hedges</code>	Hedges
<code>hschmidt</code>	Hunter–Schmidt

<i>cefemethod</i>	Description
-------------------	-------------

<code>maenszel</code>	Mantel–Haenszel
<code>invvariance</code>	inverse variance
<code>ivariance</code>	synonym for <code>invvariance</code>

<i>reopts</i>	Description
<code>tau2(#)</code>	sensitivity meta-analysis using a fixed value of between-study variance τ^2
<code>i2(#)</code>	sensitivity meta-analysis using a fixed value of heterogeneity statistic I^2
<code>predinterval [(#)]</code>	report prediction interval for the overall effect size
<code>se(<i>seadj</i>)</code>	adjust standard error of the overall effect size

Options

Main

Options `random()`, `common()`, and `fixed()`, when specified with `meta summarize`, temporarily override the global model declared by `meta set` or `meta esize` during the computation. Options `random()`, `common()`, and `fixed()` may not be combined. If these options are omitted, the declared meta-analysis model is assumed; see [Declaring a meta-analysis model](#) in [META] [meta data](#). Also see [Meta-analysis models](#) in [META] [Intro](#).

`random` and `random(remethod)` specify that a random-effects model be assumed for meta-analysis; see [Random-effects model](#) in [META] [Intro](#).

remethod specifies the type of estimator for the between-study variance τ^2 . *remethod* is one of `reml`, `mle`, `ebayes`, `dlaird`, `sjonkman`, `hedges`, or `hschmidt`. `random` is a synonym for `random(reml)`. See [Options](#) in [META] [meta esize](#) for more information.

`common` and `common(cefemethod)` specify that a common-effect model be assumed for meta-analysis; see [Common-effect \(“fixed-effect”\) model](#) in [META] [Intro](#). Also see the [discussion](#) in [META] [meta data](#) about common-effect versus fixed-effects models.

`common` implies `common(mhaenszel)` for effect sizes `lnoratio`, `lnrratio`, and `rdiff` and `common(invvariance)` for all other effect sizes. `common(mhaenszel)` is supported only with effect sizes `lnoratio`, `lnrratio`, and `rdiff`.

cefemethod is one of `mhaenszel` or `invvariance` (synonym `ivariance`). See [Options](#) in [META] [meta esize](#) for more information.

`fixed` and `fixed(cefemethod)` specify that a fixed-effects model be assumed for meta-analysis; see [Fixed-effects model](#) in [META] [Intro](#). Also see the [discussion](#) in [META] [meta data](#) about fixed-effects versus common-effect models.

`fixed` implies `fixed(mhaenszel)` for effect sizes `lnoratio`, `lnrratio`, and `rdiff` and `fixed(invvariance)` for all other effect sizes. `fixed(mhaenszel)` is supported only with effect sizes `lnoratio`, `lnrratio`, and `rdiff`.

cefemethod is one of `mhaenszel` or `invvariance` (synonym `ivariance`); see [Options](#) in [META] [meta esize](#) for more information.

`subgroup(varlist)` specifies that a subgroup meta-analysis (subgroup analysis) be performed for each variable in *varlist*. Subgroup analysis performs meta-analysis separately for each variable in *varlist* and for each group as defined by that variable. The specified meta-analysis model is assumed for each subgroup. This analysis is useful when the results of all studies are too heterogeneous to be combined into one estimate but the results are similar within certain groups of studies. The specified variables can be numeric or string variables. When multiple variables are specified, only the subgroup results are displayed; that is, the results from individual studies are suppressed for brevity. Only one of `subgroup()` or `cumulative()` may be specified.

`cumulative(ordervar [, ascending|descending by(byvar)])` performs a cumulative meta-analysis (CMA). CMA performs multiple meta-analyses and accumulates the results by adding one study at a time to each subsequent analysis. It is useful for monitoring the results of the studies as new studies become available. The studies enter the CMA based on the ordered values of variable *ordervar*. *ordervar* must be a numeric variable. By default, ascending order is assumed unless the suboption `descending` is specified; only one of `ascending` or `descending` is allowed. The `by(byvar)` option specifies that the CMA be stratified by variable *byvar*.

reopts are `tau2(#)`, `i2(#)`, `predinterval[(#)]`, and `se(khartung[, truncated])`. These options are used with random-effects meta-analysis.

`tau2(#)` specifies the value of the between-study variance parameter, τ^2 , to use for the random-effects meta-analysis. This option is useful for exploring the sensitivity of the results to different levels of between-study heterogeneity. Only one of `tau2()` or `i2()` may be specified. This option is not allowed in combination with `subgroup()` or `cumulative()`.

`i2(#)` specifies the value of the heterogeneity statistic I^2 (as a percentage) to use for the random-effects meta-analysis. This option is useful for exploring the sensitivity of the results to different levels of between-study heterogeneity. Only one of `i2()` or `tau2()` may be specified. This option is not allowed in combination with `subgroup()` or `cumulative()`.

`predinterval` and `predinterval(#)` specify that the 95% or #% **prediction interval** be reported for the overall effect size in addition to the confidence interval. # specifies the confidence level of the prediction interval. The prediction interval provides plausible ranges for the effect size in a future, new study.

`se(seadj)` specifies that the adjustment *seadj* be applied to the standard error of the overall effect size. Additionally, the test of significance of the overall effect size is based on a Student's *t* distribution instead of the normal distribution.

seadj is `khartung[, truncated]`. Adjustment `khartung` specifies that the Knapp–Hartung adjustment (Hartung and Knapp 2001a, 2001b; Knapp and Hartung 2003), also known as the Sidik–Jonkman adjustment (Sidik and Jonkman 2002), be applied to the standard error of the overall effect size. `hknapp` and `sjonkman` are synonyms for `khartung`. `truncated` specifies that the truncated Knapp–Hartung adjustment (Knapp and Hartung 2003), also known as the modified Knapp–Hartung adjustment, be used.

Options

`level(#)` specifies the confidence level, as a percentage, for confidence intervals. The default is as declared for the meta-analysis session; see [Declaring a confidence level for meta-analysis in \[META\] meta data](#). Also see option `level()` in [\[META\] meta set](#).

eform_option is one of `eform`, `eform(string)`, `or`, or `rr`. It reports exponentiated effect sizes and transforms their respective confidence intervals, whenever applicable. By default, the results are displayed in the metric declared with `meta set` or `meta esize` such as log odds-ratios and log risk-ratios. *eform_option* affects how results are displayed, not how they are estimated and stored.

`eform(string)` labels the exponentiated effect sizes as *string*; the other options use default labels.

The default label is specific to the chosen effect size. For example, option `eform` uses Odds Ratio when used with log odds-ratios declared with `meta esize` or Risk Ratio when used with the declared log risk-ratios. Option `or` is a synonym for `eform` when log odds-ratio is declared, and option `rr` is a synonym for `eform` when log risk-ratio is declared. If option `eslabel(eslab)` is specified during declaration, then `eform` will use the `exp(eslab)` label or, if `eslab` is too long, the `exp(ES)` label.

`tdistribution` reports a t test instead of a z test for the overall effect size. This option may not be combined with option `subgroup()`, `cumulative()`, or `se()`.

`nostudies` (synonym `nostudy`) suppresses the display of information such as effect sizes and their CIs for individual studies from the output table.

`noheader` suppresses the output header.

`metashow` and `nometashow` display or suppress the meta setting information. By default, this information is displayed at the top of the output. You can also specify `nometashow` with `meta update` to suppress the meta setting output for the entire meta-analysis session.

`display_options`: `cformat(%fmt)`, `pformat(%fmt)`, and `sformat(%fmt)`; see [R] [Estimation options](#). The defaults are `cformat(%9.3f)`, `pformat(%5.3f)`, and `sformat(%8.2f)`.

`wgtformat(%fmt)` specifies how to format the weight column in the output table. The default is `wgtformat(%5.2f)`. The maximum format width is 5.

`ordformat(%fmt)` specifies the format for the values of the order variable, specified in `cumulative(ordervar)`. The default is `ordformat(%9.0g)`. The maximum format width is 9.

Maximization

`maximize_options`: `iterate(#)`, `tolerance(#)`, `nrtolerance(#)`, `nonrtolerance` (see [R] [Maximize](#)), `from(#)`, and `showtrace`. These options control the iterative estimation of the between-study variance parameter, τ^2 , with random-effects methods `reml`, `mle`, and `ebayes`. These options are seldom used.

`from(#)` specifies the initial value for τ^2 during estimation. By default, the initial value for τ^2 is the noniterative Hedges estimator.

`showtrace` displays the iteration log that contains the estimated parameter τ^2 , its relative difference with the value from the previous iteration, and the scaled gradient.

Remarks and examples

Remarks are presented under the following headings:

[Introduction](#)

[Examples of using meta summarize](#)

Introduction

Meta-analysis helps answer research questions based on the results of multiple studies. Does exercise prolong life? Does lack of sleep increase the risk of cancer? Does daylight saving save energy? Or does performing the duck-face technique while taking a selfie increase the number of likes on Facebook? These (except perhaps the last one) and many other research questions have been investigated by multiple studies. These studies may have reported conflicting results: some may have shown effects in one direction, some in the opposite, and others may have shown none that are statistically significant. Meta-analysis uses quantitative methods to explore these conflicting results and, whenever possible, provide a unified conclusion based on the results of the individual studies.

Meta-analysis combines the results of similar multiple studies into a single result. Studies typically report some measures of outcomes, or effect sizes, and their precision (standard errors or CIs). Meta-analysis combines the individual effects sizes to provide various meta-analysis summaries. The main summaries are the overall effect size and its precision. Other meta-analysis summaries include the test of significance of the overall effect size, between-study heterogeneity summaries such as the

I^2 statistic, and the test of homogeneity between studies. The `meta summarize` command reports such summaries.

Estimating the overall effect size, θ , and its precision based on the results of multiple studies is at the heart of meta-analysis. There are various methods for estimating θ , which depend on the research goals and model assumptions about the studies. The estimate of the overall (combined) ES is computed as the weighted average of the study-specific effect sizes, with larger weights given to more precise (larger) studies:

$$\hat{\theta} = \frac{\sum_{j=1}^K w_j \hat{\theta}_j}{\sum_{j=1}^K w_j}$$

The weights are determined by the chosen meta-analysis model, estimation method, and potentially the type of effect size; see [Methods and formulas](#) for details. (In [\[META\] Intro](#), we used θ_{pop} to denote the population parameter of interest. For simplicity, here and in the rest of the documentation, we will use θ .)

As we described in [Meta-analysis models](#) in [\[META\] Intro](#), the choice of a meta-analysis model is important not only for estimation but also for interpretation of $\hat{\theta}$. `meta summarize` supports random-effects (`random`), fixed-effects (`fixed`), and common-effect (`common`) meta-analysis models. Each meta-analysis model provides various estimation methods such as the random-effects REML method, `random(reml)`, and fixed-effects Mantel–Haenszel method, `fixed(mhaenszel)`. The default model and method are as declared with `meta set` or `meta esize`; see [Declaring a meta-analysis model](#) in [\[META\] meta data](#). Note that the Mantel–Haenszel method is available only with effect sizes `lnratio`, `lnrratio`, and `rdiff` declared by using `meta esize`; see [\[META\] meta esize](#).

For random-effects models, you can perform sensitivity meta-analysis to explore the impact of different levels of heterogeneity on the results. You can use the `tau2(#)` option to specify different fixed values for the between-study variance τ^2 . Or you can fix the percentage of variation in the effect sizes because of heterogeneity by specifying the values for the I^2 statistic in the `i2(#)` option. With random-effects models, you can also compute prediction intervals for $\hat{\theta}$, `predinterval(#)`, and use the alternative standard-error estimators, `se()`.

You can perform subgroup analysis, `subgroup()`, or CMA, `cumulative()`; see [Subgroup meta-analysis](#) and [Cumulative meta-analysis](#) in [\[META\] Intro](#). Also see [Subgroup meta-analysis](#) and [Cumulative meta-analysis](#) in [Methods and formulas](#) below.

You can specify the desired confidence level with `level()`; report exponentiated results by specifying `eform`; report a t test, instead of a z test, for the overall effect size by specifying `tdistribution`; and more.

In the next section, we demonstrate various usages of `meta summarize`.

Examples of using meta summarize

Recall the pupil IQ data (Raudenbush and Bryk 1985; Raudenbush 1984) described in *Effects of teacher expectancy on pupil IQ (pupiliq.dta)* of [META] **meta**. Here we will use its declared version and will focus on the demonstration of various options of `meta summarize` and explanation of its output.

```
. use https://www.stata-press.com/data/r16/pupiliqset
(Effects of teacher expectancy on pupil IQ; set with -meta set-)
. keep in 1/10
(9 observations deleted)
. meta query, short
-> meta set stdmdiff se , studylabel(studylbl) eslabel(Std. Mean Diff.)
Effect-size label: Std. Mean Diff.
Effect-size type: Generic
Effect size: stdmdiff
Std. Err.: se
Model: Random-effects
Method: REML
```

For brevity, we consider only the first 10 studies. We use `meta query, short` to remind us about the main settings of the declaration step. Our data were declared by using `meta set` with variables `stdmdiff` and `se` specifying the effect sizes and their standard errors, respectively. The declared meta-analysis model is the default random-effects model with the REML estimation method.

Examples are presented under the following headings:

- Example 1: Default random-effects meta-analysis*
- Example 2: DerSimonian–Laird random-effects method*
- Example 3: Fixed-effects meta-analysis*
- Example 4: Common-effect meta-analysis*
- Example 5: Knapp–Hartung standard-error adjustment*
- Example 6: Prediction interval*
- Example 7: Sensitivity meta-analysis*
- Example 8: Other options: CI level, t distribution, eform*
- Example 9: Subgroup meta-analysis*
- Example 10: Cumulative meta-analysis*

► Example 1: Default random-effects meta-analysis

We type `meta summarize` to obtain a standard meta-analysis summary.

```
. meta summarize
      Effect-size label: Std. Mean Diff.
      Effect size: stdmdiff
      Std. Err.: se
      Study label: studylbl

Meta-analysis summary                                Number of studies =    10
Random-effects model                                Heterogeneity:
Method: REML                                         tau2 = 0.0754
                                                    I2 (%) = 74.98
                                                    H2 = 4.00
```

Effect Size: Std. Mean Diff.

Study	Effect Size	[95% Conf. Interval]		% Weight
Rosenthal et al., 1974	0.030	-0.215	0.275	12.39
Conn et al., 1968	0.120	-0.168	0.408	11.62
Jose & Cody, 1971	-0.140	-0.467	0.187	10.92
Pellegrini & Hicks, 1972	1.180	0.449	1.911	5.25
Pellegrini & Hicks, 1972	0.260	-0.463	0.983	5.33
Evans & Rosenthal, 1969	-0.060	-0.262	0.142	13.11
Fielder et al., 1971	-0.020	-0.222	0.182	13.11
Claiborn, 1969	-0.320	-0.751	0.111	9.11
Kester, 1969	0.270	-0.051	0.591	11.02
Maxwell, 1970	0.800	0.308	1.292	8.15
theta	0.134	-0.075	0.342	

Test of theta = 0: z = 1.26

Prob > |z| = 0.2085

Test of homogeneity: Q = chi2(9) = 26.21

Prob > Q = 0.0019

As with other `meta` commands, a short information about meta settings is displayed directly following the `meta summarize` command. It can be suppressed with the `nometashow` option; see [example 2](#).

Next, the header reports the information about the meta-analysis model and method, the number of studies (10), and several heterogeneity statistics. The output table reports the effect sizes and their 95% CIs for individual studies and the estimate of the overall, combined ES, labeled as `theta`, and its 95% CI. The test of significance of the overall effect size and the homogeneity test are reported at the bottom of the table.

Because our declared effect-size label, `Std. Mean Diff.`, was too long to fit as the column header, `meta summarize` used the generic column label `Effect Size` but displayed the specified label in the table legend.

The mean effect size in our example is 0.134 with the 95% CI of $[-0.075, 0.342]$. This estimate is computed as the weighted average of the study-specific effect sizes, with the weights representing precision of the studies. The percentages of the total weight for each study are reported in the `% Weight` column. The more precise the study is, the larger its weight percentage. For example, studies 6 and 7, with labels `Evans & Rosenthal, 1969` and `Fielder et al., 1971`, have the largest weight percentage among the studies of about 13% (each). Thus, their effect-size estimates, -0.06 and -0.02 , have the largest weights in the weighted-average estimate.

The 95% CI for the overall mean estimate and the test of $H_0: \theta = 0$ with the z -test statistic of 1.26 and the p -value of 0.2085 suggest that θ is not statistically significantly different from 0. We should be careful, however, with our conclusions in the presence of between-study heterogeneity.

The heterogeneity statistic I^2 , reported in the header, is about 75%, which means that 75% of the variability in the effect-size estimates is because of the between-study differences rather than the sampling variation. According to Higgins et al. (2003), this value of I^2 corresponds to “large heterogeneity”. (The authors suggest that $I^2 = 25\%$ should indicate “small heterogeneity”, $I^2 = 50\%$ indicate “medium heterogeneity”, and $I^2 = 75\%$ indicate “large heterogeneity”.) The between-study variance τ^2 is estimated to be 0.0754. The homogeneity test of $H_0: \theta_1 = \theta_2 = \dots = \theta_{10}$ reports the Q test statistic of 26.21 with a p -value of 0.0019.

When there are few studies, which is typical in meta-analysis, the homogeneity test is known to have low power, which means that it may not detect clinically significant heterogeneity (Hedges and Pigott 2001). Thus, you should use caution when interpreting nonsignificant results as “no heterogeneity”. In fact, many experts (for example, Berman and Parker [2002]) recommend using a 10% significance level instead of the classical 5% level to determine statistical significance when using this test. On the other hand, when there are many studies, this test is known to have excessive power, which means that it tends to detect heterogeneity that is clinically insignificant (Hardy and Thompson 1998).

In our example, the p -value of the homogeneity test is $0.0019 < 0.05 < 0.1$, so there is definitely statistical evidence of the between-study heterogeneity. See example 9 for one way to account for the heterogeneity.

◀

► Example 2: DerSimonian–Laird random-effects method

Continuing with example 1, let’s use the DerSimonian–Laird random-effects method instead of the default (declared) REML method. Let’s also suppress the meta setting information displayed at the top of the command output by using the `nometashow` option.

```
. meta summarize, random(dlaird) nometashow
Meta-analysis summary          Number of studies =    10
Random-effects model          Heterogeneity:
Method: DerSimonian-Laird      tau2 =    0.0481
                                I2 (%) =    65.66
                                H2 =     2.91
```

Effect Size: Std. Mean Diff.

Study	Effect Size	[95% Conf. Interval]		% Weight
Rosenthal et al., 1974	0.030	-0.215	0.275	13.00
Conn et al., 1968	0.120	-0.168	0.408	11.88
Jose & Cody, 1971	-0.140	-0.467	0.187	10.90
Pellegrini & Hicks, 1972	1.180	0.449	1.911	4.42
Pellegrini & Hicks, 1972	0.260	-0.463	0.983	4.49
Evans & Rosenthal, 1969	-0.060	-0.262	0.142	14.11
Fielder et al., 1971	-0.020	-0.222	0.182	14.11
Claiborn, 1969	-0.320	-0.751	0.111	8.58
Kester, 1969	0.270	-0.051	0.591	11.04
Maxwell, 1970	0.800	0.308	1.292	7.45
theta	0.117	-0.061	0.296	

Test of theta = 0: z = 1.29

Prob > |z| = 0.1967

Test of homogeneity: Q = chi2(9) = 26.21

Prob > Q = 0.0019

The results are now based on the DerSimonian–Laird method, and the header is updated to reflect this. This method is one of the many [random-effects methods](#) for estimating the between-study variance

τ^2 . Its estimate is 0.0481. In random-effects models, the weights depend on τ^2 and thus will differ across different random-effects methods. The mean effect-size estimate under the DerSimonian–Laird method is 0.117 with the 95% CI of $[-0.061, 0.296]$. This estimate is similar to the 0.134 estimate we obtained in [example 1](#). We also arrive at the same inferential conclusion of no statistical significance of the mean effect size as in the previous example.

To shorten the output, let's suppress the meta setting information from the output of `meta summarize` for all remaining examples. We can use `meta update` to update our current meta settings.

```
. quietly meta update, nometashow
```

We specified the `nometashow` option with `meta update` to suppress the display of the meta setting information in all meta commands; see [Modifying default meta settings](#) in [\[META\] meta data](#). ◀

► Example 3: Fixed-effects meta-analysis

In [example 1](#), we assumed a random-effects meta-analysis model. We can use the `fixed` option to specify a fixed-effects meta-analysis model.

```
. meta summarize, fixed
Meta-analysis summary                Number of studies =    10
Fixed-effects model                  Heterogeneity:
Method: Inverse-variance            I2 (%) =    65.66
                                      H2 =    2.91
```

Effect Size: Std. Mean Diff.

Study	Effect Size	[95% Conf. Interval]		% Weight
Rosenthal et al., 1974	0.030	-0.215	0.275	15.13
Conn et al., 1968	0.120	-0.168	0.408	10.94
Jose & Cody, 1971	-0.140	-0.467	0.187	8.48
Pellegrini & Hicks, 1972	1.180	0.449	1.911	1.70
Pellegrini & Hicks, 1972	0.260	-0.463	0.983	1.74
Evans & Rosenthal, 1969	-0.060	-0.262	0.142	22.29
Fielder et al., 1971	-0.020	-0.222	0.182	22.29
Claiborn, 1969	-0.320	-0.751	0.111	4.89
Kester, 1969	0.270	-0.051	0.591	8.79
Maxwell, 1970	0.800	0.308	1.292	3.75
theta	0.051	-0.045	0.146	

Test of theta = 0: z = 1.04

Prob > |z| = 0.2974

Test of homogeneity: Q = chi2(9) = 26.21

Prob > Q = 0.0019

As reported in the header, `fixed` implied the inverse-variance estimation method. The between-group variance parameter is not estimated with fixed-effects models, so the heterogeneity summary does not report `tau2`. Under this model, the mean effect-size estimate is 0.051 with the 95% CI of $[-0.045, 0.146]$. As we explain in [Comparison between the models and interpretation of their results](#) in [\[META\] Intro](#), in a fixed-effects model, `theta` estimates the weighted average of the true study-specific standardized mean differences. Our interpretation is also limited to these 10 studies that we observed in our meta-analysis. That is, the weighted average of the standardized mean differences of these 10 studies is not statistically significantly different from 0. ◀

► Example 4: Common-effect meta-analysis

From [example 1](#) and [example 3](#), we determined that there is substantial between-study variability in these data. Thus, a common-effect model, which assumes that all study-specific effects are the same, is not reasonable for these data. But we will demonstrate it for illustration purposes.

```
. meta summarize, common
Meta-analysis summary           Number of studies =    10
Common-effect model
Method: Inverse-variance
      Effect Size: Std. Mean Diff.
```

Study	Effect Size	[95% Conf. Interval]		% Weight
Rosenthal et al., 1974	0.030	-0.215	0.275	15.13
Conn et al., 1968	0.120	-0.168	0.408	10.94
Jose & Cody, 1971	-0.140	-0.467	0.187	8.48
Pellegrini & Hicks, 1972	1.180	0.449	1.911	1.70
Pellegrini & Hicks, 1972	0.260	-0.463	0.983	1.74
Evans & Rosenthal, 1969	-0.060	-0.262	0.142	22.29
Fielder et al., 1971	-0.020	-0.222	0.182	22.29
Claiborn, 1969	-0.320	-0.751	0.111	4.89
Kester, 1969	0.270	-0.051	0.591	8.79
Maxwell, 1970	0.800	0.308	1.292	3.75
theta	0.051	-0.045	0.146	

Test of theta = 0: z = 1.04

Prob > |z| = 0.2974

We use the `common` option to specify a common-effect model. Because this model implies no heterogeneity, the corresponding summaries and the homogeneity test are not reported for this model. As we point out in [Comparison between the models and interpretation of their results](#) in [\[META\] Intro](#), a common-effect model is computationally the same as a fixed-effects model. So we obtain the exact same results as in [example 3](#). However, the interpretation of our results is different. Here `theta` estimates a single effect, which is common to all studies. Although the two models produce the same results, to encourage proper interpretation, we provide both options, `common` and `fixed`, to distinguish between these models; see [Declaring a meta-analysis model](#) in [\[META\] meta data](#) for details.

◀

► Example 5: Knapp–Hartung standard-error adjustment

Let's return to our random-effects model from [example 1](#). For random-effects models, `meta summarize` provides several additional options, which we explore in the next three examples.

The Knapp–Hartung adjustment (also known as the Sidik–Jonkman adjustment) to the standard error of the overall effect size ([Knapp and Hartung 2003](#) and [Hartung and Knapp 2001a, 2001b](#)) is sometimes used in practice. We can specify it with the `se(khartung)` option. We also specify the `nostudies` option to suppress the output from individual studies because it is unaffected by the `se(khartung)` option.

```
. meta summarize, se(khartung) nostudies
```

Meta-analysis summary	Number of studies = 10
Random-effects model	Heterogeneity:
Method: REML	tau2 = 0.0754
SE adjustment: Knapp-Hartung	I2 (%) = 74.98
	H2 = 4.00

theta: Overall Std. Mean Diff.

	Estimate	Std. Err.	t	P> t	[95% Conf. Interval]
theta	.1335309	.1215065	1.10	0.300	-.1413358 .4083976

Test of homogeneity: Q = chi2(9) = 26.21 Prob > Q = 0.0019

Without the individual studies, the output table is slightly different. The test of significance is now reported in the output table instead of at the bottom of the output table.

The estimate `theta` is the same as in [example 1](#), 0.134, but it is reported with more digits in this table. The confidence intervals and the test of significance are different. In addition to making an adjustment to the standard error, Knapp and Hartung also use a Student's t distribution as a sampling distribution instead of the normal distribution. Thus, the t statistic is reported in the output table instead of the z statistic. Regardless, we still conclude that our overall effect size is not statistically significant.

Another standard-error adjustment, also used in practice, is the so-called truncated or modified Knapp–Hartung adjustment; see [Methods and formulas](#) for details. This adjustment can be specified with the `se(khartung, truncated)` option.

```
. meta summarize, se(khartung, truncated)
(output omitted)
```

◀

▷ Example 6: Prediction interval

Recall from [Random-effects model](#) in [\[META\] Intro](#) that a random-effects model implies that the observed studies in a meta-analysis represent a sample from a larger population of similar studies. What if we want to estimate the plausible ranges for the overall effect size in a new, future study? We cannot use the confidence interval for the overall effect size because it does not incorporate the uncertainty in estimating the between-study variance, which is important if we want to predict an effect in a new study. We can compute the prediction interval.

```
. meta summarize, predinterval(90) nostudies
Meta-analysis summary                    Number of studies =    10
Random-effects model                    Heterogeneity:
Method: REML                            tau2 = 0.0754
                                           I2 (%) = 74.98
                                           H2 = 4.00
```

theta: Overall Std. Mean Diff.

	Estimate	Std. Err.	z	P> z	[95% Conf. Interval]
theta	.1335309	.1061617	1.26	0.208	-.0745422 .3416041

90% prediction interval for theta: [-0.414, 0.681]

Test of homogeneity: Q = chi2(9) = 26.21 Prob > Q = 0.0019

We specified `predinterval(90)` to compute the 90% prediction interval for the mean effect size; use `predinterval` to compute the 95% interval. Following [example 5](#), we also used `nostudies` to suppress individual studies.

The 90% prediction interval, reported at the bottom of the table, is [-0.414, 0.681]. The prediction interval will be wider than the confidence interval because it additionally accounts for the uncertainty in the between-study variability.

◀

▶ Example 7: Sensitivity meta-analysis

For random-effects models, we can perform sensitivity analysis to explore various levels of heterogeneity between studies. Let's see how our results change for different values of the between-study variance τ^2 and the heterogeneity statistic I^2 .

Let's compute the results assuming that τ^2 equals 0.25.

```
. meta summarize, tau2(0.25) nostudies
Sensitivity meta-analysis summary        Number of studies =    10
Random-effects model                    Heterogeneity:
Method: User-specified tau2            tau2 = 0.2500
                                           I2 (%) = 90.86
                                           H2 = 10.94
```

theta: Overall Std. Mean Diff.

	Estimate	Std. Err.	z	P> z	[95% Conf. Interval]
theta	.173588	.171407	1.01	0.311	-.1623636 .5095395

Test of homogeneity: Q = chi2(9) = 26.21

Prob > Q = 0.0019

Our estimate of the mean effect size is 0.174 with the 95% CI of [-0.162, 0.51] compared with 0.134 with the 95% CI of [-0.075, 0.342] from [example 1](#).

The specified value of τ^2 corresponds to an I^2 of about 91%. Let's now compute the results assuming I^2 of 10%.

```
. meta summarize, i2(10) nostudies
Sensitivity meta-analysis summary          Number of studies =    10
Random-effects model                      Heterogeneity:
Method: User-specified I2                 tau2 =    0.0028
                                           I2 (%) =   10.00
                                           H2 =     1.11
```

theta: Overall Std. Mean Diff.

	Estimate	Std. Err.	z	P> z	[95% Conf. Interval]
theta	.0589369	.0527232	1.12	0.264	-.0443987 .1622724

Test of homogeneity: Q = chi2(9) = 26.21 Prob > Q = 0.0019

The estimate of the mean effect size is 0.059 with the 95% CI of $[-0.044, 0.162]$. The corresponding τ^2 value is 0.0028.

In both cases above, the mean effect size is not statistically significant.

◀

► Example 8: Other options: CI level, t distribution, eform

`meta summarize` provides other options such as `level()` to temporarily change the declared confidence level and `tdistribution` to use a Student's t distribution as the sampling distribution instead of the default normal distribution.

```
. meta summarize, level(90) tdistribution
Meta-analysis summary          Number of studies =    10
Random-effects model          Heterogeneity:
Method: REML                  tau2 =    0.0754
                               I2 (%) =   74.98
                               H2 =     4.00
```

Effect Size: Std. Mean Diff.

Study	Effect Size	[90% Conf. Interval]	% Weight
Rosenthal et al., 1974	0.030	-0.176 0.236	12.39
Conn et al., 1968	0.120	-0.122 0.362	11.62
Jose & Cody, 1971	-0.140	-0.415 0.135	10.92
Pellegrini & Hicks, 1972	1.180	0.566 1.794	5.25
Pellegrini & Hicks, 1972	0.260	-0.347 0.867	5.33
Evans & Rosenthal, 1969	-0.060	-0.229 0.109	13.11
Fielder et al., 1971	-0.020	-0.189 0.149	13.11
Claiborn, 1969	-0.320	-0.682 0.042	9.11
Kester, 1969	0.270	0.000 0.540	11.02
Maxwell, 1970	0.800	0.387 1.213	8.15
theta	0.134	-0.061 0.328	

Test of theta = 0: t(9) = 1.26 Prob > |t| = 0.2085
 Test of homogeneity: Q = chi2(9) = 26.21 Prob > Q = 0.0019

Notice that all CIs, including those for the individual studies, now correspond to the 90% confidence level, compared with [example 1](#). Also, the significance test now uses the Student's t distribution with 9 degrees of freedom, but the conclusion remains the same—the mean effect size is not statistically significant.

You may also find `meta summarize`'s option `eform` useful when dealing with the effect sizes in the log-transformed metric such as log odds-ratios or log risk-ratios. By default, `meta summarize` reports results in the declared metric, which should be chosen such that the sampling distributions of the effect sizes are well approximated by normal distributions. It may be more convenient, however, to display the final results in the original metric. When you specify the `eform` option, it reports the exponentiated results and the corresponding CIs. Note that the significance tests and other summary measures are still computed based on the nonexponentiated results.

It does not make sense to exponentiate standardized mean differences in our example, but we will do this just to demonstrate the option.

```
. meta summarize, eform
Meta-analysis summary          Number of studies =    10
Random-effects model          Heterogeneity:
Method: REML                   tau2 =    0.0754
                                I2 (%) =    74.98
                                H2 =     4.00
```

exp(ES): exp(Std. Mean Diff.)

Study	exp(ES)	[95% Conf. Interval]		% Weight
Rosenthal et al., 1974	1.030	0.807	1.317	12.39
Conn et al., 1968	1.127	0.845	1.504	11.62
Jose & Cody, 1971	0.869	0.627	1.206	10.92
Pellegrini & Hicks, 1972	3.254	1.567	6.760	5.25
Pellegrini & Hicks, 1972	1.297	0.629	2.673	5.33
Evans & Rosenthal, 1969	0.942	0.770	1.152	13.11
Fielder et al., 1971	0.980	0.801	1.199	13.11
Claiborn, 1969	0.726	0.472	1.118	9.11
Kester, 1969	1.310	0.950	1.807	11.02
Maxwell, 1970	2.226	1.361	3.640	8.15
exp(theta)	1.143	0.928	1.407	

```
Test of theta = 0: z = 1.26          Prob > |z| = 0.2085
Test of homogeneity: Q = chi2(9) = 26.21      Prob > Q = 0.0019
```

`meta summarize, eform` reports exponentiated effect sizes and their corresponding CIs. It labels the effect-size column as `exp(ES)`, but you can change this label to `string` by specifying `eform(string)`.

Note that the `eform` option worked in our example because `meta set` declared our precomputed effect sizes as generic. They could have been log odds-ratios, in which case `eform` would make perfect sense. However, if you use `meta esize` to compute the standardized mean differences (for example, Hedges's g) and try to use `eform` with `meta summarize`, you will receive an error message because `meta summarize` knows that exponentiation is not appropriate with effect sizes that correspond to continuous data. With effect sizes `lnratio` (or `lnorpeto`) and `lnrratio` computed by `meta esize`, you can also use the respective options `or` and `rr`, which are synonyms for `eform` in those cases. These options (and `eform`) will label your results as Odds Ratio (Peto's OR) and Risk Ratio.

◀

► Example 9: Subgroup meta-analysis

In [example 1](#) and [example 3](#), we identified the presence of substantial heterogeneity between the observed studies. Sometimes, the heterogeneity can be explained by some study-level covariates, also known as moderators. With categorical moderators, we can perform subgroup analysis, which performs meta-analysis separately for each category of each moderator.

We have binary variable `week1`, which records whether teachers had prior contact with students for more than 1 week or for 1 week or less. Let's use this variable as the moderator in our subgroup analysis. We specify the variable `week1` in the `subgroup()` option.

```
. meta summarize, subgroup(week1)
Subgroup meta-analysis summary          Number of studies =    10
Random-effects model
Method: REML
Group: week1
```

Effect Size: Std. Mean Diff.

Study	Effect Size	[95% Conf. Interval]		% Weight
Group: <= 1 week				
Pellegrini & Hicks, 1972	1.180	0.449	1.911	5.25
Pellegrini & Hicks, 1972	0.260	-0.463	0.983	5.33
Kester, 1969	0.270	-0.051	0.591	11.02
Maxwell, 1970	0.800	0.308	1.292	8.15
theta	0.581	0.174	0.989	
Group: > 1 week				
Rosenthal et al., 1974	0.030	-0.215	0.275	12.39
Conn et al., 1968	0.120	-0.168	0.408	11.62
Jose & Cody, 1971	-0.140	-0.467	0.187	10.92
Evans & Rosenthal, 1969	-0.060	-0.262	0.142	13.11
Fielder et al., 1971	-0.020	-0.222	0.182	13.11
Claiborn, 1969	-0.320	-0.751	0.111	9.11
theta	-0.033	-0.137	0.071	
Overall				
theta	0.134	-0.075	0.342	

Heterogeneity summary

Group	df	Q	P > Q	tau2	% I2	H2
<= 1 week	3	7.14	0.068	0.095	57.03	2.33
> 1 week	5	3.53	0.618	0.000	0.00	1.00
Overall	9	26.21	0.002	0.075	74.98	4.00

Test of group differences: $Q_b = \text{chi2}(1) = 8.18$

Prob > $Q_b = 0.004$

We now have two output tables. Our main table now reports results from individual studies separately for each group, in addition to the group-specific overall effect size. The overall effect size computed using all studies is reported at the bottom under `Overall`.

The second table reports the group-specific and overall heterogeneity summaries. The test of group differences is reported at the bottom of this table.

The estimated `theta` for the group with contact `<= 1 week` is 0.581 with the 95% CI of [0.174, 0.989]. The mean effect size in this group is statistically significant at the 5% level. The estimated `theta` for the group with contact `> 1 week` is -0.033 with the 95% CI of [-0.137, 0.071]. The mean effect size in this group is not statistically significant at the 5% level.

If we look at the heterogeneity summaries, the `<= 1 week` group still has some unexplained between-study heterogeneity with an estimated I^2 of 57% and a p -value of the homogeneity test of $0.068 < 0.1$. There does not appear to be any between-study heterogeneity in the `> 1 week` group: I^2 is essentially 0%, and the homogeneity test p -value is 0.618.

We should interpret our results with caution because each subgroup analysis used a few studies, with the ≤ 1 week group having only 4 studies.

We can specify multiple variables in the `subgroup()` option. Let's also include variable `tester` in our subgroup analysis.

```
. meta summarize, subgroup(week1 tester)
Subgroup meta-analysis summary                Number of studies =    10
Random-effects model
Method: REML
Group: week1 tester
```

Group		No. of Studies	Std. Mean Diff.	[95% Conf. Interval]		P-value
week1						
	≤ 1 week	4	0.581	0.174	0.989	0.005
	> 1 week	6	-0.033	-0.137	0.071	0.535
tester						
	aware	7	0.059	-0.129	0.247	0.535
	blind	3	0.316	-0.206	0.837	0.235
Overall						
	theta	10	0.134	-0.075	0.342	0.208

Heterogeneity summary

Group		df	Q	P > Q	tau2	% I2	H2
week1							
	≤ 1 week	3	7.14	0.068	0.095	57.03	2.33
	> 1 week	5	3.53	0.618	0.000	0.00	1.00
tester							
	aware	6	16.35	0.012	0.035	59.07	2.44
	blind	2	9.31	0.009	0.154	75.14	4.02
Overall							
		9	26.21	0.002	0.075	74.98	4.00

Tests of group differences

		df	Q_b	P > Q_b
week1				
	tester	1	8.18	0.004
tester				
		1	0.82	0.365

With more than one variable in `subgroup()`, `meta summarize` reports three output tables. To conserve space, the main table does not report individual studies but reports the number of studies in each group. It also reports the p -values of the corresponding significance tests of the overall effect sizes in each group.

The heterogeneity table reports the group summaries for each variable, in addition to the overall summaries. The new table reports the results of tests of subgroup differences for each variable.

The studies appear to be homogeneous across the levels of the `tester` variable.

► Example 10: Cumulative meta-analysis

CMA (Lau et al. 1992; Sterne 2016) performs multiple meta-analyses by accumulating studies one at a time. The studies are first ordered with respect to the variable of interest, the ordering variable. Meta-analysis summaries are then computed for the first study, for the first two studies, for the first three studies, and so on. The last meta-analysis will correspond to the standard meta-analysis using all studies.

CMA is useful, for instance, for identifying the point in time of the potential change in the direction or significance of the effect size when the ordering variable is time. You can use the `cumulative()` option to perform CMA.

For demonstration purposes, let's use year as our ordering variable.

```
. meta summarize, cumulative(year)
Cumulative meta-analysis summary           Number of studies =    10
Random-effects model
Method: REML
Order variable: year
theta: Overall Std. Mean Diff.
```

Study	theta	[95% Conf. Interval]	P-value	year
Conn et al., 1968	0.120	-0.168 0.408	0.414	1968
Evans & Rosent~1969	-0.001	-0.166 0.165	0.995	1969
Claiborn, 1969	-0.042	-0.201 0.117	0.605	1969
Kester, 1969	0.022	-0.177 0.221	0.830	1969
Maxwell, 1970	0.140	-0.178 0.459	0.389	1970
Jose & Cody, 1971	0.089	-0.177 0.355	0.510	1971
Fielder et al., 1~1	0.064	-0.141 0.270	0.539	1971
Pellegrini & H~1972	0.161	-0.117 0.438	0.257	1972
Pellegrini & H~1972	0.161	-0.090 0.413	0.208	1972
Rosenthal et., 1~4	0.134	-0.075 0.342	0.208	1974

The output table reports the overall effect size and its CIs for each cumulative analysis. The P-value column contains the *p*-values of the significance tests of the overall effect sizes from these analyses. The last column displays the values of the ordering variable.

In our example, no particular trend is apparent.

We can perform stratified CMA by specifying a categorical variable in `cumulative()`'s option `by()`. To demonstrate, we also specify `cumulative()`'s option `descending` to list results in descending order of year.

```
. meta summarize, cumulative(year, by(week1) descending)
Stratified cumulative meta-analysis summary      Number of studies =      10
Random-effects model
Method: REML
Order variable: year (descending)
Stratum: week1

theta: Overall Std. Mean Diff.
```

Study	theta	[95% Conf. Interval]	P-value	year
Group: <= 1 week				
Pellegrini & H~1972	0.260	-0.463 0.983	0.481	1972
Pellegrini & H~1972	0.718	-0.183 1.620	0.118	1972
Maxwell, 1970	0.755	0.320 1.190	0.001	1970
Kester, 1969	0.581	0.174 0.989	0.005	1969
Group: > 1 week				
Rosenthal et., 1~4	0.030	-0.215 0.275	0.810	1974
Fielder et al., 1~1	0.000	-0.156 0.156	0.998	1971
Jose & Cody, 1971	-0.026	-0.166 0.115	0.720	1971
Claiborn, 1969	-0.054	-0.188 0.080	0.429	1969
Evans & Rosent~1969	-0.056	-0.167 0.056	0.326	1969
Conn et al., 1968	-0.033	-0.137 0.071	0.535	1968

CMA is performed separately for each group of week1.

Also see *Cumulative meta-analysis* in [META] meta.

◀

Stored results

meta summarize stores the following in r():

Scalars

r(N)	number of observations
r(theta)	overall effect size
r(se)	standard error of overall effect size
r(ci_lb)	lower CI bound for overall effect size
r(ci_ub)	upper CI bound for overall effect size
r(tau2)	between-study variance
r(I2)	I^2 heterogeneity statistic (not for CE model)
r(H2)	H^2 heterogeneity statistic (not for CE model)
r(z)	z statistic for test of significance of overall effect size (when se() not specified)
r(t)	t statistic for test of significance of overall effect size (when se() specified)
r(df)	degrees of freedom for t distribution
r(p)	p -value for test of significance of overall effect size
r(Q)	Cochran's Q heterogeneity test statistic (not for CE model)
r(df_Q)	degrees of freedom for heterogeneity test
r(p_Q)	p -value for heterogeneity test
r(Q_b)	Cochran's Q statistic for test of group differences (for subgroup() with one variable)
r(df_Q_b)	degrees of freedom for test of group differences
r(p_Q_b)	p -value for test of group differences
r(seadj)	standard-error adjustment
r(level)	confidence level for CIs
r(pi_lb)	lower bound of prediction interval
r(pi_ub)	upper bound of prediction interval
r(pilevel)	confidence level for prediction interval
r(converged)	1 if converged, 0 otherwise (with iterative random-effects methods)

Macros

r(model)	meta-analysis model
r(method)	meta-analysis estimation method
r(subgroupvars)	names of subgroup-analysis variables
r(ordervar)	name of order variable used in option cumulative()
r(byvar)	name of variable used in suboption by() within option cumulative()
r(direction)	ascending or descending
r(seadjtype)	type of standard-error adjustment

Matrices

r(esgroup)	ESs and CIs from subgroup analysis
r(hetgroup)	heterogeneity summary from subgroup analysis
r(diffgroup)	results for tests of group differences from subgroup analysis
r(cumul)	results from cumulative meta-analysis

meta summarize also creates a system variable, `_meta_weight`, that contains study weights. Also see *Stored results* in [META] **meta set** and *Stored results* in [META] **meta esize** for other system variables.

Methods and formulas

Methods and formulas are presented under the following headings:

Fixed-effects and common-effect methods for combining study estimates

Inverse-variance method

Mantel–Haenszel method for binary outcomes

Peto’s method for odds ratios

Random-effects methods for combining study estimates

Iterative methods

Noniterative methods

Knapp–Hartung standard-error adjustment

Prediction intervals

Confidence intervals and significance test

Heterogeneity measures

Homogeneity test

Subgroup meta-analysis

Fixed-effects model

Random-effects model

Cumulative meta-analysis

The formulas and methods below are based on Veroniki et al. (2016), Viechtbauer et al. (2015), Borenstein et al. (2009), Schwarzer, Carpenter, and Rücker (2015), Kontopantelis and Reeves (2016), Fisher (2016), and Bradburn, Deeks, and Altman (2016).

Fixed-effects and common-effect methods for combining study estimates

Consider the data from K independent studies. Let $\hat{\theta}_j$ be the estimate of the population effect size θ_j reported by the j th study and $\hat{\sigma}_j^2$ be the corresponding estimate of the within-study variance, which is equal to the squared standard error of $\hat{\theta}_j$. $\hat{\theta}_j$ is one of Hedges’s g_j , Cohen’s d_j , $\ln(\widehat{OR}_j)$, $\ln(\widehat{RR}_j)$, and so on, as defined in *Methods and formulas* of [META] **meta esize**, or a generic (precomputed) effect size as declared by [META] **meta set**.

Consider a fixed-effects model (Hedges and Vevea 1998; Rice, Higgins, and Lumley 2018) from *Meta-analysis models* in [META] **Intro**,

$$\hat{\theta}_j = \theta_j + \epsilon_j \quad \epsilon_j \sim N(0, \hat{\sigma}_j^2)$$

where $\widehat{\sigma}_j^2$'s are treated as known values that do not require estimation. Under the assumption that $\theta_1 = \theta_2 = \dots = \theta_K = \theta$, the above fixed-effects model simplifies to a common-effect model (Hedges 1982; Rosenthal and Rubin 1982):

$$\widehat{\theta}_j = \theta + \epsilon_j \quad \epsilon_j \sim N(0, \widehat{\sigma}_j^2)$$

The estimation methods we describe below are the same for the two models, but the interpretation of the estimates is different; see [Comparison between the models and interpretation of their results](#) in [META] Intro. The two models estimate different population parameters. A common-effect model estimates the common effect $\theta_{\text{pop}} = \theta$, whereas a fixed-effects model estimates a weighted average of the study-specific effects $\widehat{\theta}_j$'s,

$$\theta_{\text{pop}} = \text{Ave}(\theta_j) = \frac{\sum_{j=1}^K W_j \theta_j}{\sum_{j=1}^K W_j}$$

where W_j 's represent true, unknown weights, which are defined in Rice, Higgins, and Lumley (2018, eq. 3). For simplicity, in what follows, we will use θ to mean θ_{pop} .

Inverse-variance method

Under the inverse-variance method, the MLE of θ is

$$\widehat{\theta}_{\text{IV}} = \frac{\sum_{j=1}^K \widehat{\theta}_j / \widehat{\sigma}_j^2}{\sum_{j=1}^K 1 / \widehat{\sigma}_j^2} = \frac{\sum_{j=1}^K w_j \widehat{\theta}_j}{\sum_{j=1}^K w_j}$$

where the weight $w_j = 1 / \widehat{\sigma}_j^2$ is used to estimate the true weight W_j . The inverse-variance method takes its name from the weights being the reciprocal of the effect-size variances.

The variance estimate of $\widehat{\theta}_{\text{IV}}$

$$\widehat{\text{Var}}(\widehat{\theta}_{\text{IV}}) = \frac{1}{w.}$$

where $w. = \sum_{j=1}^K w_j$.

Mantel–Haenszel method for binary outcomes

For meta-analysis that compares two binary outcomes, the Mantel–Haenszel method can be used to combine odds ratios (OR), risk ratios (RR), and risk differences (RD) instead of the inverse-variance method. The classical Mantel–Haenszel method (Mantel and Haenszel 1959) is used for OR, and its extension by Greenland and Robins (1985) is used for RR and RD. The Mantel–Haenszel method may be preferable with sparse data (Emerson 1994). This is the default pooling method in [meta esize](#) for the effect sizes mentioned above with fixed-effects and common-effect models.

Consider the following 2×2 table for the j th study.

group	event	no event	size
treatment	a_j	b_j	$n_{1j} = a_j + b_j$
control	c_j	d_j	$n_{2j} = c_j + d_j$

The sample size for the j th study is denoted by $n_j = n_{1j} + n_{2j}$.

For the overall risk difference, the formula is

$$\hat{\theta}_{\text{MH}} = \frac{\sum_{j=1}^K w_j^{(\text{MH})} \times \hat{\theta}_j}{\sum_{j=1}^K w_j^{(\text{MH})}}$$

where $\hat{\theta}_j$ is $\widehat{\text{RD}}$ from the j th study.

Unlike the inverse-variance method, with log odds-ratios and log risk-ratios, the Mantel–Haenszel method combines the individual effect sizes in the original metric and then takes the log to obtain the final overall log odds-ratio or log risk-ratio estimate,

$$\hat{\theta}_{\text{MH}} = \ln \left\{ \frac{\sum_{j=1}^K w_j^{(\text{MH})} \times \exp(\hat{\theta}_j)}{\sum_{j=1}^K w_j^{(\text{MH})}} \right\}$$

where $\hat{\theta}_j$ is $\ln(\widehat{\text{OR}})$ or $\ln(\widehat{\text{RR}})$ from the j th study.

The MH weights are defined as follows. In the formula for the overall risk difference, the weight assigned to each study is

$$w_j^{(\text{MH})} = \frac{n_{1j}n_{2j}}{n_j}$$

For the overall log risk-ratio, the j th weight is given by

$$w_j^{(\text{MH})} = \frac{n_{1j}c_j}{n_j}$$

And for the overall log odds-ratio, the j th weight is given by

$$w_j^{(\text{MH})} = \frac{b_jc_j}{n_j}$$

An estimator of the variance of the overall risk difference $\hat{\theta}_{\text{MH}} = \widehat{\text{RD}}_{\text{MH}}$ (Greenland and Robins 1985) is

$$\widehat{\text{Var}}(\widehat{\text{RD}}_{\text{MH}}) = \frac{\sum_{j=1}^K (a_j b_j n_{2j}^3 + c_j d_j n_{1j}^3) / n_{1j} n_{2j} n_j^2}{\left(\sum_{j=1}^K n_{1j} n_{2j} / n_j \right)^2}$$

An estimator of the variance of the overall log risk-ratio $\hat{\theta}_{\text{MH}} = \ln(\widehat{\text{RR}}_{\text{MH}})$ (Greenland and Robins 1985) is

$$\widehat{\text{Var}}\{\ln(\widehat{\text{RR}}_{\text{MH}})\} = \frac{\sum_{j=1}^K \{n_{1j}n_{2j}(a_j + c_j) - a_jc_jn_j\} / n_j^2}{\left(\sum_{j=1}^K a_jn_{2j} / n_j \right) \times \left(\sum_{j=1}^K c_jn_{1j} / n_j \right)}$$

And an estimator of the variance of the overall log odds-ratio $\hat{\theta}_{MH} = \ln(\widehat{OR}_{MH})$ (Robins, Breslow, and Greenland 1986a; Robins, Greenland, and Breslow 1986b) is

$$\widehat{\text{Var}} \left\{ \ln(\widehat{OR}_{MH}) \right\} = \frac{\sum_{j=1}^K P_j R_j}{2 \left(\sum_{j=1}^K R_j \right)^2} + \frac{\sum_{j=1}^K (P_j S_j + Q_j R_j)}{2 \sum_{j=1}^K R_j \sum_{j=1}^K S_j} + \frac{\sum_{j=1}^K Q_j S_j}{2 \left(\sum_{j=1}^K S_j \right)^2}$$

where

$$P_j = \frac{a_j + d_j}{n_j}, \quad Q_j = \frac{b_j + c_j}{n_j}, \quad R_j = \frac{a_j d_j}{n_j}, \quad \text{and} \quad S_j = \frac{b_j c_j}{n_j}$$

Greenland and Robins (1985) and Robins, Breslow, and Greenland (1986a) demonstrate consistency of all the above variance estimators in the two cases they call a sparse-data limiting model, in which the number of 2×2 tables (studies) increases but the cell sizes remain fixed, and a large-strata limiting model, in which the number of studies remains fixed but individual cell sizes increase.

Peto's method for odds ratios

An alternative to the Mantel–Haenszel method for combining odds ratios is the Peto's method (Peto et al. 1977; Yusuf et al. 1985). It is based on the inverse-variance method but uses an alternate way to compute the odds ratios (and consequently the log odds-ratio).

Let $\ln(\widehat{OR}_j^{\text{Peto}})$ be Peto's log odds-ratio for the j th study as defined in *Odds ratio* in [META] **meta esize**. Then, Peto's overall log odds-ratio is defined following the inverse-variance method as follows,

$$\hat{\theta}_{\text{Peto}} = \ln(\widehat{OR}^{\text{Peto}}) = \frac{\sum_{j=1}^K w_j \ln(\widehat{OR}_j^{\text{Peto}})}{\sum_{j=1}^K w_j}$$

where $w_j = 1/\hat{\sigma}_j^2 = \text{Var}(a_j)$ and $\text{Var}(a_j)$ is as defined in *Methods and formulas* of [META] **meta esize** of [META] **meta esize**.

The variance estimate is

$$\widehat{\text{Var}} \left\{ \ln(\widehat{OR}^{\text{Peto}}) \right\} = \frac{1}{\sum_{j=1}^K w_j}$$

Random-effects methods for combining study estimates

Suppose that the observed study-specific effect sizes represent a random sample from a population of effect sizes that is normally distributed with mean θ and variance τ^2 .

Consider a random-effects model (Hedges 1983; DerSimonian and Laird 1986) from *Meta-analysis models* in [META] **Intro**,

$$\hat{\theta}_j = \theta_j + \epsilon_j = \theta + u_j + \epsilon_j$$

where ϵ_j and u_j are assumed to be independent with $\epsilon_j \sim N(0, \hat{\sigma}_j^2)$ and $u_j \sim N(0, \tau^2)$.

The overall effect $E(\hat{\theta}_j) = \theta$ is estimated as the weighted average,

$$\hat{\theta}^* = \frac{\sum_{j=1}^K w_j^* \hat{\theta}_j}{\sum_{j=1}^K w_j^*} \quad (1)$$

where $w_j^* = 1/(\hat{\sigma}_j^2 + \hat{\tau}^2)$. The variance of $\hat{\theta}^*$ is estimated by

$$\widehat{\text{Var}}(\hat{\theta}^*) = \frac{1}{w^*}$$

where $w^* = \sum_{j=1}^K w_j^*$.

Different estimators of the between-study variance, τ^2 , lead to different estimators of θ . **meta summarize** supports seven estimation methods of τ^2 . Three methods are iterative: the maximum likelihood (ML) estimator (Hardy and Thompson 1996); the restricted maximum-likelihood (REML) estimator (Raudenbush 2009); and the empirical Bayes (EB) estimator (Morris 1983; Berkey et al. 1995), also known as the Paule–Mandel estimator (Paule and Mandel 1982). Four methods are noniterative (have a closed-form expression): DerSimonian–Laird (DL) estimator (DerSimonian and Laird 1986); Hedges estimator (HE) (Hedges 1983; Hedges and Olkin 1985), also known as the Cochran estimator or variance-component estimator; Hunter–Schmidt (HS) estimator (Schmidt and Hunter 2015); and Sidik–Jonkman (SJ) estimator (Sidik and Jonkman 2005).

The formulas for and properties of these estimators have been discussed at length in Veroniki et al. (2016). Expressions for these estimators are given in the more general context of meta-regression in *Methods and formulas* of [META] **meta regress**. Below, we provide the simplified expressions when no covariates (moderators) are included in the regression model. The simplified expressions were obtained by replacing the \mathbf{X} matrix with $K \times 1$ column vector of 1s.

Iterative methods

The ML method (Hardy and Thompson 1996; Thompson and Sharp 1999) computes the MLE of τ^2 by maximizing the following log-likelihood function,

$$\ln L_{\text{ML}}(\tau^2) = -\frac{K}{2} \ln(2\pi) - \frac{1}{2} \sum_{j=1}^K \ln(\hat{\sigma}_j^2 + \tau^2) - \frac{1}{2} \sum_{j=1}^K \frac{(\hat{\theta}_j - \hat{\theta}^*)^2}{\hat{\sigma}_j^2 + \tau^2}$$

with respect to τ^2 , where $\hat{\theta}^*$ is defined in (1) and is based on the current value of $\hat{\tau}^2$.

The ML method is asymptotically efficient but may produce biased results in small samples. The REML method estimates τ^2 by accounting for the uncertainty in the estimation of θ , which leads to nearly an unbiased estimate of τ^2 .

The REML log-likelihood function is

$$\ln L_{\text{REML}}(\tau^2) = \ln L_{\text{ML}}(\tau^2) - \frac{1}{2} \ln \left\{ \sum_{j=1}^K (\hat{\sigma}_j^2 + \tau^2)^{-1} \right\} + \frac{\ln(2\pi)}{2}$$

The EB estimator and a description of the iterative process for each estimator in this section is presented in the *Methods and formulas* of [META] **meta regress**.

Noniterative methods

The methods in this section do not make any assumptions about the distribution of the random effects. They also do not require any iteration.

The most popular noniterative estimation method is the DL method. This is a method of moment estimator for τ^2 , and it is defined as follows,

$$\hat{\tau}_{\text{DL}}^2 = \frac{Q - (K - 1)}{\sum_{j=1}^K w_j - \sum_{j=1}^K w_j^2 / \sum_{j=1}^K w_j}$$

where $Q = \sum_{j=1}^K w_j (\hat{\theta}_j - \hat{\theta}_{\text{IV}})^2$ and $w_j = 1/\hat{\sigma}_j^2$.

Because $\hat{\tau}_{\text{DL}}^2$ is negative when $Q < K - 1$, it is truncated at 0 in practice, and thus $\max(0, \hat{\tau}_{\text{DL}}^2)$ is used to estimate the between-study variance:

$$\hat{\tau}_{\text{DL}}^2 = \max \left\{ 0, \frac{\sum_{j=1}^K w_j (\hat{\theta}_j - \hat{\theta}_{\text{IV}})^2 - (K - 1)}{\sum_{j=1}^K w_j - \sum_{j=1}^K w_j^2 / \sum_{j=1}^K w_j} \right\}$$

The HE estimator is another method of moment estimator defined as follows,

$$\hat{\tau}_{\text{HE}}^2 = \max \left\{ 0, \frac{1}{K - 1} \sum_{j=1}^K (\hat{\theta}_j - \bar{\theta})^2 - \frac{1}{K} \sum_{j=1}^K \hat{\sigma}_j^2 \right\}$$

where $\bar{\theta} = (\sum_{j=1}^K \hat{\theta}_j)/K$.

The HS estimator is given by

$$\hat{\tau}_{\text{HS}}^2 = \max \left\{ 0, \frac{Q - K}{\sum_{j=1}^K w_j} \right\}$$

For the SJ estimator, consider an initial estimate of τ^2 , given by

$$\hat{\tau}_0^2 = \frac{\sum_{j=1}^K (\hat{\theta}_j - \bar{\theta})^2}{K}$$

Then, the estimator is defined as

$$\widehat{\tau}_{\text{SJ}}^2 = \frac{\sum_{j=1}^K w_j^{\text{SJ}} \left(\widehat{\theta}_j - \widehat{\theta}^{\text{SJ}} \right)^2}{K - 1}$$

where $w_j^{\text{SJ}} = \widehat{\tau}_0^2 / (\widehat{\sigma}_j^2 + \widehat{\tau}_0^2)$ and $\widehat{\theta}^{\text{SJ}} = \sum_{j=1}^K w_j^{\text{SJ}} \widehat{\theta}_j / \sum_{j=1}^K w_j^{\text{SJ}}$.

Knapp–Hartung standard-error adjustment

Hartung and Knapp (2001a) and Sidik and Jonkman (2002) proposed an adjustment to the variance of $\widehat{\theta}^*$ to account for the uncertainty in estimating τ^2 , which is used in the expression for weights. They proposed to multiply $\widehat{\text{Var}}(\widehat{\theta}^*) = 1/w^*$ by the following quadratic form,

$$q_{\text{KH}} = \frac{1}{K - 1} \sum_{j=1}^K w_j^* \left(\widehat{\theta}_j - \widehat{\theta}^* \right)^2$$

or by $\max(1, q_{\text{KH}})$.

The variance estimator for $\widehat{\theta}^*$ can then be defined as

$$\widehat{\text{Var}}_{\text{HK}} \left(\widehat{\theta}^* \right) = \begin{cases} q_{\text{KH}} \times 1/w^* & \text{with option se(khartung)} \\ \max(1, q_{\text{KH}}) \times 1/w^* & \text{with option se(khartung, truncated)} \end{cases}$$

Hartung (1999) established that the statistic

$$\frac{\widehat{\theta}^* - \theta}{\sqrt{\widehat{\text{Var}}_{\text{HK}} \left(\widehat{\theta}^* \right)}}$$

has a Student's t distribution with $K - 1$ degrees of freedom.

Correspondingly, the $(1 - \alpha) \times 100\%$ CI for θ using the Knapp–Hartung standard error is

$$\widehat{\theta}^* \pm t_{K-1, 1-\alpha/2} \sqrt{\widehat{\text{Var}}_{\text{HK}} \left(\widehat{\theta}^* \right)}$$

where $t_{K-1, 1-\alpha/2}$ denotes the $1 - \alpha/2$ quantile of the Student's t distribution with $K - 1$ degrees of freedom.

The test statistic for the significance test of an overall effect, $H_0: \theta = 0$, is

$$\frac{\hat{\theta}^*}{\sqrt{\widehat{\text{Var}}_{\text{HK}}(\hat{\theta}^*)}}$$

and has the Student's t distribution with $K - 1$ degrees of freedom.

Also see [Sidik and Jonkman \(2002, 2003\)](#) and [Cornell et al. \(2014\)](#) for more discussion about the Knapp–Hartung adjustment.

Prediction intervals

In a random-effects model, you can compute a prediction interval ([Higgins, Thompson, and Spiegelhalter 2009](#)) that estimates plausible ranges for θ in a future study. Compared with the CI, a prediction interval incorporates the uncertainty in estimating τ^2 in the computation.

A $(1 - \alpha) \times 100\%$ prediction interval is defined as

$$\hat{\theta}^* \pm t_{K-2, 1-\alpha/2} \sqrt{\widehat{\text{Var}}(\hat{\theta}^*) + \hat{\tau}^2}$$

where $t_{K-1, 1-\alpha/2}$ denotes the $1 - \alpha/2$ quantile of the Student's t distribution with $K - 2$ degrees of freedom. This prediction interval may be specified with the `predinterval()` option.

Confidence intervals and significance test

Let $\hat{\theta}$ be any of the estimators considered in the previous sections such as $\hat{\theta}_{\text{IV}}$ or $\hat{\theta}^*$. The $(1 - \alpha) \times 100\%$ confidence interval for θ is

$$\hat{\theta} \pm z_{1-\alpha/2} \sqrt{\widehat{\text{Var}}(\hat{\theta})}$$

where $z_{1-\alpha/2}$ is the $(1 - \alpha/2)$ th quantile of the standard normal distribution.

We reject the hypothesis of no treatment effect $H_0: \theta = 0$ at level α , if

$$\frac{|\hat{\theta}|}{\sqrt{\widehat{\text{Var}}(\hat{\theta})}} > z_{1-\alpha/2}$$

If the `tdistribution` option is specified, the $z_{1-\alpha/2}$ critical value is replaced with the $t_{K-1, 1-\alpha/2}$ critical value in the above formulas.

Heterogeneity measures

The homogeneity test can be used to test whether the study-specific effects are the same; see *Homogeneity test*. But with a small number of studies, this test may have low power (Hedges and Pigott 2001). Also, it does not provide an estimate of the magnitude of the between-study heterogeneity. Some authors (for example, Higgins and Thompson [2002] and Higgins et al. [2003]) suggest examining the heterogeneity statistics rather than relying solely on the homogeneity test.

Higgins and Thompson (2002) proposed two heterogeneity measures: I^2 and H^2 . We define them separately for random-effects and fixed-effects models.

For a random-effects model, the two heterogeneity measures are defined as follows:

$$I^2 = \frac{\hat{\tau}^2}{\hat{\tau}^2 + s^2} \times 100\% \quad (2)$$

and

$$H^2 = \frac{\hat{\tau}^2 + s^2}{s^2} \quad (3)$$

where

$$s^2 = \frac{K - 1}{\sum_{j=1}^K w_j - \sum_{j=1}^K w_j^2 / \sum_{j=1}^K w_j}$$

is the within-study variance and $\hat{\tau}^2$ is an estimator of the between-study variance. The values of I^2 and H^2 will vary depending on which estimator of $\hat{\tau}^2$ is specified in the `random()` option.

For a fixed-effects model, the expressions for I^2 and H^2 are given by

$$I^2 = \left\{ \frac{Q - (K - 1)}{Q} \right\} \times 100\%$$

and

$$H^2 = \frac{Q}{K - 1}$$

where Q is defined in *Homogeneity test*.

The formulas above for I^2 and H^2 are equivalent to the corresponding formulas (2) and (3), when the DL method is used to estimate τ^2 . I^2 is negative when $Q < (K - 1)$ and is thus reset to zero in that case.

Homogeneity test

Consider a test of $H_0: \theta_1 = \theta_2 = \dots = \theta_K = \theta$, known as the homogeneity test, that evaluates whether the effect sizes are the same across the studies. It uses the following test statistic,

$$Q = \sum_{j=1}^K w_j (\hat{\theta}_j - \hat{\theta})^2 = \sum_{j=1}^K w_j \hat{\theta}_j^2 - \frac{\left(\sum_{j=1}^K w_j \hat{\theta}_j \right)^2}{\sum_{j=1}^K w_j}$$

where w_j , $\hat{\theta}_j$, and $\hat{\theta}$ depend on the type of the effect size and the chosen meta-analysis model and estimation method.

Under the null hypothesis of homogeneity, Q follows a χ^2 distribution with $K - 1$ degrees of freedom.

Hedges and Pigott (2001) showed that the test has low power when the number of studies (K) is small, which is typical in meta-analysis. This means that the null hypothesis of homogeneity is not rejected as often as it should be. Thus, for the homogeneity test, the meta-analysis literature (for example, Petitti [2001]; Berman and Parker [2002]; Sutton and Higgins [2008]) suggests using the significance level $\alpha = 0.1$ instead of the conventional $\alpha = 0.05$.

The homogeneity test checks for the potential presence of heterogeneity but does not estimate the magnitude of the heterogeneity. Thus, many authors (for example, Higgins and Thompson [2002]; Higgins et al. [2003]) suggest exploring the heterogeneity statistics rather than solely relying on the test. See *Heterogeneity measures*.

Subgroup meta-analysis

When the `subgroup(varname)` option is specified, we assume that the K studies are partitioned into L subgroups defined by `varname`. Estimates of the overall effect size and their corresponding standard errors are calculated for each of the L subgroups.

Let $\hat{\theta}_{jl}$ be the effect-size estimate from study j within subgroup l and $\hat{\sigma}_{jl}^2$ be the corresponding variance, where $l = 1, 2, \dots, L$ and $j = 1, 2, \dots, K_l$.

Below, we describe the formulas separately for fixed-effects and random-effects models. The formulas for the common-effect model are the same as for the fixed-effects model. When you specify a common-effect model with subgroup analysis, this model is assumed within each subgroup $l = 1, 2, \dots, L$, but not for the entire sample of studies.

Fixed-effects model

In what follows, we assume the inverse-variance method, but the same principles apply to the Mantel–Haenszel method.

In subgroup analysis, a fixed-effects model may be formulated as

$$\hat{\theta}_{jl} = \theta_{jl} + \epsilon_{jl}, \quad \epsilon_{jl} \sim N(0, \hat{\sigma}_{jl}^2)$$

For the l th group, $\hat{\theta}_{IV,l}$ is a weighted average of the effect sizes $\hat{\theta}_{jl}$ with weights $w_{jl} = 1/\hat{\sigma}_{jl}^2$:

$$\hat{\theta}_{IV,l} = \frac{\sum_{j=1}^{K_l} w_{jl} \hat{\theta}_{jl}}{\sum_{j=1}^{K_l} w_{jl}}$$

The variance estimate of $\hat{\theta}_{IV,l}$ is

$$\widehat{\text{Var}}(\hat{\theta}_{IV,l}) = \frac{1}{w_l}$$

where $w_l = \sum_{j=1}^{K_l} w_{jl}$.

Other meta-analytic quantities such as I_l^2 and Q_l may also be computed for the l th subgroup just as we described in the previous sections.

The Cochran's Q statistic can be extended to test for differences between the L subgroups:

$$Q_b = \sum_{l=1}^L w_{.l} \left(\hat{\theta}_{IV,l} - \frac{\sum_{l=1}^L w_{.l} \hat{\theta}_{IV,l}}{\sum_{l=1}^L w_{.l}} \right)^2$$

The subscript b in Q_b stands for “between” to emphasize that Q_b tests for “between-group” differences.

Under the null hypothesis of homogeneity between the subgroups ($\theta_{.1} = \theta_{.2} = \dots = \theta_{.L} = \theta$), the statistic Q_b has a χ^2 distribution with $L - 1$ degrees of freedom.

Random-effects model

Consider a random-effects model with L subgroups and separate between-study variances τ_l^2 :

$$\hat{\theta}_{jl} = \theta_{.l} + u_{jl} + \epsilon_{jl} \quad \epsilon_{jl} \sim N(0, \hat{\sigma}_{jl}^2) \quad u_{jl} \sim N(0, \tau_l^2)$$

The formulas for the random-effects model are the same as for the above fixed-effects model, except we replace the weights with the random-effects weights.

The estimate, $\hat{\theta}_l^*$, and its variance in the l th group are

$$\hat{\theta}_l^* = \frac{\sum_{j=1}^{K_l} w_{jl}^* \hat{\theta}_{jl}}{\sum_{j=1}^{K_l} w_{jl}^*}$$

$$\widehat{\text{Var}}(\hat{\theta}_l^*) = \frac{1}{w_{.l}^*}$$

where $w_{jl}^* = 1/(\hat{\sigma}_{jl}^2 + \hat{\tau}_l^2)$ and $w_{.l}^* = \sum_{j=1}^{K_l} w_{jl}^*$.

The Cochran's statistic for testing differences between the L subgroups is defined as

$$Q_b^* = \sum_{l=1}^L w_{.l}^* \left(\hat{\theta}_l^* - \frac{\sum_{l=1}^L w_{.l}^* \hat{\theta}_l^*}{\sum_{l=1}^L w_{.l}^*} \right)^2$$

Under the null hypothesis of homogeneity between the subgroups ($\theta_{.1} = \theta_{.2} = \dots = \theta_{.L} = \theta$), Q_b^* has a χ^2 distribution with $L - 1$ degrees of freedom.

Also see [Borenstein et al. \(2009, chap. 19\)](#) and [Schwarzer, Carpenter, and Rucker \(2015\)](#).

Cumulative meta-analysis

To perform CMA, we first sort the studies in ascending order according to the values of the variable specified in the `cumulative()` option. If suboption `descending` is specified within the `cumulative()` option, the order is reversed. Mathematically, this corresponds to sorting the pairs $(\hat{\theta}_j, \hat{\sigma}_j^2)$ in the specified order. Let $(\hat{\theta}_j^s, \hat{\sigma}_j^{2,s})$ denote the sorted pairs.

CMA estimates K overall effect sizes $\widehat{\theta}_j^c$'s as follows,

$$\begin{aligned}\widehat{\theta}_1^c &= \widehat{\theta}_1^s \\ \widehat{\theta}_2^c &= \text{MA} \left(\widehat{\theta}_1^s, \widehat{\theta}_2^s \right) \\ \widehat{\theta}_3^c &= \text{MA} \left(\widehat{\theta}_1^s, \widehat{\theta}_2^s, \widehat{\theta}_3^s \right) \\ &\vdots \\ \widehat{\theta}_j^c &= \text{MA} \left(\widehat{\theta}_1^s, \widehat{\theta}_2^s, \widehat{\theta}_3^s, \dots, \widehat{\theta}_j^s \right) \\ &\vdots \\ \widehat{\theta}_K^c &= \text{MA} \left(\widehat{\theta}_1^s, \widehat{\theta}_2^s, \widehat{\theta}_3^s, \dots, \widehat{\theta}_K^s \right)\end{aligned}$$

where $\text{MA} \left(\widehat{\theta}_1^s, \widehat{\theta}_2^s, \widehat{\theta}_3^s, \dots, \widehat{\theta}_j^s \right)$ denotes a meta-analysis applied to the sorted studies 1 through j . Note that the meta-analysis also depends on the values $\widehat{\sigma}_j^{2,s}$ but we omitted them from $\text{MA}()$ for notational convenience.

If suboption `by(byvar)` is specified within the `cumulative()` option, the above procedure is repeated for each subgroup defined by variable `byvar`.

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Also see

- [META] [meta data](#) — Declare meta-analysis data
- [META] [meta forestplot](#) — Forest plots
- [META] [meta regress](#) — Meta-analysis regression
- [META] [meta](#) — Introduction to meta
- [META] [Glossary](#)
- [META] [Intro](#) — Introduction to meta-analysis

Title

meta labbeplot — L'Abbé plots

Description	Quick start	Menu	Syntax
Options	Remarks and examples	Stored results	Methods and formulas
References	Also see		

Description

`meta labbeplot` produces L'Abbé plots for a meta-analysis that compares the binary outcomes of two groups. These plots are useful for assessing heterogeneity and comparing study-specific event rates in the two groups.

Quick start

Construct a L'Abbé plot based on the effect size for binary data computed by `meta esize`

```
meta labbeplot
```

As above, but request that the overall effect size be computed using a random-effects REML method instead of the default common-effect inverse-variance method

```
meta labbeplot, random(reml)
```

As above, but specify that study-marker sizes be proportional to weights from a random-effects model instead of the default common-effect model

```
meta labbeplot, random(reml) reweighted
```

Modify the default looks of the reference line and the overall effect-size line

```
meta labbeplot, rlopts(lcolor(red)) esopts(lpattern(solid))
```

Menu

Statistics > Meta-analysis

Syntax

meta labbeplot [*if*] [*in*] [, *options*]

<i>options</i>	Description
----------------	-------------

Main	
<u>random</u> [<i>(remethod)</i>]	random-effects meta-analysis
<u>common</u> [<i>(cefemethod)</i>]	common-effect meta-analysis
<u>fixed</u> [<i>(cefemethod)</i>]	fixed-effects meta-analysis
<u>reweighted</u>	make bubble size depend on random-effects weights
[<i>no</i>] <u>metashow</u>	display or suppress meta settings in the output
<u>graph_options</u>	affect rendition of overall L'Abbé plot

<i>remethod</i>	Description
-----------------	-------------

<u>reml</u>	restricted maximum likelihood; the default
<u>mle</u>	maximum likelihood
<u>ebayes</u>	empirical Bayes
<u>dlaird</u>	DerSimonian–Laird
<u>sjonkman</u>	Sidik–Jonkman
<u>hedges</u>	Hedges
<u>hschmidt</u>	Hunter–Schmidt

<i>cefemethod</i>	Description
-------------------	-------------

<u>mhaenszel</u>	Mantel–Haenszel
<u>invvariance</u>	inverse variance
<u>ivariance</u>	synonym for invvariance

<i>graph_options</i>	Description
----------------------	-------------

RL options	
<u>rlopts</u> (<i>line_options</i>)	affect rendition of the plotted reference line indicating no effect
ES options	
<u>esopts</u> (<i>line_options</i>)	affect rendition of the plotted estimated effect-size line
Add plots	
<u>addplot</u> (<i>plot</i>)	add other plots to the contour plot
Y axis, X axis, Titles, Legend, Overall	
<u>twoway_options</u>	any options other than by() documented in [G-3] <i>twoway_options</i>

Options

Main

Options `random()`, `common()`, and `fixed()` specify a meta-analysis model to use when estimating the overall effect size. For historical reasons, the default is `common(invvariance)`, regardless of the global model declared by `meta esize`. Specify one of these options with `meta labbeplot` to override this default. Options `random()`, `common()`, and `fixed()` may not be combined. Also see *Meta-analysis models* in [META] [Intro](#).

`random` and `random(remethod)` specify that a random-effects model be assumed for meta-analysis; see *Random-effects model* in [META] [Intro](#).

remethod specifies the type of estimator for the between-study variance τ^2 . *remethod* is one of `reml`, `mle`, `ebayes`, `dlaird`, `sjonkman`, `hedges`, or `hschmidt`. `random` is a synonym for `random(reml)`. See *Options* in [META] [meta esize](#) for more information.

`common` and `common(cefemethod)` specify that a common-effect model be assumed for meta-analysis; see *Common-effect (“fixed-effect”) model* in [META] [Intro](#). Also see the [discussion](#) in [META] [meta data](#) about common-effect versus fixed-effects models.

`common` implies `common(mhaenszel)`.

cefemethod is one of `mhaenszel` or `invvariance` (synonym `ivariance`). See *Options* in [META] [meta esize](#) for more information.

`fixed` and `fixed(cefemethod)` specify that a fixed-effects model be assumed for meta-analysis; see *Fixed-effects model* in [META] [Intro](#). Also see the [discussion](#) in [META] [meta data](#) about fixed-effects versus common-effect models.

`fixed` implies `fixed(mhaenszel)`.

cefemethod is one of `mhaenszel` or `invvariance` (synonym `ivariance`); see *Options* in [META] [meta esize](#) for more information.

`reweighted` is used with random-effects meta-analysis. It specifies that the sizes of the bubbles be proportional to the weights from the random-effects meta-analysis, $w_j^* = 1/(\hat{\sigma}_j^2 + \hat{\tau}^2)$. By default, the sizes are proportional to the precision of each study, $w_j = 1/\hat{\sigma}_j^2$.

`metashow` and `nometashow` display or suppress the meta setting information. By default, this information is displayed at the top of the output. You can also specify `nometashow` with `meta update` to suppress the meta setting output for the entire meta-analysis session.

RL options

`rlopts(line_options)` affects the rendition of the plotted reference (diagonal) line that indicates no effect of the intervention or treatment; see [G-3] [line_options](#).

ES options

`esopts(line_options)` affects the rendition of the dashed line that plots the estimated overall effect size; see [G-3] [line_options](#).

Add plots

`addplot(plot)` allows adding more graph twoway plots to the graph; see [G-3] [addplot_option](#).

Y axis, X axis, Titles, Legend, Overall

`twoway_options` are any of the options documented in [G-3] [twoway_options](#), excluding `by()`. These include options for titling the graph (see [G-3] [title_options](#)) and for saving the graph to disk (see [G-3] [saving_option](#)).

Remarks and examples

The L'Abbé plot (L'Abbé, Detsky, and O'Rourke 1987) is a scatterplot of the summary outcome measure such as log odds in the control group on the x axis and of that in the treatment group on the y axis. This plot is used with binary data declared by `meta esize`. The plotted summary outcome measure depends on the chosen effect size. It is log odds when the effect size is log odds-ratio, log risk when the effect size is log risk-ratio, and risk when the effect size is risk difference. The summary outcome measures are plotted as circles with their sizes (areas) proportional to study precisions. The plot also contains a reference (diagonal) line, which indicates identical outcomes in the two groups and thus represents no effect, and the estimated overall effect-size line.

The L'Abbé plot explores between-study heterogeneity by comparing group-level summary outcome measures across studies. It can also be used to determine which type of effect size is more homogeneous across studies. Compared with other meta-analysis graphs, one important advantage of the L'Abbé plot is that it displays the data on individual studies for each of the two groups. Thus, in addition to identifying outlying studies, it can also identify the outlying groups within studies. Also see Anzures-Cabrera and Higgins (2010) for more detail.

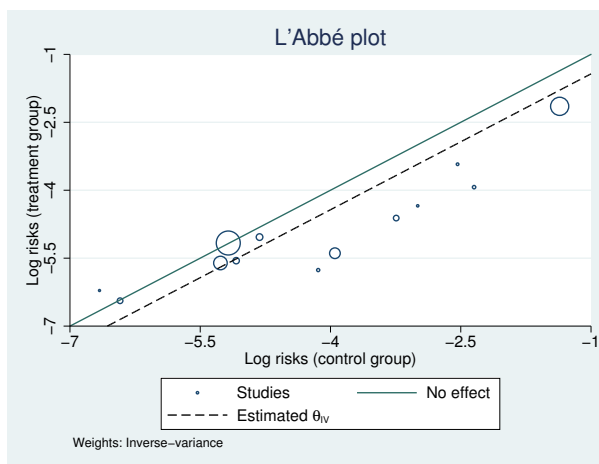
`meta labbeplot` produces L'Abbé plots. The plotted summary outcome measure is determined automatically based on the declared effect size. Unlike other `meta` commands, for historical reasons, `meta labbeplot` assumes a common-effect model with the inverse-variance method when computing the overall effect size to be plotted. You can use `random()`, `common()`, or `fixed()` to specify a different meta-analysis model or method. By default, `meta labbeplot` uses the precision weights, $1/\hat{\sigma}_j^2$, but, with a random-effects model, you can instead choose to use the random-effects weights, $1/(\hat{\sigma}_j^2 + \hat{\tau}^2)$. You can also control the look of the plotted reference and effect-size lines by specifying the `rlopts()` and `esopts()` options.

► Example 1: Basic L'Abbé plot

Consider the declared version of the BCG dataset, `bcgset.dta`, which we used in, for instance, [example 1](#) of [META] [meta regress](#). Let's produce the L'Abbé plot for these data.

```
. use https://www.stata-press.com/data/r16/bcgset
(Efficacy of BCG vaccine against tuberculosis; set with -meta esize-)
. meta labbeplot

Effect-size label:  Log Risk-Ratio
Effect size:       _meta_es
Std. Err.:        _meta_se
Summary data:     npost nnegt npostc nnegc
Model:            Common-effect
Method:          Inverse-variance
```



From the displayed meta settings, the declared effect size is a log risk-ratio. Thus, `meta labbeplot` plots the log risks on the scatterplot. The treatment-group log risk is on the y axis, and the control-group log risk is on the x axis. The sizes of the plotted markers (circles) are proportional to the precision of the trials. Large circles represent more precise, larger trials, whereas small circles represent less precise, smaller trials.

The solid reference line ($y = x$) represents the “no-effect” line. That is, the log risks (or risks) in the two groups for the trials on the line are either the same or very similar. There are two trials that are on the line in our example: one is a large trial, the other one is a small trial. The log risks for these trials are very similar in the two groups, and the corresponding log risk-ratios are close to zero.

If a circle is above the reference line, the risk in the treatment group is higher than the risk in the control group for that study. Conversely, if a circle is below the line, the risk in the treatment group is lower than the risk in the control group. In our example, one trial is above the reference line, suggesting that the risk in the treatment group is higher, but this is a very small trial. The remaining trials are below the line, suggesting that the risk is lower in the treatment group. However, the trials demonstrating large differences between the groups are also smaller (less precise) trials.

The dashed line is the overall effect-size line. The intercept of this line equals the estimate of the overall effect size, which is the overall log risk-ratio in our example. The actual estimate of the overall effect size is not important in the L'Abbé plot. What is important is whether the circles follow the effect-size line or deviate from it. When the circles deviate from the effect-size line greatly, this may be a sign of study heterogeneity. In our example, there are at least five trials that are far away

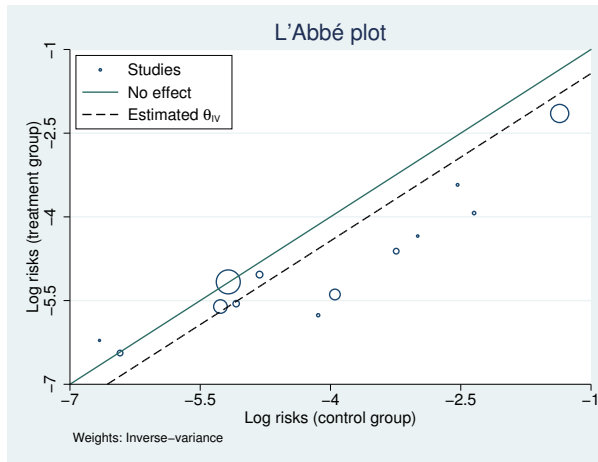
from the effect-size line. We should suspect the presence of heterogeneity in these data. In fact, we did establish in [example 1](#) of [\[META\] meta regress](#) that there is at least one moderator, the distance from the equator, that explains some of the variation in the trial effect sizes.

◀

► Example 2: Custom legend

Continuing with [example 1](#), let's demonstrate how we can customize the look of the legend produced by default.

```
. meta labbeplot, legend(cols(1) symxsize(*0.6) position(10) ring(0))
Effect-size label: Log Risk-Ratio
Effect size:      _meta_es
Std. Err.:       _meta_se
Summary data:    npost nnegt npostc nnegc
Model:           Common-effect
Method:          Inverse-variance
```



We customized the legend with a few suboptions specified in `legend()`. We used the `cols(1)` suboption to list all the legend keys in one column. We used `symxsize(*0.6)` to set the width of the key symbols to 60% of their default width. We used `position(10)` to position the label at 10 o'clock and `ring(0)` to place the legend inside the plot region.

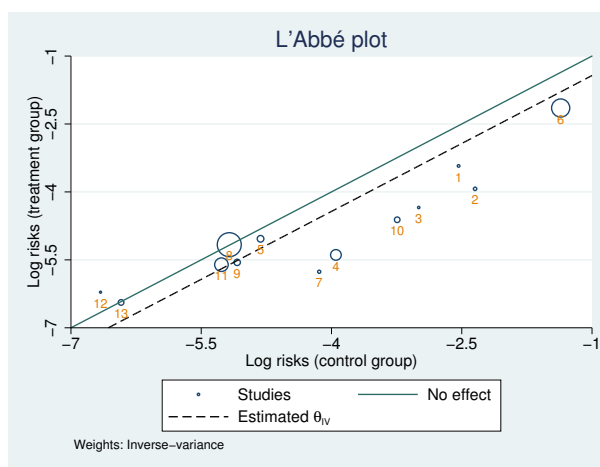
◀

► Example 3: Labeling trials

Continuing with [example 1](#), let's say it would be nice to mark the circles with the trial labels. We use the `addplot()` option and follow similar steps to those described in [example 3](#) of [\[META\] estat bubbleplot](#), except here we generate new variables for the added scatterplot.

```
. generate double lnriskt = ln(npost/(npost + nnegt))
. generate double lnriskc = ln(nposc/(nposc + nnegc))
. local opts msymbol(none) mlabel(trial) mlabpos(6)
. meta labbeplot, addplot(scatter lnriskt lnriskc, 'opts' legend(order(1 2 3)))
```

```
Effect-size label: Log Risk-Ratio
Effect size:      _meta_es
Std. Err.:       _meta_se
Summary data:    npost nnegt nposc nnegc
Model:           Common-effect
Method:          Inverse-variance
```



First, we generated two new variables, `lnriskt` and `lnriskc`, that contain the log risks in the treatment and control groups. Then, we used `addplot()` to overlay the same scatterplot as produced by `meta labbeplot` but without the markers and with marker labels. We specified other options to improve the look of the graph; see [example 3](#) of [\[META\] estat bubbleplot](#) for details. Also see [example 4](#) of [\[META\] estat bubbleplot](#) for how to further improve the positioning of the labels.

Stored results

meta labbeplot stores the following in `r()`:

Scalars

<code>r(theta)</code>	estimated overall effect size
<code>r(xmin)</code>	minimum value in the control group (x axis)
<code>r(xmax)</code>	maximum value in the control group
<code>r(ymin)</code>	minimum value in the treatment group (y axis)
<code>r(ymax)</code>	maximum value in the treatment group

Macros

<code>r(model)</code>	meta-analysis model
<code>r(method)</code>	meta-analysis estimation method

Methods and formulas

Let a_j , b_j , c_j , and d_j define cell counts of a 2×2 table for study j ; see [Effect sizes for binary outcomes](#) in [META] **meta esize**. Let y_j and x_j be the summary measures such as log odds for study j in the treatment and control groups. The L'Abbé plot produces a scatterplot of (y_j, x_j) with the sizes of markers (areas of circles) proportional to the weights $w_j = 1/\hat{\sigma}_j^2$ or, if `reweighted` is specified with a random-effects model, $w_j = 1/(\hat{\sigma}_j^2 + \hat{\tau}^2)$.

When the effect size is risk difference, y_j and x_j are the risks given by

$$y_j = \frac{a_j}{a_j + b_j} \quad \text{and} \quad x_j = \frac{c_j}{c_j + d_j}$$

When the effect size is log risk-ratio, y_j and x_j are the log risks given by

$$y_j = \log\left(\frac{a_j}{a_j + b_j}\right) \quad \text{and} \quad x_j = \log\left(\frac{c_j}{c_j + d_j}\right)$$

When the effect size is log odds-ratio, y_j and x_j are the log odds given by

$$y_j = \log\left(\frac{a_j}{b_j}\right) \quad \text{and} \quad x_j = \log\left(\frac{c_j}{d_j}\right)$$

The plotted reference line is the diagonal line. Studies that have the same values of the summary outcome measures in the two groups will have $y_j = x_j$ and thus will fall on the reference line.

The effect-size (dashed) line is a 45-degree line with an intercept equal to the estimated overall effect size. By default, the overall effect-size is estimated assuming a common-effect model with the inverse-variance method, but this can be changed by specifying one of `random()`, `common()`, or `fixed()`.

References

- Anzures-Cabrera, J., and J. P. T. Higgins. 2010. Graphical displays for meta-analysis: An overview with suggestions for practice. *Research Synthesis Methods* 1: 66–80.
- L'Abbé, K. A., A. S. Detsky, and K. O'Rourke. 1987. Meta-analysis in clinical research. *Annals of Internal Medicine Journal* 107: 224–233.

Also see

[META] **meta data** — Declare meta-analysis data

[META] **meta esize** — Compute effect sizes and declare meta-analysis data

[META] **meta regress** — Meta-analysis regression

[META] **meta** — Introduction to meta

[META] **Glossary**

[META] **Intro** — Introduction to meta-analysis

Title

meta regress — Meta-analysis regression

Description	Quick start	Menu	Syntax
Options	Remarks and examples	Stored results	Methods and formulas
References	Also see		

Description

`meta regress` performs meta-analysis regression, or meta-regression, which is a linear regression of the study effect sizes on study-level covariates (moderators). Meta-regression investigates whether between-study heterogeneity can be explained by one or more moderators. You can think of meta-regression as a standard meta-analysis that incorporates moderators into the model. `meta regress` performs both random-effects and fixed-effects meta-regression.

Quick start

Perform meta-regression of the effect size, `_meta_es`, on covariate (moderator) `x1`

```
meta regress x1
```

As above, but assume a DerSimonian–Laird random-effects method instead of the method declared by either `meta set` or `meta esize`

```
meta regress x1, random(dlaird)
```

Add a factor variable `a`, and request a Knapp–Hartung adjustment to the standard errors of coefficients

```
meta regress x1 i.a, random(dlaird) se(khartung)
```

Perform a sensitivity analysis by assuming a fixed value of 0.2 for the between-study variance τ^2

```
meta regress x1 i.a, tau2(0.2)
```

Menu

Statistics > Meta-analysis

Syntax

Meta-regression using meta-analysis model as declared with meta set or meta esize

```
meta regress moderators [if] [in] [, reopts options]
```

Random-effects meta-regression

```
meta regress moderators [if] [in], random[ (remethod) ] [reopts options]
```

Fixed-effects meta-regression

```
meta regress moderators [if] [in], fixed [multiplicative options]
```

Constant-only meta-regression

```
meta regress _cons [if] [in] [, modelopts]
```

<i>reopts</i>	Description
<u>tau2</u> (#)	sensitivity meta-analysis using a fixed value of between-study variance τ^2
<u>i2</u> (#)	sensitivity meta-analysis using a fixed value of heterogeneity statistic I_{res}^2
<u>se</u> (<i>seadj</i>)	adjust standard errors of the coefficients

<i>options</i>	Description
Model	
<u>noconstant</u>	suppress constant term
<u>tdistribution</u>	report <i>t</i> tests instead of <i>z</i> tests for the coefficients
Reporting	
<u>level</u> (#)	set confidence level; default is as declared for meta-analysis
<u>noheader</u>	suppress output header
[<u>no</u>] <u>metashow</u>	display or suppress meta settings in the output
<u>display_options</u>	control columns and column formats, row spacing, line width, display of omitted variables and base and empty cells, and factor-variable labeling
Maximization	
<u>maximize_options</u>	control the maximization process; seldom used
<u>coeflegend</u>	display legend instead of statistics

moderators may contain factor variables; see [U] 11.4.3 **Factor variables**.

coeflegend does not appear in the dialog box.

See [U] 20 **Estimation and postestimation commands** for more capabilities of estimation commands.

<i>remethod</i>	Description
<code>rem1</code>	restricted maximum likelihood; the default
<code>mle</code>	maximum likelihood
<code>ebayes</code>	empirical Bayes
<code>dlaird</code>	DerSimonian–Laird
<code>sjonkman</code>	Sidik–Jonkman
<code>hedges</code>	Hedges
<code>hschmidt</code>	Hunter–Schmidt

modelopts is any option except `noconstant`.

Options

Model

`noconstant`; see [R] [Estimation options](#). This option is not allowed with constant-only meta-regression.

Options `random()` and `fixed`, when specified with `meta regress`, temporarily override the global model declared by `meta set` or `meta esize` during the computation. Options `random()` and `fixed` may not be combined. If these options are omitted, the declared meta-analysis model is assumed; see [Declaring a meta-analysis model](#) in [META] [meta data](#). Also see [Meta-analysis models](#) in [META] [Intro](#).

`random` and `random(remethod)` specify that a random-effects model be assumed for meta-regression; see [Random-effects model](#) in [META] [Intro](#).

remethod specifies the type of estimator for the between-study variance τ^2 . *remethod* is one of `rem1`, `mle`, `ebayes`, `dlaird`, `sjonkman`, `hedges`, or `hschmidt`. `random` is a synonym for `random(rem1)`. See [Options](#) in [META] [meta esize](#) for more information.

`fixed` specifies that a fixed-effects model be assumed for meta-regression; see [Fixed-effects model](#) in [META] [Intro](#). It uses the inverse-variance estimation method; see [Meta-analysis estimation methods](#) in [META] [Intro](#).

reopts are `tau2(#)`, `i2(#)`, and `se(khartung[, truncated])`. These options are used with random-effects meta-regression.

`tau2(#)` specifies the value of the between-study variance parameter, τ^2 , to use for the random-effects meta-regression. This option is useful for exploring the sensitivity of the results to different levels of between-study heterogeneity. Only one of `tau2()` or `i2()` may be specified.

`i2(#)` specifies the value of the residual heterogeneity statistic I^2_{res} (as a percentage) to use for the random-effects meta-regression. This option is useful for exploring the sensitivity of the results to different levels of between-study heterogeneity. Only one of `i2()` or `tau2()` may be specified.

`se(seadj)` specifies that the adjustment *seadj* be applied to the standard errors of the coefficients. Additionally, the tests of significance of the coefficients are based on a Student's *t* distribution instead of the normal distribution.

seadj is `khartung[, truncated]`. Adjustment `khartung` specifies that the Knapp–Hartung adjustment (Hartung and Knapp 2001a, 2001b; Knapp and Hartung 2003), also known as the Sidik–Jonkman adjustment (Sidik and Jonkman 2002), be applied to the standard errors of the coefficients. `hknapp` and `sjonkman` are synonyms for `khartung`. `truncated` specifies

that the truncated Knapp–Hartung adjustment (Knapp and Hartung 2003), also known as the modified Knapp–Hartung adjustment, be used.

`multiplicative` performs a fixed-effects meta-regression that accounts for residual heterogeneity by including a multiplicative variance parameter ϕ . ϕ is referred to as an “(over)dispersion parameter”. See [Introduction](#) for details.

`tdistribution` reports t tests instead of z tests for the coefficients. This option is useful, for instance, when `meta regress` is used to conduct a regression-based test for funnel-plot asymmetry. Traditionally, the test statistic from this test is compared with critical values from a Student’s t distribution instead of the default normal distribution. This option may not be combined with option `se()`.

Reporting

`level(#)` specifies the confidence level, as a percentage, for confidence intervals. The default is as declared for the meta-analysis session; see [Declaring a confidence level for meta-analysis](#) in [\[META\] meta data](#). Also see option `level()` in [\[META\] meta set](#).

`noheader` suppresses the output header, either at estimation or upon replay.

`metashow` and `nometashow` display or suppress the meta setting information. By default, this information is displayed at the top of the output. You can also specify `nometashow` with `meta update` to suppress the meta setting output for the entire meta-analysis session.

display_options: `nocl`, `nopvalues`, `noomitted`, `vsquish`, `noemptycells`, `baselevels`, `allbaselevels`, `nofvlabel`, `fvwrap(#)`, `fvwrapon(style)`, `cformat(%fmt)`, `pformat(%fmt)`, `sformat(%fmt)`, and `nolstretch`; see [\[R\] Estimation options](#).

Maximization

maximize_options: `iterate(#)`, `tolerance(#)`, `nrtolerance(#)`, `nonrtolerance` (see [\[R\] Maximize](#)), `from(#)`, and `showtrace`. These options control the iterative estimation of the between-study variance parameter, τ^2 , with random-effects methods `reml`, `mle`, and `ebayes`. These options are seldom used.

`from(#)` specifies the initial value for τ^2 during estimation. By default, the initial value for τ^2 is the noniterative Hedges estimator.

`showtrace` displays the iteration log that contains the estimated parameter τ^2 , its relative difference with the value from the previous iteration, and the scaled gradient.

The following option is available with `meta regress` but is not shown in the dialog box:

`coeflegend`; see [\[R\] Estimation options](#).

Remarks and examples

Remarks are presented under the following headings:

[Introduction](#)

[Examples of using meta regress](#)

Introduction

Meta-regression is a regression performed in the context of meta-analysis. It is used to study the relationship between study effect sizes and covariates. Meta-regression is analogous to standard regression used when individual data are available, but in meta-regression, the observations are the studies, the outcome of interest is the effect size, and the covariates are recorded at the study level. The study-level covariates in meta-regression are known as moderators. Several examples of moderators include study location, study test environment, drug administration method. For a general overview and discussions about meta-regression, see [Berlin and Antman \(1992\)](#), [Berkey et al. \(1995\)](#), and [Thompson and Higgins \(2002\)](#).

The goal of meta-regression is to explore and explain the between-study heterogeneity as a function of moderators. Two types of regression models, fixed-effects (FE) and random-effects (RE), are available. An FE meta-regression assumes that all heterogeneity between study effect sizes can be accounted for by the included moderators. An RE meta-regression accounts for potential additional variability unexplained by the included moderators, also known as residual heterogeneity. Because a common-effect meta-analysis model implies no study heterogeneity, it is not applicable to meta-regression, except in a less interesting case of a constant-only model, which is equivalent to the standard common-effect meta-analysis; see [\[META\] meta summarize](#).

`meta regress` fits meta-regression. Use the `random()` option to fit an RE meta-regression and the `fixed` option to fit an FE meta-regression. Also see [Default meta-analysis model and method](#) in [\[META\] meta data](#) to learn about the default regression model used by `meta regress`.

For the j th study, let $\hat{\theta}_j$ denote the effect size, $\hat{\sigma}_j^2$ its variance, and \mathbf{x}_j be a $1 \times p$ vector of moderators with the corresponding unknown $p \times 1$ coefficient vector β .

An FE meta-regression ([Greenland 1987](#)) is given by

$$\hat{\theta}_j = \mathbf{x}_j\beta + \epsilon_j, \text{ weighted by } w_j = \frac{1}{\hat{\sigma}_j^2}, \text{ where } \epsilon_j \sim N(0, \hat{\sigma}_j^2)$$

Residual heterogeneity may be incorporated into an FE meta-regression via a multiplicative factor, ϕ , applied to each of the variances $\hat{\sigma}_j^2$. This leads to a multiplicative meta-regression or FE meta-regression with multiplicative dispersion parameter ([Thompson and Sharp 1999](#))

$$\hat{\theta}_j = \mathbf{x}_j\beta + \epsilon_j^\phi, \text{ weighted by } w_j = \frac{1}{\hat{\sigma}_j^2}, \text{ where } \epsilon_j^\phi \sim N(0, \hat{\sigma}_j^2\phi)$$

This regression model may be specified by the combination of `fixed` and `multiplicative` options.

Another method of incorporating residual heterogeneity is to include an additive between-study variance component, τ^2 , that leads to an RE meta-regression ([Berkey et al. 1995](#)), also known as a mixed model in the meta-analysis literature:

$$\hat{\theta}_j = \mathbf{x}_j\beta + \epsilon_j^* = \mathbf{x}_j\beta + u_j + \epsilon_j, \text{ weighted by } w_j^* = \frac{1}{\hat{\sigma}_j^2 + \hat{\tau}^2}, \text{ where } \epsilon_j^* \sim N(0, \hat{\sigma}_j^2 + \tau^2)$$

As we mentioned earlier, an RE meta-regression assumes that the moderators explain only part of the heterogeneity, and a random-effects term $u_j \sim N(0, \tau^2)$ is used to account for the remainder.

Harbord and Higgins (2016) point out that some authors (Thompson and Sharp 1999; Higgins and Thompson 2004) argue that an FE meta-regression should not be used because, in practice, the included moderators rarely capture all the between-study heterogeneity and that the failure of the FE regression to capture the extra between-study heterogeneity can lead to excessive type I errors. Also, the results from an FE meta-regression, including its multiplicative version, may not be generalized to populations from which the observed studies are a sample (Konstantopoulos and Hedges 2009). If you do not specify a meta-analysis model with `meta set` or `meta esize` during declaration, an RE meta-regression will be assumed by `meta regress`.

Meta-regression can also be considered an extension of subgroup analysis (see `meta summarize`, `subgroup()` in [META] `meta summarize`) to include continuous moderators in addition to the categorical ones. In particular, an FE meta-regression with the subgroup variable specified as a factor variable (see [U] 11.4.3 **Factor variables**) is equivalent to the FE subgroup analysis on that variable.

It is recommended that you have at least 10 studies per moderator to perform meta-regression (Borenstein et al. 2009, chap. 20). Otherwise, you may not be able to estimate the effects of moderators reliably. For more recommendations regarding meta-regression, see Schmidt and Hunter (2015, chap. 9), Deeks, Macaskill, and Irwig (2005), Harbord and Higgins (2016), Sharp (2016), and Thompson and Higgins (2002).

Examples of using meta regress

Consider a dataset from Colditz et al. (1994) of clinical trials that explore the efficacy of a Bacillus Calmette-Guérin (BCG) vaccine in the prevention of tuberculosis (TB). This dataset was introduced in *Efficacy of BCG vaccine against tuberculosis (bcg.dta)* of [META] `meta`. In this section, we use its declared version and focus on the demonstration of various options of `meta regress` and explanation of its output.

```
. use https://www.stata-press.com/data/r16/bcgset
(Efficacy of BCG vaccine against tuberculosis; set with -meta esize-)

. meta query, short
-> meta esize npost nnegt nposc nnegc, esize(lnrratio) studylabel(study1b1)

Effect-size label:  Log Risk-Ratio
Effect-size type:  lnrratio
Effect size:       _meta_es
Std. Err.:        _meta_se
Model:            Random-effects
Method:           REML
```

`meta query, short` reminds us about the main settings of the declaration step. Our data were declared by using `meta esize` with variables `npost`, `nnegt`, `nposc`, and `nnegc` representing the summary data from 2×2 tables, which record the numbers of positive and negative TB cases in the treatment and control groups. The computed effect sizes are log risk-ratios; their values and standard errors are stored in the respective **system variables** `_meta_es` and `_meta_se`. The `study1b1` variable supplies the study labels to be used in the output. The declared meta-analysis model is the default random-effects model with the REML estimation method.

Examples are presented under the following headings:

- Example 1: Random-effects meta-regression*
- Example 2: Sidik–Jonkman random-effects method*
- Example 3: Truncated Knapp–Hartung standard-error adjustment*
- Example 4: Sensitivity meta-analysis*
- Example 5: Fixed-effects meta-regression*
- Example 6: Multiplicative meta-regression*
- Example 7: Constant-only model*

► Example 1: Random-effects meta-regression

In [example 8](#) of [\[META\] meta](#), following [Berkey et al. \(1995\)](#), we fit a meta-regression with a centered absolute latitude, `latitude_c`, as the moderator to address heterogeneity. Let's refit this model here and focus on the specification and output from `meta regress`.

```
. meta regress latitude_c
      Effect-size label:  Log Risk-Ratio
      Effect size:      _meta_es
      Std. Err.:       _meta_se

Random-effects meta-regression          Number of obs =          13
Method: REML                           Residual heterogeneity:
                                          tau2 = .07635
                                          I2 (%) = 68.39
                                          H2 = 3.16
                                          R-squared (%) = 75.63
Wald chi2(1) = 16.36
Prob > chi2 = 0.0001
```

_meta_es	Coef.	Std. Err.	z	P> z	[95% Conf. Interval]
latitude_c	-.0291017	.0071953	-4.04	0.000	-.0432043 - .0149991
_cons	-.7223204	.1076535	-6.71	0.000	-.9333174 - .5113234

Test of residual homogeneity: $Q_{res} = \chi^2(11) = 30.73$ Prob > $Q_{res} = 0.0012$

Unlike with many Stata regression commands, we do not specify the dependent variable with `meta regress`. The command includes it automatically from the declared [meta settings](#). `meta regress` provides a short summary of the settings, which you can suppress with the `nometashow` option. [System variable](#) `_meta_es` contains the effect sizes and is thus used as the dependent variable. [System variable](#) `_meta_se` contains effect-size standard errors; it is used to construct the weights for the regression.

The header includes the information about the meta-analysis model and reports various summaries such as heterogeneity statistics and the model test. For example, the results are based on 13 studies. The reported I^2_{res} statistic is 68%, which still suggests moderate heterogeneity, using the categorization of [Higgins et al. \(2003\)](#), even after including `latitude_c` as the moderator. In other words, 68% of the variability in the residuals is still attributed to the between-study variation, whereas only 32% is attributed to the within-study variation. The adjusted R^2 statistic can be used to assess the proportion of between-study variance explained by the covariates; see [\(4\)](#) in [Methods and formulas](#) for its definition used in the meta-analysis literature. Here roughly 76% of the between-study variance is explained by the covariate `latitude_c`.

The output header also displays a model test that all coefficients other than the intercept are equal to zero based on the χ^2 distribution with $p - 1$ degrees of freedom. In our example, the χ^2 test statistic is 16.36 with a p -value of 0.0001. We have only one moderator, so the results of the model test in our example are equivalent to the z test (χ^2 value equals squared z value) of the coefficient of `latitude_c` reported in the output table.

The regression coefficient for `latitude_c` is -0.029 , which means that every one degree of latitude corresponds to a decrease of 0.0291 units in log risk-ratio. The intercept, $\widehat{\beta}_0$, is -0.722 , which means that the overall risk ratio at the mean latitude (`latitude_c = 0` corresponds to `latitude` ≈ 33.46) is $\exp(-0.722) = 0.46$. Both of these coefficients are statistically significantly different from zero based on the reported z tests.

Finally, a test of residual homogeneity is reported at the bottom of the output. The test statistic Q_{res} is 30.73 with a p -value of 0.0012, which suggests the presence of heterogeneity among the residuals.



□ Technical note

Heterogeneity statistics I^2_{res} and H^2_{res} , reported under **Residual heterogeneity:** in the header, are extensions of the corresponding statistics I^2 and H^2 from standard meta-analysis to meta-regression (Higgins and Thompson 2002). They measure the remaining between-study heterogeneity among the residuals after adjusting for the variability due to moderators. Similarly, the test of residual homogeneity based on the Q_{res} statistic is the extension of the standard meta-analysis homogeneity test based on the Cochran's Q statistic to meta-regression. See [Residual heterogeneity measures](#) and [Residual homogeneity test](#) in *Methods and formulas*.



▷ Example 2: Sidik–Jonkman random-effects method

Continuing with [example 1](#), let's demonstrate the use of a different RE method, for instance, the Sidik–Jonkman method, instead of the default REML method.

```
. meta regress latitude_c, random(sjonkman)
      Effect-size label:  Log Risk-Ratio
      Effect size:      _meta_es
      Std. Err.:       _meta_se

Random-effects meta-regression
Method: Sidik-Jonkman
Number of obs =      13
Residual heterogeneity:
      tau2 =      .2318
      I2 (%) =     86.79
      H2 =       7.57
      R-squared (%) = 32.90
Wald chi2(1) =      6.50
Prob > chi2 =      0.0108
```

_meta_es	Coef.	Std. Err.	z	P> z	[95% Conf. Interval]	
latitude_c	-.0280714	.0110142	-2.55	0.011	-.0496589	-.0064838
_cons	-.7410395	.1602117	-4.63	0.000	-1.055049	-.4270304

```
Test of residual homogeneity: Q_res = chi2(11) = 30.73  Prob > Q_res = 0.0012
```

The estimate of the regression coefficient for `latitude_c` is -0.028 and is similar to the REML estimate of -0.029 , but the standard errors are quite different: 0.011 versus 0.007. Recall that REML assumes that the error distribution is normal, whereas the Sidik–Jonkman estimator does not. Thus, its standard-error estimates are likely to be larger than those from REML. The estimates of the between-study variance, τ^2 , are also very different: 0.23 compared with the REML estimate of 0.08.



► Example 3: Truncated Knapp–Hartung standard-error adjustment

Continuing with [example 1](#), let's use an alternative standard-error computation sometimes used in practice—the truncated Knapp–Hartung method.

```
. meta regress latitude_c, se(khartung, truncated)
      Effect-size label:  Log Risk-Ratio
      Effect size:      _meta_es
      Std. Err.:       _meta_se

Random-effects meta-regression                Number of obs =      13
Method: REML                                Residual heterogeneity:
SE adjustment: Truncated Knapp-Hartung      tau2 = .07635
                                             I2 (%) = 68.39
                                             H2 = 3.16
                                             R-squared (%) = 75.63
Model F(1,11) = 12.59
Prob > F = 0.0046
```

_meta_es	Coef.	Std. Err.	t	P> t	[95% Conf. Interval]	
latitude_c	-.0291017	.0082014	-3.55	0.005	-.0471529	-.0110505
_cons	-.7223204	.1227061	-5.89	0.000	-.9923946	-.4522462

Test of residual homogeneity: $Q_{\text{res}} = \text{chi2}(11) = 30.73$ Prob > $Q_{\text{res}} = 0.0012$

The reported standard errors are larger than those from [example 1](#). This is expected because the Knapp–Hartung adjustment incorporates the uncertainty in estimating τ^2 in the standard-error computation. Also, the inferences for the tests of coefficients and the model test are now based on the Student's t and F distributions, respectively, instead of the default normal and χ^2 distributions.

◀

► Example 4: Sensitivity meta-analysis

We can perform sensitivity analysis to explore the impact of the various levels of heterogeneity on the regression results. Continuing with [example 1](#), let's fit a meta-regression assuming that the residual heterogeneity statistic I_{res}^2 equals 90%.

```
. meta regress latitude_c, i2(90)
      Effect-size label:  Log Risk-Ratio
      Effect size:      _meta_es
      Std. Err.:       _meta_se

Random-effects meta-regression                Number of obs =      13
Method: User-specified I2                    Residual heterogeneity:
                                             tau2 = .3176
                                             I2 (%) = 90.00
                                             H2 = 10.00
Wald chi2(1) = 4.89
Prob > chi2 = 0.0269
```

_meta_es	Coef.	Std. Err.	z	P> z	[95% Conf. Interval]	
latitude_c	-.0277589	.0125474	-2.21	0.027	-.0523514	-.0031664
_cons	-.7443082	.1812664	-4.11	0.000	-1.099584	-.3890326

Test of residual homogeneity: $Q_{\text{res}} = \text{chi2}(11) = 30.73$ Prob > $Q_{\text{res}} = 0.0012$

The estimate of the coefficient for `latitude_c` is now -0.028 with a standard-error estimate of 0.01 .

Let's now fit a meta-regression assuming the between-study variance of 0.01.

```
. meta regress latitude_c, tau2(0.01)
      Effect-size label:  Log Risk-Ratio
      Effect size:      _meta_es
      Std. Err.:       _meta_se

Random-effects meta-regression
Method: User-specified tau2

Number of obs =      13
Residual heterogeneity:
      tau2 =      .01
      I2 (%) =     22.08
      H2 =      1.28
Wald chi2(1) =     57.62
Prob > chi2 =     0.0000
```

_meta_es	Coef.	Std. Err.	z	P> z	[95% Conf. Interval]
latitude_c	-.0295601	.0038942	-7.59	0.000	-.0371926 - .0219277
_cons	-.6767043	.0617892	-10.95	0.000	-.7978089 - .5555998

Test of residual homogeneity: $Q_{res} = \text{chi2}(11) = 30.73$ Prob > $Q_{res} = 0.0012$

The specified value of τ^2 corresponds to the I^2_{res} value of 22.08%. The coefficient estimate is now -0.03 with a standard error of 0.004.

In both sensitivity analyses, `latitude_c` remained a statistically significant moderator for the log risk-ratios.

◀

▶ Example 5: Fixed-effects meta-regression

Instead of an RE meta-regression as in [example 1](#), we can use the `fixed` option to fit an FE meta-regression. The use of an FE meta-regression is usually discouraged in the meta-analysis literature because it assumes that all between-study heterogeneity is accounted for by the specified moderators ([Harbord and Higgins 2016](#); [Thompson and Sharp 1999](#); [Higgins and Thompson 2004](#)). This is often an unrealistic assumption in meta-analysis. We fit this model in our example for the purpose of demonstration.

```
. meta regress latitude_c, fixed
      Effect-size label:  Log Risk-Ratio
      Effect size:      _meta_es
      Std. Err.:       _meta_se

Fixed-effects meta-regression
Method: Inverse-variance

Number of obs =      13
Wald chi2(1) =     121.50
Prob > chi2 =     0.0000
```

_meta_es	Coef.	Std. Err.	z	P> z	[95% Conf. Interval]
latitude_c	-.0292369	.0026524	-11.02	0.000	-.0344356 - .0240383
_cons	-.6347482	.0445446	-14.25	0.000	-.7220541 - .5474423

Because the FE regression assumes no additional residual heterogeneity, the residual heterogeneity statistics and the residual homogeneity test are not reported with `meta regress, fixed`.

The coefficient estimates are similar to those from [example 1](#), but standard errors from the FE regression are smaller. This is because the FE regression does not account for the residual heterogeneity that is not explained by the included moderators.

Considering the presence of residual heterogeneity in these data, we should go back to our RE analysis or explore the multiplicative meta-regression, which we demonstrate in [example 6](#). ◀

▶ Example 6: Multiplicative meta-regression

An FE meta-regression in [example 5](#) does not account for residual heterogeneity. An extension of this regression model that does, known as a multiplicative meta-regression (see [Introduction](#) and [Methods and formulas](#)), has been considered in the meta-analysis literature. An RE meta-regression is the preferred analysis these days, but we provide the multiplicative meta-regression for completeness.

Continuing with [example 5](#), we add the multiplicative option to fit an FE meta-regression with a multiplicative dispersion parameter ϕ .

```
. meta regress latitude_c, fixed multiplicative
      Effect-size label:  Log Risk-Ratio
      Effect size:      _meta_es
      Std. Err.:       _meta_se

Fixed-effects meta-regression
Error: Multiplicative
Method: Inverse-variance

Number of obs =      13
Dispersion phi =     2.79
Wald chi2(1)  =     43.49
Prob > chi2   =     0.0000
```

_meta_es	Coef.	Std. Err.	z	P> z	[95% Conf. Interval]	
latitude_c	-.0292369	.0044335	-6.59	0.000	-.0379265	-.0205474
_cons	-.6347482	.0744564	-8.53	0.000	-.7806801	-.4888163

```
Test of residual homogeneity: Q_res = chi2(11) = 30.73  Prob > Q_res = 0.0012
```

The estimate of the dispersion parameter, reported in the header as Dispersion phi, is 2.79. It is greater than 1, which suggests the presence of residual heterogeneity in these data. The coefficient estimates are the same as those in [example 5](#), but the standard errors are about two times larger. ◀

▶ Example 7: Constant-only model

The primary use of `meta regress` is to fit meta-regression models containing moderators. You can also fit a constant-only model (without moderators), although this is less common in the context of meta-regression.

To fit a constant-only model with many regression estimation commands, you simply omit the covariates in the command specification. This would not work with `meta regress` because, without the dependent-variable specification, we would have to type

```
. meta regress
```

which means replaying previous estimation results consistently across Stata. The above will either issue an error that previous estimation results are not found or redisplay the results from the previous `meta regress` specification.

Instead, to fit a constant-only model with `meta regress`, you specify the designator `_cons` following the command name.

```
. meta regress _cons
      Effect-size label:  Log Risk-Ratio
      Effect size:      _meta_es
      Std. Err.:       _meta_se

Random-effects meta-regression
Method: REML
Number of obs =      13
Residual heterogeneity:
      tau2 =    .3132
      I2 (%) =   92.22
      H2 =    12.86
Wald chi2(0) =      .
Prob > chi2 =      .
```

<code>_meta_es</code>	Coef.	Std. Err.	z	P> z	[95% Conf. Interval]	
<code>_cons</code>	-.7145323	.1797815	-3.97	0.000	-1.066898	-.362167

Test of residual homogeneity: $Q_{res} = \text{chi2}(12) = 152.23$ Prob > $Q_{res} = 0.0000$

Note that the estimated value of $\hat{\tau}^2$ is now 0.313, whereas in [example 1](#) it was 0.076. That is, the inclusion of covariate `latitude_c` in [example 1](#) reduced $\hat{\tau}^2$ from 0.313 to 0.076 for a relative reduction of $(0.313 - 0.076)/0.313 \approx 76\%$.

The reason a constant-only meta-regression is not as common is because it produces the same results as a standard meta-analysis.

```
. meta summarize, nostudies
      Effect-size label:  Log Risk-Ratio
      Effect size:      _meta_es
      Std. Err.:       _meta_se
      Study label:     studylbl

Meta-analysis summary
Random-effects model
Method: REML
Number of studies =    13
Heterogeneity:
      tau2 =   0.3132
      I2 (%) =   92.22
      H2 =    12.86
```

theta: Overall Log Risk-Ratio

	Estimate	Std. Err.	z	P> z	[95% Conf. Interval]	
theta	-.7145323	.1797815	-3.97	0.000	-1.066898	-.362167

Test of homogeneity: $Q = \text{chi2}(12) = 152.23$ Prob > $Q = 0.0000$

See [\[META\] meta summarize](#) for details.

Stored results

`meta regress` stores the following in `e()`:

Scalars

<code>e(N)</code>	number of observations (studies)
<code>e(df_m)</code>	model degrees of freedom
<code>e(df_r)</code>	residual degrees of freedom
<code>e(chi2)</code>	model χ^2 Wald test statistic
<code>e(F)</code>	model F statistic
<code>e(p)</code>	p -value for model test
<code>e(phi)</code>	dispersion parameter
<code>e(tau2)</code>	between-study variance
<code>e(I2_res)</code>	I_{res}^2 heterogeneity statistic
<code>e(H2_res)</code>	H_{res}^2 heterogeneity statistic
<code>e(R2)</code>	R^2 heterogeneity measure
<code>e(Q_res)</code>	Cochran's Q residual homogeneity test statistic
<code>e(df_Q_res)</code>	degrees of freedom for residual homogeneity test
<code>e(p_Q_res)</code>	p -value for residual homogeneity test
<code>e(seadj)</code>	standard-error adjustment
<code>e(level)</code>	confidence level for CIs
<code>e(converged)</code>	1 if converged, 0 otherwise (with iterative random-effects methods)

Macros

<code>e(cmd)</code>	<code>meta regress</code>
<code>e(cmdline)</code>	command as typed
<code>e(depvar)</code>	name of dependent variable, <code>_meta_es</code>
<code>e(indepvars)</code>	names of independent variables (moderators)
<code>e(title)</code>	title in estimation output
<code>e(model)</code>	meta-analysis model
<code>e(method)</code>	meta-analysis estimation method
<code>e(seadjtype)</code>	type of standard-error adjustment
<code>e(properties)</code>	<code>b V</code>
<code>e(estat_cmd)</code>	program used to implement <code>estat</code>
<code>e(predict)</code>	program used to implement <code>predict</code>
<code>e(marginsok)</code>	predictions allowed by <code>margins</code>
<code>e(marginsnotok)</code>	predictions disallowed by <code>margins</code>
<code>e(marginsdefault)</code>	default <code>predict()</code> specification for <code>margins</code>
<code>e(asbalanced)</code>	factor variables <code>fvset</code> as <code>asbalanced</code>
<code>e(asobserved)</code>	factor variables <code>fvset</code> as <code>asobserved</code>

Matrices

<code>e(b)</code>	coefficient vector
<code>e(V)</code>	variance-covariance matrix of the estimators

Functions

<code>e(sample)</code>	marks estimation sample
------------------------	-------------------------

`meta regress` also creates a system variable, `_meta_regweight`, that contains meta-regression weights.

Methods and formulas

Methods and formulas are presented under the following headings:

Fixed-effects meta-regression
Random-effects meta-regression
 Iterative methods for computing $\hat{\tau}^2$
 Noniterative methods for computing $\hat{\tau}^2$
 Knapp-Hartung standard-error adjustment
Residual homogeneity test
Residual heterogeneity measures

Fixed-effects meta-regression

For an overview of estimation methods used by meta-regression, see [Berkey et al. \(1995\)](#), [Sidik and Jonkman \(2005\)](#), and [Viechtbauer et al. \(2015\)](#).

Consider an FE meta-analysis, where $\hat{\theta}_j \sim N(\theta_j, \hat{\sigma}_j^2)$, θ_j is the true effect size for study j , $\hat{\theta}_j$ is the estimated effect size, and $\hat{\sigma}_j^2$ is the variance of $\hat{\theta}_j$. In an FE meta-regression ([Greenland 1987](#)), the study-specific mean, θ_j , is expressed as

$$\theta_j = \beta_0 + \beta_1 x_{1j} + \dots + \beta_{p-1} x_{p-1,j} = \mathbf{x}_j \boldsymbol{\beta}$$

where $\mathbf{x}_j = (1, x_{1j}, \dots, x_{p-1,j})$ is a $1 \times p$ vector of categorical and continuous moderators (covariates) and $\boldsymbol{\beta}$ is a $p \times 1$ vector of regression coefficients to be estimated.

Defining $K \times p$ matrix $\mathbf{X} = (\mathbf{x}'_1, \mathbf{x}'_2, \dots, \mathbf{x}'_K)'$ and $\hat{\boldsymbol{\theta}} = (\hat{\theta}_1, \hat{\theta}_2, \dots, \hat{\theta}_K)'$. Let $w_j = 1/\hat{\sigma}_j^2$ be the weight associated with study j in an FE meta-analysis. The vector of estimated regression coefficients is

$$\hat{\boldsymbol{\beta}} = (\mathbf{X}' \mathbf{W} \mathbf{X})^{-1} \mathbf{X}' \mathbf{W} \hat{\boldsymbol{\theta}}$$

where $\mathbf{W} = \text{diag}(w_1, w_2, \dots, w_K)$.

The above FE regression does not account for residual heterogeneity. This can lead to coefficient standard errors that are too small. [Thompson and Sharp \(1999\)](#) incorporated residual heterogeneity into the model by including a multiplicative variance parameter:

$$\hat{\theta}_j \sim N(\mathbf{x}_j \boldsymbol{\beta}, \phi \sigma_j^2)$$

For a multiplicative FE meta-regression, \mathbf{W} in the above is replaced with $\mathbf{W}^\phi = \text{diag}(w_1^\phi, w_2^\phi, \dots, w_K^\phi)$, where the weights are defined as $w_j^\phi = 1/(\hat{\phi} \hat{\sigma}_j^2)$. $\hat{\phi}$ is estimated as the mean squared error from the weighted linear regression with weights proportional to $1/\hat{\sigma}_j^2$.

Next, we present another method of incorporating residual heterogeneity by including an additive between-study variance parameter.

Random-effects meta-regression

An RE meta-regression ([Berkey et al. 1995](#)) model may be expressed as

$$\hat{\theta}_j = \mathbf{x}_j \boldsymbol{\beta} + u_j + \epsilon_j \quad u_j \sim N(0, \tau^2) \quad \epsilon_j \sim N(0, \hat{\sigma}_j^2)$$

All algorithms for RE meta-regression first estimate the between-study variance, τ^2 . The regression coefficients are then estimated via weighted least squares,

$$\hat{\boldsymbol{\beta}}^* = (\mathbf{X}' \mathbf{W}^* \mathbf{X})^{-1} \mathbf{X}' \mathbf{W}^* \hat{\boldsymbol{\theta}}$$

where $\mathbf{W}^* = \text{diag}(w_1^*, w_2^*, \dots, w_K^*)$ and $w_j^* = 1/(\hat{\sigma}_j^2 + \hat{\tau}^2)$.

All the estimators of τ^2 can be expressed in terms of the matrix

$$\mathbf{P} = \mathbf{A} - \mathbf{A}\mathbf{X}(\mathbf{X}'\mathbf{A}\mathbf{X})^{-1}\mathbf{X}'\mathbf{A} \quad (1)$$

where \mathbf{A} is a $p \times p$ diagonal weight matrix whose elements depend on the type of estimator (Viechtbauer et al. 2015).

The formulas in the following sections are based on Viechtbauer et al. (2015).

Iterative methods for computing $\hat{\tau}^2$

The three estimators described below do not have a closed-form solution, and an iterative algorithm is needed to obtain an estimate of τ^2 . The Fisher scoring algorithm, described below, is used to estimate τ^2 .

All three estimators start with an initial estimate of τ^2 based on the Hedges estimator, $\hat{\tau}_0^2 = \hat{\tau}_{\text{HE}}^2$, but you can specify your own initial estimate in the `from()` option. The estimate is then updated at each iteration via the formula,

$$\hat{\tau}_{\text{new}}^2 = \hat{\tau}_{\text{current}}^2 + \delta$$

where δ is a function of $\hat{\tau}_{\text{current}}^2$ and its functional form depends on the estimation method.

The iteration terminates when `reldif($\hat{\tau}_{\text{new}}^2, \hat{\tau}_{\text{current}}^2$)` is less than `tolerance()` and the scaled gradient, computed based on the log-likelihood functions provided below, is less than `nrtolerance()`; see [R] [Maximize](#).

The MLE of τ^2 is the value that maximizes the log-likelihood function (Hardy and Thompson 1996)

$$\ln L_{\text{ML}}(\tau^2) = -\frac{1}{2} \left\{ K \ln(2\pi) + \ln |\tau^2 \mathbf{I} + \mathbf{W}^{-1}| + \hat{\boldsymbol{\theta}}' \mathbf{P} \hat{\boldsymbol{\theta}} \right\}$$

The MLE formula for δ is

$$\delta_{\text{MLE}} = \frac{\hat{\boldsymbol{\theta}}' \mathbf{P} \mathbf{P} \hat{\boldsymbol{\theta}} - \text{tr}(\mathbf{W}^*)}{\text{tr}(\mathbf{W}^* \mathbf{W}^*)}$$

The MLE estimator of τ^2 does not incorporate the uncertainty about the unknown regression coefficients $\boldsymbol{\beta}$ and thus can be negatively biased.

The REML estimate of τ^2 is the value that maximizes the restricted log-likelihood function,

$$\ln L_{\text{REML}}(\tau^2) = \ln L_{\text{ML}}(\tau^2) - \frac{1}{2} \ln \left| \sum_{j=1}^K \frac{1}{\tau^2 + \hat{\sigma}_j^2} \mathbf{x}'_j \mathbf{x}_j \right| + \frac{p}{2} \ln(2\pi)$$

and the REML formula for δ is

$$\delta_{\text{REML}} = \frac{\hat{\boldsymbol{\theta}}' \mathbf{P} \mathbf{P} \hat{\boldsymbol{\theta}} - \text{tr}(\mathbf{P})}{\text{tr}(\mathbf{P})}$$

The empirical Bayes estimator for τ^2 was introduced by Morris (1983) and was first used in the meta-analytic context by Berkey et al. (1995). This estimator is also known as the Paule–Mandel estimator (Paule and Mandel 1982). The empirical Bayes formula for δ is

$$\delta_{\text{EB}} = \frac{K/(K-p)\hat{\boldsymbol{\theta}}' \mathbf{P} \hat{\boldsymbol{\theta}} - K}{\text{tr}(\mathbf{W}^*)}$$

For the three above estimators, $\mathbf{A} = \mathbf{W}^*$ in the definition of the \mathbf{P} matrix from (1).

Noniterative methods for computing $\hat{\tau}^2$

This section describes noniterative methods, which have closed-form expressions.

The method-of-moments estimator of τ^2 (DuMouchel and Harris [1983, eq. 3.12]; also see Raudenbush [2009, eq. 16.43]), which can be viewed as an extension of the DerSimonian–Laird estimator from the RE meta-analysis to meta-regression, is

$$\begin{aligned} \hat{\tau}_{\text{DL}}^2 &= \frac{\hat{\boldsymbol{\theta}}' \mathbf{P} \hat{\boldsymbol{\theta}} - (K-p)}{\text{tr}(\mathbf{P})} \\ &= \frac{Q_{\text{res}} - (K-p)}{\text{tr}(\mathbf{W}) - \text{tr}\left\{\mathbf{W}\mathbf{X}(\mathbf{X}'\mathbf{W}\mathbf{X})^{-1}\mathbf{X}'\mathbf{W}\right\}} = \frac{Q_{\text{res}} - (K-p)}{\sum_{j=1}^K w_j (1-h_j)} \end{aligned} \tag{2}$$

where \mathbf{P} is defined in (1) with $\mathbf{A} = \mathbf{W}$, h_j is the j th diagonal element of the “hat” matrix $\mathbf{X}(\mathbf{X}'\mathbf{W}\mathbf{X})^{-1}\mathbf{X}'\mathbf{W}$, and Q_{res} is defined in (3).

For a constant-only model, when $p = 1$, (2) reduces to the DerSimonian–Laird estimator from *Noniterative methods* in [META] meta summarize.

Hedges (1983) used OLS to provide a method of moments estimator of $\hat{\tau}^2$ for the RE meta-analysis. In the context of meta-regression, the extension of the Hedges’s (HE) estimator introduced by Raudenbush (2009, eq. 16.41) is

$$\begin{aligned} \hat{\tau}_{\text{HE}}^2 &= \frac{\hat{\boldsymbol{\theta}}' \mathbf{P} \hat{\boldsymbol{\theta}} - \text{tr}(\mathbf{P}\mathbf{W}^{-1})}{K-p} \\ &= \frac{\sum_{j=1}^K \left(\hat{\theta}_j - \mathbf{x}_j \hat{\boldsymbol{\beta}}_{\text{ols}}\right)^2 - \sum_{j=1}^K \hat{\sigma}_j^2 (1-h_j^{\text{ols}})}{K-p} \end{aligned}$$

where \mathbf{P} is defined in (1) with $\mathbf{A} = \mathbf{I}$, $\hat{\boldsymbol{\beta}}_{\text{ols}} = (\mathbf{X}'\mathbf{X})^{-1}\mathbf{X}'\hat{\boldsymbol{\theta}}$, and h_j^{ols} is the j th diagonal element of the OLS hat matrix $\mathbf{X}(\mathbf{X}'\mathbf{X})^{-1}\mathbf{X}'$.

Sidik and Jonkman (2005) proposed the following estimator. Consider an initial estimate of τ^2 ,

$$\hat{\tau}_0^2 = \frac{\sum_{j=1}^K \left(\hat{\theta}_j - \bar{\theta}\right)^2}{K} \quad \bar{\theta} = \frac{\sum_{j=1}^K \hat{\theta}_j}{K}$$

Then, the estimator is defined as

$$\hat{\tau}_{\text{SJ}}^2 = \frac{\hat{\boldsymbol{\theta}}' \mathbf{P} \hat{\boldsymbol{\theta}}}{K - p} = \frac{\sum_{j=1}^K w_j^{\text{SJ}} \left(\hat{\theta}_j - \mathbf{x}_j \hat{\boldsymbol{\beta}}_{\text{SJ}} \right)^2}{K - p}$$

where $w_j^{\text{SJ}} = \hat{\tau}_0^2 / (\hat{\sigma}_j^2 + \hat{\tau}_0^2)$ is a diagonal element of \mathbf{A} from (1), $\hat{\boldsymbol{\beta}}_{\text{SJ}} = (\mathbf{X}' \mathbf{W}^{\text{SJ}} \mathbf{X})^{-1} \mathbf{X}' \mathbf{W}^{\text{SJ}} \hat{\boldsymbol{\theta}}$, and \mathbf{W}^{SJ} is a $K \times K$ diagonal matrix with elements w_j^{SJ} .

The Sidik–Jonkman estimator is not truncated because, theoretically, it should always produce a nonnegative estimate. However, [Viechtbauer et al. \(2015\)](#) point out that, technically, a negative value can be produced in practice in an unlikely case of all $\hat{\theta}_j$'s being identical.

[Viechtbauer et al. \(2015\)](#) provide the following extension for the estimator of τ^2 , which was originally introduced by [Schmidt and Hunter \(2015\)](#) in the context of RE meta-analysis, to meta-regression

$$\hat{\tau}_{\text{HS}}^2 = \frac{\hat{\boldsymbol{\theta}}' \mathbf{P} \hat{\boldsymbol{\theta}} - K}{\text{tr}(\mathbf{W})} = \frac{Q_{\text{res}} - K}{\text{tr}(\mathbf{W})}$$

where \mathbf{P} is defined in (1) with $\mathbf{A} = \mathbf{W}$.

Knapp–Hartung standard-error adjustment

By default, the inference about the regression coefficients and their confidence intervals from meta-regression is based on a normal distribution. The test of the significance of all regression coefficients is based on a χ^2 distribution with $p - 1$ degrees of freedom.

[Knapp and Hartung \(2003\)](#) proposed an adjustment to the standard errors of the estimated regression coefficients to account for the uncertainty in the estimation of τ^2 . They showed that the corresponding tests of individual regression coefficients and their confidence intervals are based on the Student's t distribution with $K - p$ degrees of freedom and that the overall test of significance is based on an F distribution with $p - 1$ numerator and $K - p$ denominator.

The Knapp–Hartung adjustment first calculates the quadratic form,

$$q_{\text{KH}} = \frac{\hat{\boldsymbol{\theta}}' \mathbf{P} \hat{\boldsymbol{\theta}}}{K - p}$$

where \mathbf{P} is defined in (1) with $\mathbf{A} = \mathbf{W}^*$. It then multiplies the regular expressions of the variances of regression coefficients by q_{KH} or, in the case of the truncated Knapp–Hartung adjustment, by $\max(1, q_{\text{KH}})$.

Residual homogeneity test

Consider a test of residual homogeneity, which mathematically translates to $H_0: \tau^2 = 0$ for the random-effects meta-regression and to $H_0: \phi = 1$ for the fixed-effects meta-regression with multiplicative dispersion parameter ϕ . This test is based on the residual weighted sum of squares, Q_{res} ,

$$Q_{\text{res}} = \sum_{j=1}^K w_j \left(\hat{\theta}_j - \mathbf{x}_j \hat{\boldsymbol{\beta}} \right)^2 = \sum_{j=1}^K \left(\frac{\hat{\theta}_j - \mathbf{x}_j \hat{\boldsymbol{\beta}}}{\hat{\sigma}_j} \right)^2 \quad (3)$$

which is a generalization of the heterogeneity test statistic, Q (see *Homogeneity test* in [META] **meta summarize**), to the context of meta-regression.

Under the null hypothesis of residual homogeneity, Q_{res} follows a χ^2 distribution with $K - p$ degrees of freedom (Seber and Lee 2003, sec. 2.4).

Residual heterogeneity measures

The I_{res}^2 statistic represents the percentage of residual between-study variation relative to the total variability. For an RE meta-regression, it is defined by Higgins and Thompson (2002) as

$$I_{\text{res}}^2 = \frac{\hat{\tau}^2}{\hat{\tau}^2 + s^2} \times 100\%$$

where $s^2 = (K - p)/\text{tr}(\mathbf{P})$ and $\mathbf{A} = \mathbf{W}$ is used to define \mathbf{P} . In the meta-regression context, the H^2 statistic is defined as

$$H_{\text{res}}^2 = \frac{\hat{\tau}^2 + s^2}{s^2}$$

Adjusted R^2 (Harbord and Higgins 2016; Borenstein et al. 2009) measures the proportion of the between-study variance that is explained by the moderators. It is defined as

$$R^2 = \frac{\hat{\tau}_c^2 - \hat{\tau}^2}{\hat{\tau}_c^2} \times 100\% \quad (4)$$

where $\hat{\tau}_c^2$ is the between-study variance estimated from a constant-only model.

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Also see

[META] **meta regress postestimation** — Postestimation tools for meta regress

[META] **meta data** — Declare meta-analysis data

[META] **meta summarize** — Summarize meta-analysis data

[META] **meta** — Introduction to meta

[META] **Glossary**

[META] **Intro** — Introduction to meta-analysis

[U] **20 Estimation and postestimation commands**

[Postestimation commands](#)[Remarks and examples](#)[Also see](#)[predict](#)[Methods and formulas](#)[margins](#)[Reference](#)

Postestimation commands

The following postestimation command is of special interest after `meta regress`:

Command	Description
<code>estat bubbleplot</code>	bubble plots

The following postestimation commands are available after `meta regress`:

Command	Description
<code>contrast</code>	contrasts and ANOVA-style joint tests of estimates
<code>estat summarize</code>	summary statistics for the estimation sample
<code>estat vce</code>	variance–covariance matrix of the estimators (VCE)
<code>estimates</code>	cataloging estimation results
<code>lincom</code>	point estimates, standard errors, testing, and inference for linear combinations of coefficients
<code>margins</code>	marginal means, predictive margins, marginal effects, and average marginal effects
<code>marginsplot</code>	graph the results from margins (profile plots, interaction plots, etc.)
<code>nlcom</code>	point estimates, standard errors, testing, and inference for nonlinear combinations of coefficients
<code>predict</code>	predictions, residuals, and other diagnostic measures
<code>predictnl</code>	point estimates, standard errors, testing, and inference for generalized predictions
<code>pwcompare</code>	pairwise comparisons of estimates
<code>test</code>	Wald tests of simple and composite linear hypotheses
<code>testnl</code>	Wald tests of nonlinear hypotheses

predict

Description for predict

`predict` creates a new variable containing predictions such as linear predictions, residuals, leverage, and standard errors. After random-effects meta-regression, you can also obtain estimates of random effects and their standard errors.

Menu for predict

Statistics > Postestimation

Syntax for predict

Syntax for obtaining BLUPs of random effects and their standard errors after RE meta-regression

```
predict [type] newvar [if] [in], reffects [se(newvar)]
```

Syntax for obtaining other predictions

```
predict [type] newvar [if] [in] [, statistic fixedonly se(sespec)]
```

<i>statistic</i>	Description
Main	
<code>xb</code>	linear prediction; the default
<code>stdp</code>	standard error of the linear prediction
<code>fitted</code>	fitted values, fixed-portion linear prediction plus predicted random effects
<code>residuals</code>	residuals, response minus fitted values
<code>leverage hat</code>	leverage (diagonal elements of hat matrix)

Unstarred statistics are available both in and out of sample; type `predict ... if e(sample) ...` if wanted only for the estimation sample.

Options for predict

Main

`xb`, the default, calculates the linear prediction $\mathbf{x}_j\hat{\beta}$. For the random-effects meta-regression, this corresponds to the fixed portion of the linear predictor based on the estimated regression coefficients. That is, this is equivalent to fixing all random effects in the model to their theoretical mean value of 0.

`stdp` calculates the standard error of the linear prediction.

`reffects` calculates best linear unbiased predictions (BLUPs) of the random effects.

`fitted` calculates the fitted values. With fixed-effects meta-regression or with random-effects meta-regression when option `fixedonly` is also specified, this option is equivalent to `xb`. For random-effects meta-regression without `fixedonly`, it calculates $\mathbf{x}_j\hat{\beta} + u_j$, which is equal to the fixed portion of the linear prediction plus predicted random effects.

`residuals` calculates the residuals, which are equal to the responses minus the fitted values. With fixed-effects meta-regression or with random-effects meta-regression when option `fixedonly` is also specified, it calculates $\hat{\theta}_j - \mathbf{x}_j\hat{\beta}$. The former are known as marginal residuals in the context of the random-effects model. For random-effects meta-regression without `fixedonly`, this option calculates $\hat{\theta}_j - (\mathbf{x}_j\hat{\beta} + u_j)$, which are known as conditional residuals.

`leverage` or `hat` calculates the diagonal elements of the projection (“hat”) matrix.

`fixedonly` specifies that all random effects be set to zero, which is equivalent to using only the fixed portion of the model, when computing results for random-effects models. This option may be specified only with statistics `fitted`, `residuals`, or `leverage`.

`se(newvar[, marginal])` calculates the standard error of the corresponding predicted values. This option may be specified only with statistics `reffects`, `fitted`, and `residuals`.

Suboption `marginal` is allowed only with random-effects meta-regression and requires option `fixedonly`. It computes marginal standard errors, when you type

```
. predict ..., statistic se(newvar, marginal) fixedonly
```

instead of the standard errors conditional on zero random effects, which are computed when you type

```
. predict ..., statistic se(newvar) fixedonly
```

`marginal` is not allowed in combination with `reffects`.

margins

Description for margins

`margins` estimates margins of response for linear predictions.

Menu for margins

Statistics > Postestimation

Syntax for margins

```
margins [marginlist] [, options]
```

```
margins [marginlist] , predict(statistic ...) [options]
```

<i>statistic</i>	Description
<code>xb</code>	linear prediction; the default
<code>fitted</code>	fitted values; implies <code>fixedonly</code>
<code>stdp</code>	not allowed with <code>margins</code>
<code>reffects</code>	not allowed with <code>margins</code>
<code>residuals</code>	not allowed with <code>margins</code>
<code>leverage</code> <code>hat</code>	not allowed with <code>margins</code>

Statistics not allowed with `margins` are functions of stochastic quantities other than $e(b)$.

For the full syntax, see [R] [margins](#).

Remarks and examples

We demonstrate some of the postestimation features, including `estat bubbleplot`, `margins`, and `predict` after `meta regress`.

► Example 1: Bubble plot

Consider the declared `BCG dataset` of clinical trials that studied the efficacy of a Bacillus Calmette-Guérin (BCG) vaccine in the prevention of tuberculosis (TB) (Colditz et al. 1994). In [example 1 of \[META\] meta regress](#), we used `meta regress` to fit a simple meta-regression to these data with the continuous moderator `latitude_c` to explore heterogeneity.

```
. use https://www.stata-press.com/data/r16/bcgset
(Efficacy of BCG vaccine against tuberculosis; set with -meta esize-)
. meta regress latitude_c

Effect-size label:  Log Risk-Ratio
Effect size:       _meta_es
Std. Err.:        _meta_se

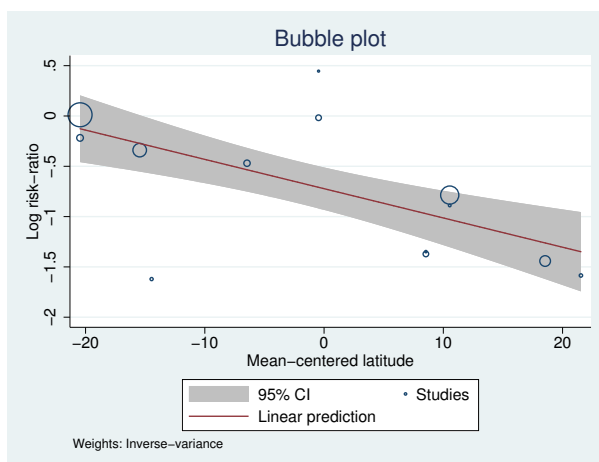
Random-effects meta-regression                Number of obs =      13
Method: REML                                Residual heterogeneity:
                                           tau2 = .07635
                                           I2 (%) = 68.39
                                           H2 = 3.16
                                           R-squared (%) = 75.63
                                           Wald chi2(1) = 16.36
                                           Prob > chi2 = 0.0001
```

_meta_es	Coef.	Std. Err.	z	P> z	[95% Conf. Interval]
latitude_c	-.0291017	.0071953	-4.04	0.000	-.0432043 -.0149991
_cons	-.7223204	.1076535	-6.71	0.000	-.9333174 -.5113234

Test of residual homogeneity: $Q_{res} = \chi^2(11) = 30.73$ Prob > $Q_{res} = 0.0012$

Whenever there is one continuous moderator in a meta-regression, a so-called bubble plot is commonly used to explore the relationship between the effect size and that moderator. Let's use `estat bubbleplot` to produce the bubble plot after the fitted meta-regression.

```
. estat bubbleplot
```



A bubble plot is a scatterplot of the observed effect sizes against the moderator overlaid with the predicted regression and confidence-intervals lines. Each study is represented by a circle (bubble) with the size (area) proportional to the study precision, $1/\hat{\sigma}_j^2$. The larger the size of the bubble, the more precise the study. The coordinates of the center of each circle show the observed value of the effect size on the y axis and that of the moderator (`latitude_c` in our example) on the x axis. The solid line shows the predicted values (predicted log risk-ratios in our example). The predicted 95% confidence intervals are also plotted.

From the plot, the log risk-ratio for the BCG vaccine declines as the distance from the equator increases. There appear to be a couple of outlying studies (see points in the bottom left and middle top sections of the plot), but their bubbles are very small, which suggests that their log risk-ratios estimates had small weights, relative to other studies, in the meta-regression. Outlying studies with large bubbles may be a source of concern because of the large differences in their effect sizes compared with those from the other studies and because of the large impact they have on the regression results. ◀

▷ Example 2: Marginal effects

Continuing with [example 1](#), we found that the log risk-ratio for the BCG decreases as the distance from the equator increases. For example, from the bubble plot, a trial conducted relatively close to the equator, say, in Thailand (with a latitude of 15 or a centered latitude of -18.5), would have a predicted log risk-ratio of about -0.2 . A trial conducted in, say, Nepal (with a latitude of 28 or a centered latitude of -5.5), would have a predicted log risk-ratio of about -0.7 . And a trial conducted in, say, Ukraine (with a latitude of 50 or a centered latitude of 16.5), would have a predicted log risk-ratio of about -1 .

Instead of relying on the graph, we can obtain more precise estimates of the predicted log risk-ratios at different latitude values by using the `margins` command as follows:

```
. margins, at(latitude_c = (-18.5 -5.5 16.5))
Adjusted predictions          Number of obs      =          13
Expression   : Fitted values; fixed portion (xb), predict(fitted fixedonly)
1._at        : latitude_c      =          -18.5
2._at        : latitude_c      =           -5.5
3._at        : latitude_c      =          16.5
```

	Delta-method				[95% Conf. Interval]	
	Margin	Std. Err.	z	P> z		
_at						
1	-.1839386	.1586092	-1.16	0.246	-.4948069	.1269297
2	-.562261	.1091839	-5.15	0.000	-.7762574	-.3482645
3	-1.202499	.1714274	-7.01	0.000	-1.53849	-.8665072

The list of numbers specified in the `at()` option are the values of the latitudes centered around the latitude mean (≈ 33.5).

Note that results produced by `margins` are on the log scale and need to be exponentiated to make interpretations on the natural (risk) scale. For instance, from the output, the risk ratio for regions with `latitude_c = 16.5` is $\exp(-1.202499) = 0.3$, which means that the vaccine is expected to reduce the risk of TB by 70% for regions with that latitude.

► Example 3: Predicted random effects

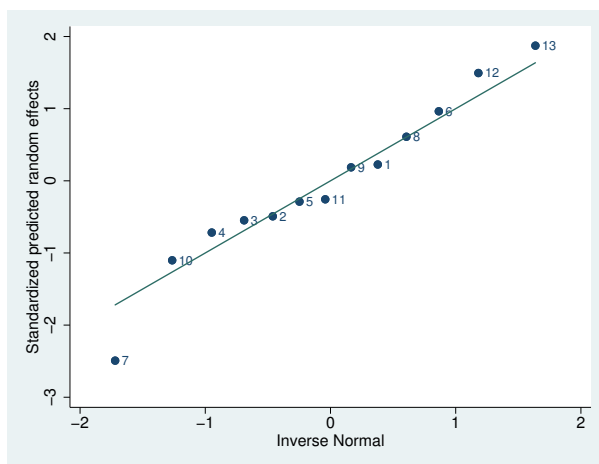
In [example 1](#), we noticed a couple of outlying studies. Let's explore this further by looking at predicted random effects from our random-effects meta-regression.

We first use `predict` with options `reffects` and `se()` to predict the random-effects and estimate their standard errors.

```
. predict double u, reffects se(se_u)
```

Then, we generate a new variable, `ustandard`, as the ratio of the predicted random effects to their standard errors and use the `qnorm` command (see [\[R\] Diagnostic plots](#)) to construct the normal quantile plot.

```
. generate double ustandard = u/se_u
. label variable ustandard "Standardized predicted random effects"
. qnorm ustandard, mlabel(trial)
```



The plot suggests that trial 7, labeled “Vandiviere et al., 1973” in our data, is an outlier. From the data, the log risk-ratio estimate for this trial is -1.62 with the corresponding risk-ratio estimate of about 0.2. This means that, in that trial, the vaccine reduced the risk of TB by roughly 80% even though this trial was conducted relatively close to the equator (in Haiti, with `latitude=19`). In fact, this trial reported the largest risk reduction (smallest log-risk-ratio value) in the meta-analysis. Compare this with trial 11 (“Comstock et al., 1974”), which was conducted in Puerto Rico and has a similar latitude (`latitude=18`) but whose estimated risk reduction was much more moderate, about 29% (with the risk-ratio estimate of $\exp(-0.34) = 0.71$). More investigation is needed to explain the extreme value reported by trial 7. Thus, in this example, you may consider reporting the results of meta-analyses with and without this trial.

◀

Methods and formulas

Methods and formulas are presented under the following headings:

[Random-effects meta-regression](#)
[Fixed-effects meta-regression](#)

The following formulas are used by `predict`. The notation is based on *Methods and formulas of [META] meta regress*.

Random-effects meta-regression

The fixed-portion of the linear prediction (option `xb`) is $\mathbf{x}_j \widehat{\boldsymbol{\beta}}$. The estimated standard error of the fixed-portion of the linear prediction (option `stdp`) for study j is

$$\widehat{\text{SE}}(\mathbf{x}_j \widehat{\boldsymbol{\beta}}) = \sqrt{\mathbf{x}_j (\mathbf{X}'\mathbf{W}^*\mathbf{X})^{-1} \mathbf{x}_j'}$$

The BLUP of the j th random effect (option `reffects`) is

$$\widehat{u}_j = \lambda_j (\widehat{\theta}_j - \mathbf{x}_j \widehat{\boldsymbol{\beta}})$$

where

$$\lambda_j = \frac{\widehat{\tau}^2}{\widehat{\tau}^2 + \widehat{\sigma}_j^2}$$

is the empirical Bayes shrinkage factor for the j th study. When the `se()` option is also specified, the estimated standard error of \widehat{u}_j is

$$\widehat{\text{SE}}(\widehat{u}_j) = \lambda_j \sqrt{\widehat{\sigma}_j^2 + \widehat{\tau}^2 - \mathbf{x}_j (\mathbf{X}'\mathbf{W}^*\mathbf{X})^{-1} \mathbf{x}_j'}$$

The fitted value (option `fitted`) is

$$\widetilde{\theta}_j = \mathbf{x}_j \widehat{\boldsymbol{\beta}} + \widehat{u}_j$$

When the `se()` option is also specified, the estimated standard error of $\widetilde{\theta}_j$ is

$$\widehat{\text{SE}}(\widetilde{\theta}_j) = \sqrt{\lambda_j^2 (\widehat{\sigma}_j^2 + \widehat{\tau}^2) + (1 - \lambda_j^2) \mathbf{x}_j (\mathbf{X}'\mathbf{W}^*\mathbf{X})^{-1} \mathbf{x}_j'}$$

The residual (option `residuals`) is

$$e_j = \widehat{\theta}_j - \widetilde{\theta}_j$$

When option `se()` is also specified, the estimated standard error of e_j is

$$\widehat{\text{SE}}(e_j) = \sqrt{(1 + \lambda_j^2) (\widehat{\sigma}_j^2 + \widehat{\tau}^2 - \mathbf{x}_j (\mathbf{X}'\mathbf{W}^*\mathbf{X})^{-1} \mathbf{x}_j')}$$

The leverage (option `hat`) are the diagonal elements of the hat matrix $\mathbf{X} (\mathbf{X}'\mathbf{W}^*\mathbf{X})^{-1} \mathbf{X}'\mathbf{W}^*$:

$$h_j^* = \frac{1}{\widehat{\tau}^2 + \widehat{\sigma}_j^2} \mathbf{x}_j (\mathbf{X}'\mathbf{W}^*\mathbf{X})^{-1} \mathbf{x}_j'$$

When the `fixedonly` option is specified, the formulas for the fitted values and residuals (including their standard errors) and leverage are adjusted by replacing the value of \hat{u}_j with 0, in which case, $\hat{\tau}^2 = 0$, $\lambda_j = 0$, and \mathbf{W}^* is replaced with $\mathbf{W} = \text{diag}(1/\hat{\sigma}_1^2, \dots, 1/\hat{\sigma}_K^2)$. In this case, the standard errors are computed conditionally on zero random effects.

If `se()`'s option `marginal` is specified, then marginal standard errors are computed. This is equivalent to computing $\widehat{\text{SE}}(\tilde{\theta}_j)$ and $\widehat{\text{SE}}(e_j)$ with $\lambda_j = 0$ but keeping $\hat{\tau}^2$ and \mathbf{W}^* unchanged.

Fixed-effects meta-regression

The linear prediction (option `xb`) is $\mathbf{x}_j \hat{\beta}$. The estimated standard error of the linear prediction (option `stdp`) for study j is

$$\widehat{\text{SE}}(\mathbf{x}_j \hat{\beta}) = \sqrt{\mathbf{x}_j (\mathbf{X}' \mathbf{W} \mathbf{X})^{-1} \mathbf{x}_j'}$$

The fitted value (option `fitted`) is the same as the linear prediction:

$$\tilde{\theta}_j = \mathbf{x}_j \hat{\beta}$$

The residual (option `residuals`) is

$$e_j = \hat{\theta}_j - \tilde{\theta}_j$$

When option `se()` is also specified, the estimated standard error of e_j is

$$\widehat{\text{SE}}(e_j) = \sqrt{(\hat{\sigma}_j^2 - \mathbf{x}_j (\mathbf{X}' \mathbf{W} \mathbf{X})^{-1} \mathbf{x}_j')}$$

The leverage (option `hat`) are the diagonal elements of the hat matrix $\mathbf{X} (\mathbf{X}' \mathbf{W} \mathbf{X})^{-1} \mathbf{X}' \mathbf{W}$:

$$h_j = \frac{1}{\hat{\sigma}_j^2} \mathbf{x}_j (\mathbf{X}' \mathbf{W} \mathbf{X})^{-1} \mathbf{x}_j'$$

For the multiplicative fixed-effects meta-regression, in the above formulas, replace \mathbf{W} with \mathbf{W}^ϕ and $\hat{\sigma}_j^2$ with $\hat{\phi} \hat{\sigma}_j^2$, where $\hat{\phi}$ is defined in *Fixed-effects meta-regression* in [META] meta regress.

Reference

- Colditz, G. A., T. F. Brewer, C. S. Berkey, M. E. Wilson, E. Burdick, H. V. Fineberg, and F. Mosteller. 1994. Efficacy of BCG vaccine in the prevention of tuberculosis: Meta-analysis of the published literature. *Journal of the American Medical Association* 271: 698–702.

Also see

[META] [meta regress](#) — Meta-analysis regression

[META] [meta data](#) — Declare meta-analysis data

[META] [meta](#) — Introduction to meta

[META] [Glossary](#)

[META] [Intro](#) — Introduction to meta-analysis

[U] [20 Estimation and postestimation commands](#)

Title

estat bubbleplot — Bubble plots after meta regress

[Description](#)
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Description

`estat bubbleplot` produces bubble plots after simple meta-regression with one continuous moderator performed by using [meta regress](#). The bubble plot is a scatterplot of effect sizes against a [moderator](#) of interest overlaid with the predicted regression line and confidence-interval bands. In a bubble plot, the marker sizes, “bubbles”, are proportional to study weights.

Quick start

Fit a random-effects meta-regression with a continuous moderator, `x`

```
meta regress x, random
```

Construct a bubble plot for `x`

```
estat bubbleplot
```

As above, but specify that the size of the marker representing studies be proportional to the random-effects weights instead of the default fixed-effects weights

```
estat bubbleplot, reweighted
```

Construct a bubble plot with a 90% confidence interval

```
estat bubbleplot, level(90)
```

Menu

Statistics > Meta-analysis

Syntax

```
estat bubbleplot [if] [in] [, options]
```

<i>options</i>	Description
Main	
<code>reweighted</code>	make bubble size depend on random-effects weights
<code>[no]regline</code>	display or suppress the regression line
<code>[no]ci</code>	display or suppress the confidence intervals
<code>level(#)</code>	set confidence level; default is as declared for meta-analysis
<code>n(#)</code>	evaluate CI lines at # points; default is n(100)
Fitted line	
<code>lineopts(<i>line_options</i>)</code>	affect rendition of the plotted regression line
CI plot	
<code>ciopts(<i>ciopts</i>)</code>	affect rendition of the plotted CI band
Add plots	
<code>addplot(<i>plot</i>)</code>	add other plots to the bubble plot
Y axis, X axis, Titles, Legend, Overall	
<code>twoway_options</code>	any options other than by() documented in [G-3] twoway_options

Options

Main

`reweighted` is used with random-effects meta-regression. It specifies that the sizes of the bubbles be proportional to the weights from the random-effects meta-regression, $w_j^* = 1/(\hat{\sigma}_j^2 + \hat{\tau}^2)$. By default, the sizes are proportional to the precision of each study, $w_j = 1/\hat{\sigma}_j^2$.

`regline` and `noregline` display or suppress the rendition of the regression line. The default, `regline`, is to display the regression line. Option `noregline` implies option `noci`.

`ci` and `noci` display or suppress confidence intervals. The default, `ci`, is to display them.

`level(#)` specifies the confidence level, as a percentage, for confidence intervals. The default is as declared for the meta-analysis session; see [Declaring a confidence level for meta-analysis in \[META\] meta data](#). Also see option `level()` in [\[META\] meta set](#).

`n(#)` specifies the number of points at which to evaluate the CIs. The default is n(100).

Fitted line

`lineopts(line_options)` affects the rendition of the plotted regression line; see [\[G-3\] line_options](#).

CI plot

`ciopts(ciopts)` affects the rendition of the CI band in the bubble plot. *ciopts* are any options as defined in [\[G-2\] graph twoway rline](#) and option `recast(rarea)` as described in [\[G-3\] advanced_options](#).

Add plots

`addplot(plot)` allows adding more graph twoway plots to the graph; see [G-3] [addplot_option](#).

Y axis, X axis, Titles, Legend, Overall

`twoway_options` are any of the options documented in [G-3] [twoway_options](#), excluding `by()`. These include options for titling the graph (see [G-3] [title_options](#)) and for saving the graph to disk (see [G-3] [saving_option](#)).

Remarks and examples

Remarks are presented under the following headings:

[Introduction](#)

[Examples of using estat bubbleplot](#)

Introduction

A bubble plot (Berkey et al. 1995; Thompson and Sharp 1999; Thompson and Higgins 2002) is used after simple meta-regression with a continuous moderator to describe the relation between the effect size and the corresponding moderator. It is used as a tool to assess how well the regression model fits the data and to potentially identify influential and outlying studies. The bubble plot is a scatterplot with the study-specific effect sizes plotted on the y axis and the moderator of interest from the meta-regression plotted on the x axis. The sizes of the markers or “bubbles” are proportional to the precision of each study. The more precise (larger) the study, the larger the size of the bubble. The predicted regression line and confidence bands are overlaid with the scatterplot.

`estat bubbleplot` produces bubble plots after simple meta-regression with a continuous moderator performed by using `meta regress`. Traditionally, the weights used to determine the sizes of the bubbles are the inverses of the effect-size variances, $1/\hat{\sigma}_j^2$. After a random-effects meta-regression, you can specify the `reweighted` option to instead use the random-effects weights, $1/(\hat{\sigma}_j^2 + \hat{\tau}^2)$.

The predicted regression line and the 95% confidence intervals are plotted by default. You can specify the `level()` option to obtain other confidence intervals. You can control the look of the lines by specifying the options `lineopts()` and `ciopts()`. You can also suppress the lines by specifying the options `noregline` and `noci`.

Examples of using estat bubbleplot

In the examples that follow, we demonstrate how to create and customize bubble plots after a meta-regression. Consider the BCG dataset from [Examples of using meta regress](#) in [META] [meta regress](#).

```
. use https://www.stata-press.com/data/r16/bcgset
(Efficacy of BCG vaccine against tuberculosis; set with -meta esize-)
. meta query, short
-> meta esize npost nnegt nposc nnegc, esize(lnrratio) studylabel(studylbl)

Effect-size label:  Log Risk-Ratio
Effect-size type:   lnrratio
Effect size:       _meta_es
Std. Err.:         _meta_se
Model:             Random-effects
Method:            REML
```

To create these plots, we first fit the random-effects meta-regression shown in [example 1](#) of [\[META\] meta regress](#), but our focus here is not on the interpretation of these plots but on the variety of bubble plots that can be created.

```
. meta regress latitude_c
      Effect-size label: Log Risk-Ratio
      Effect size:      _meta_es
      Std. Err.:       _meta_se

Random-effects meta-regression
Method: REML
Number of obs =      13
Residual heterogeneity:
      tau2 = .07635
      I2 (%) = 68.39
      H2 = 3.16
R-squared (%) = 75.63
Wald chi2(1) = 16.36
Prob > chi2 = 0.0001
```

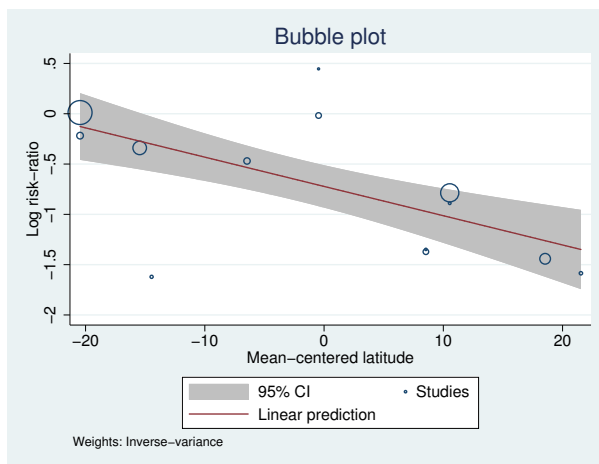
_meta_es	Coef.	Std. Err.	z	P> z	[95% Conf. Interval]	
latitude_c	-.0291017	.0071953	-4.04	0.000	-.0432043	-.0149991
_cons	-.7223204	.1076535	-6.71	0.000	-.9333174	-.5113234

Test of residual homogeneity: $Q_{res} = \chi^2(11) = 30.73$ Prob > $Q_{res} = 0.0012$

► Example 1: A basic bubble plot

To construct a bubble plot after performing a [simple meta-regression](#), we simply type

```
. estat bubbleplot
```

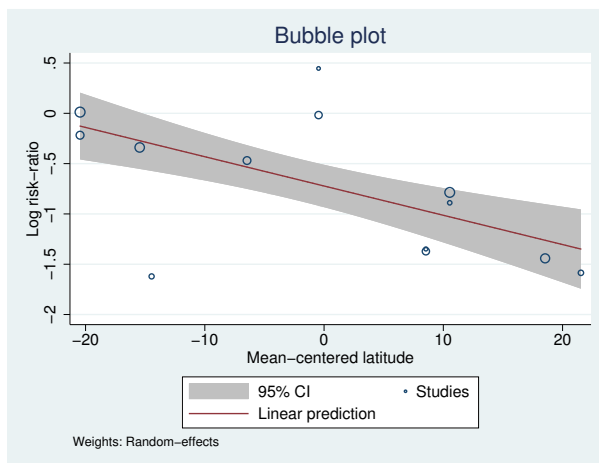


The graph shows the log risk-ratios plotted against the mean-centered latitudes of the studies' locations. By default, the regression line and corresponding confidence intervals are plotted. We could suppress these and plot just the bubbles with options `noregline` and `noci`, respectively. The regression line provides a good fit of the data because most studies are relatively close to it. The log risk-ratios for the BCG vaccine decline with increasing latitude. For more interpretation of the above bubble plot, refer to [example 1](#) of [\[META\] meta regress postestimation](#).

▷ Example 2: Reweighting the bubbles

By default, the bubble sizes are proportional to trial precisions, $1/\hat{\sigma}_j^2$. With the `reweighted` option, we can make the bubble sizes proportional to the weights from the random-effects meta-regression, $1/(\hat{\sigma}_j^2 + \hat{\tau}^2)$. For example, continuing with [example 1](#), we can reweight the bubbles as follows:

```
. estat bubbleplot, reweighted
```

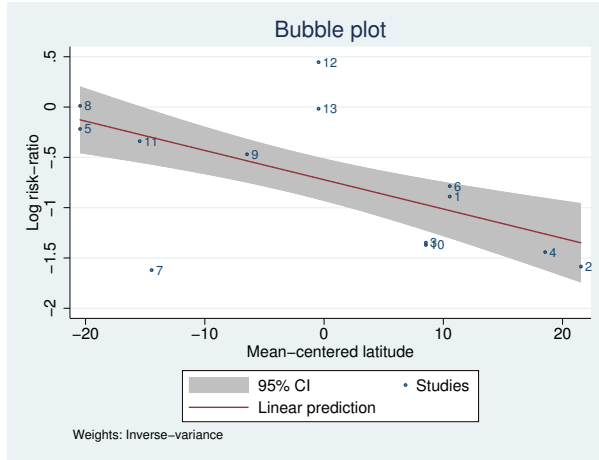


With random-effects weights, the sizes of the bubbles are more uniform across the studies as compared with precision (fixed-effects) weights used in [example 1](#). This will always be true except when $\hat{\tau}^2 = 0$, in which case the bubble sizes will be identical with both types of weights.

▷ Example 3: Using addplot() to add labels for the trials

Below, we discuss how you can add labels to the trials, which are represented by the hollow circles on a bubble plot. Typically, we use `mlabel(varname)` to add marker labels. For example, if we wish to label the trials according to their trial ID, `trial`, we type

```
. estat bubbleplot, mlabel(trial)
```



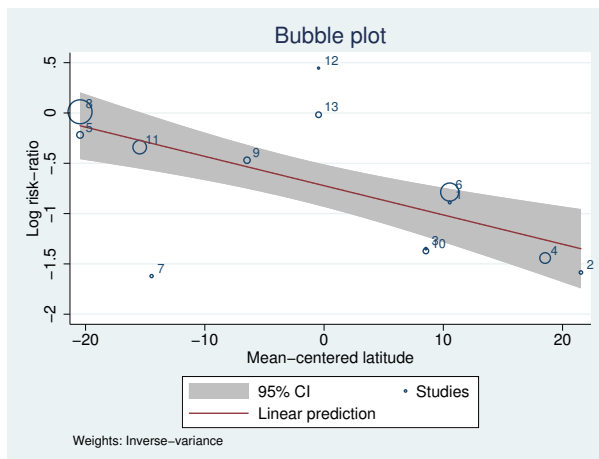
Specifying the `mlabel()` option causes all the markers to have the same size. One way to get around this is by using the `addplot()` option.

We can use `addplot()` to overlay an exact copy of the properly weighted bubble plot but without plotting the markers and symbols, that is, using the `msymbol(none)` option. We can then add labels to these nonplotted symbols to obtain the desired plot. Here is our minimal `addplot()` specification:

```
addplot(scatter _meta_es latitude_c, msymbol(none) mlabel(trial))
```

The full specification is

```
. local opts msymbol(none) mlabel(trial) mlabcolor(navy) legend(order(1 2 3))
. estat bubbleplot, addplot(scatter _meta_es latitude_c, 'opts' mlabpos(2))
```



We used additional options to fine-tune the bubble plot. The `mllabcolor(navy)` option controls the color of the study labels (navy here). The `legend(order(1 2 3))` option prevents the display of a legend key for the added scatterplot in the legend box at the bottom of the plot. Finally, the `mllabpos(2)` option specifies that marker labels be drawn at the 2 o'clock position.

◀

► Example 4: Adjusting label positions

Continuing with [example 3](#), let's customize the labels further. For example, marker labels 3 and 10 (and 6 and 1) are not easily distinguishable. You may provide individual marker label positions for each study by using the `mllabvpos(varname)` option. `varname` must be created to hold the individual positions (an integer number between 0 to 12) of each marker.

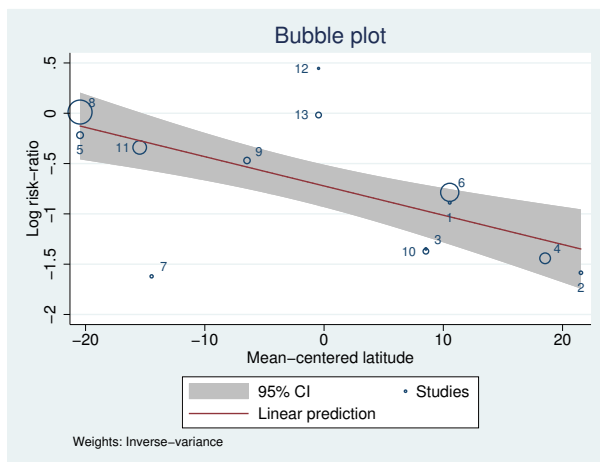
Let's generate our position variable.

```
. generate byte pos = 2
. quietly replace pos = 9 in 10/13
. quietly replace pos = 6 if inlist(trial,1,2,5)
```

We generated a new variable `pos` to hold the individual positions of each marker label. We chose to draw labels at 9 o'clock for trials 10 to 13, at 6 o'clock for trials 1, 2, and 5, and at 2 o'clock for the other trials.

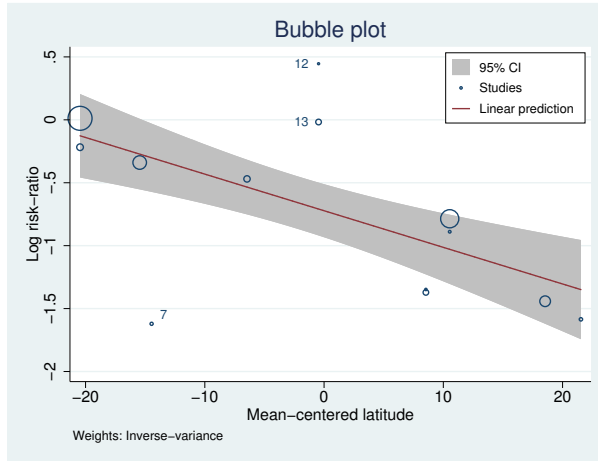
We now use a similar specification of `addplot()` from [example 3](#), but here we add `mllabvpos(pos)` and `mllabgap(*2)` to double the gap size between the marker labels and the markers so that the trial labels do not touch the hollow circles; see trials 6 and 8.

```
. estat bubbleplot, addplot(scatter _meta_es latitude_c, mllabvpos(pos)
> mllabgap(*2) 'opts')
```



We can modify other aspects of the graph such as the legend. Let's place the legend inside the plot region. We also specify if `inlist(trial,7,12,13)` with `estat bubbleplot` to display trial labels only for specific trials (for example, outliers, trials with large weights, and so on).

```
. local legopts legend(ring(0) position(2) cols(1) size(small) symxsize(*0.3))
. estat bubbleplot, addplot(scatter _meta_es latitude_c
> if inlist(trial,7,12,13), mlabvpos(pos) mlabgap(*2) 'opts') 'legopts'
```



Within the `legend()` option (see [G-3] [legend_options](#)), `ring(0)` and `position(2)` specify that the legend be placed inside the plot region at the 2 o'clock position. `cols(1)` specifies that all legend keys be displayed in one column. `size(small)` specifies that a small font be used for the legend key text, and `symxsize(*0.3)` sets the width of the key symbols to 30% of their default width.

◀

Methods and formulas

`estat bubbleplot` produces a scatterplot with the effect sizes, $\hat{\theta}_j$, stored in the system variable `_meta_es` on the y axis and a moderator from the meta-regression on the x axis. By default, the bubble size is proportional to $w_j = 1/\hat{\sigma}_j^2$. For a random-effects meta-regression, if you specify the `reweighted` option, the weights $w_j^* = 1/(\hat{\sigma}_j^2 + \hat{\tau}^2)$ will be used.

For a simple meta-regression with moderator x_1 , the plotted predicted line is $\hat{\beta}_0 + \hat{\beta}_1 x_1 = \mathbf{x}_j \hat{\beta}$. The CIs are computed as

$$\mathbf{x}_j \hat{\beta} \pm z_{1-\alpha/2} \widehat{SE}(\mathbf{x}_j \hat{\beta})$$

where the computation of $\widehat{SE}(\mathbf{x}_j \hat{\beta})$ is described in [META] [meta regress postestimation](#). The `n()` option specifies how many evaluation points are used to construct the CI plots. By default, 100 points are used. When the `se()` or `tdistribution` option is specified with `meta regress`, the confidence intervals use the $t_{K-2, 1-\alpha/2}$ critical value.

References

- Berkey, C. S., D. C. Hoaglin, F. Mosteller, and G. A. Colditz. 1995. A random-effects regression model for meta-analysis. *Statistics in Medicine* 14: 395–411.
- Thompson, S. G., and J. P. T. Higgins. 2002. How should meta-regression analyses be undertaken and interpreted? *Statistics in Medicine* 21: 1559–1573.
- Thompson, S. G., and S. J. Sharp. 1999. Explaining heterogeneity in meta-analysis: A comparison of methods. *Statistics in Medicine* 18: 2693–2708.

Also see

- [META] [meta regress](#) — Meta-analysis regression
- [META] [meta regress postestimation](#) — Postestimation tools for meta regress
- [META] [meta](#) — Introduction to meta
- [META] [Glossary](#)
- [META] [Intro](#) — Introduction to meta-analysis

Title

meta funnelplot — Funnel plots

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Description

`meta funnelplot` produces funnel plots, which are used to explore the presence of [small-study effects](#) often associated with publication bias. A funnel plot is a scatterplot of study-specific effect sizes on the x axis against measures of study precision such as standard errors and inverse standard errors on the y axis. In the absence of small-study effects, the plot should look symmetrical. `meta funnelplot` can also draw contour-enhanced funnel plots, which are useful for investigating whether the plot asymmetry can be attributed to publication bias.

Quick start

Construct a funnel plot for `meta data`, which was declared by either `meta set` or `meta esize`

```
meta funnelplot
```

Specify 1%, 5%, and 10% significance contours to produce a contour-enhanced funnel plot

```
meta funnelplot, contours(1 5 10)
```

As above, but base the significance contours on a one-sided lower-tailed z test, and request separate plots for each group of variable `groupvar`

```
meta funnelplot, contours(1 5 10, lower) by(groupvar)
```

Specify the inverse standard error as the precision metric on the y axis

```
meta funnelplot, metric(invse)
```

Menu

Statistics > Meta-analysis

Syntax

Construct a funnel plot

```
meta funnelplot [if] [in] [, level(#) options]
```

Construct a contour-enhanced funnel plot

```
meta funnelplot [if] [in], contours(contourspec) [options]
```

options

Description

<i>options</i>	Description
Model	
random [<i>remethod</i>]	random-effects meta-analysis
common [<i>cefemethod</i>]	common-effect meta-analysis
fixed [<i>cefemethod</i>]	fixed-effects meta-analysis
Options	
by (<i>varlist</i> , ...)	construct a separate plot for each group formed by <i>varlist</i>
metric (<i>metric</i>)	specify <i>y</i> -axis metric; default is metric (<i>se</i>)
n (#)	evaluate CI lines or significance contours at # points; default is n (300)
[<i>no</i>] metashow	display or suppress meta settings in the output
graph_options	affect rendition of overall funnel plot

remethod

Description

reml	restricted maximum likelihood; the default
mle	maximum likelihood
ebayes	empirical Bayes
dlaird	DerSimonian–Laird
sjonkman	Sidik–Jonkman
hedges	Hedges
hschmidt	Hunter–Schmidt

cefemethod

Description

mhaenszel	Mantel–Haenszel
invvariance	inverse variance
ivariance	synonym for invvariance

<i>graph_options</i>	Description
ES line <code>esopts</code> (<i>line_options</i>)	affect rendition of estimated effect-size line
CI plot * <code>ciopts</code> (<i>ciopts</i>)	affect rendition of the confidence intervals
Add plots <code>addplot</code> (<i>plot</i>)	add other plots to the funnel plot
Y axis, X axis, Titles, Legend, Overall <i>twoway_options</i>	any options documented in [G-3] <i>twoway_options</i>

* `ciopts`(*ciopts*) is not available for a contour-enhanced funnel plot.

Options

Main

`contours`(*contourspec*) specifies that a contour-enhanced funnel plot be plotted instead of the default standard funnel plot; see *Contour-enhanced funnel plots*. This option may not be combined with options `ciopts`() and `level`().

contourspec is `numlist`[, lower upper lines *graph_options*]. *numlist* specifies the levels of significance (as percentages) and may contain no more than 8 integer values between 1 and 50.

`lower` and `upper` specify that the significance contours be based on one-sided lower- or upper-tailed z tests of individual effect sizes. In other words, the studies in the shaded area of a specific contour c are considered not statistically significant based on one-sided lower- or upper-tailed z tests with $\alpha = c/100$. By default, the contours correspond to the two-sided z tests.

`lines` specifies that only the contours lines be plotted. That is, no shaded regions will be displayed.

graph_options are any of the options documented in [G-3] *area_options* except `recast`() or, if option `lines` is specified, any of the options documented in [G-3] *line_options*.

Model

Options `random`(), `common`(), and `fixed`() specify a meta-analysis model to use when estimating the overall effect size. For historical reasons, the default is `common`(*invariance*), regardless of the global model declared by `meta set` or `meta esize`. Specify one of these options with `meta funnelplot` to override this default. Options `random`(), `common`(), and `fixed`() may not be combined. Also see *Meta-analysis models* in [META] **Intro**.

`random` and `random`(*remethod*) specify that a random-effects model be assumed for meta-analysis; see *Random-effects model* in [META] **Intro**.

remethod specifies the type of estimator for the between-study variance τ^2 . *remethod* is one of `reml`, `mle`, `ebayes`, `dlaird`, `sjonkman`, `hedges`, or `hschmidt`. `random` is a synonym for `random`(`reml`). See *Options* in [META] **meta esize** for more information.

`common` and `common`(*cefemethod*) specify that a common-effect model be assumed for meta-analysis; see *Common-effect (“fixed-effect”) model* in [META] **Intro**. Also see the *discussion* in [META] **meta data** about common-effect versus fixed-effects models.

common implies common(mhaenszel) for effect sizes lnratio, lnrratio, and rdiff and common(invvariance) for all other effect sizes. common(mhaenszel) is supported only with effect sizes lnratio, lnrratio, and rdiff.

cefemethod is one of mhaenszel or invvariance (synonym ivariance). See [Options](#) in [META] [meta esize](#) for more information.

fixed and fixed(cefemethod) specify that a fixed-effects model be assumed for meta-analysis; see [Fixed-effects model](#) in [META] [Intro](#). Also see the [discussion](#) in [META] [meta data](#) about fixed-effects versus common-effect models.

fixed implies fixed(mhaenszel) for effect sizes lnratio, lnrratio, and rdiff and fixed(invvariance) for all other effect sizes. fixed(mhaenszel) is supported only with effect sizes lnratio, lnrratio, and rdiff.

cefemethod is one of mhaenszel or invvariance (synonym ivariance); see [Options](#) in [META] [meta esize](#) for more information.

Options

by(*varlist* [, *byopts*]) specifies that a separate plot for each group defined by *varlist* be produced. *byopts* are any of the options documented in [G-3] [by_option](#). by() is useful to explore publication bias in the presence of between-study heterogeneity induced by a set of categorical variables. These variables must then be specified in the by() option.

metric(*metric*) specifies the precision metric on the *y* axis. *metric* is one of se, invse, var, invvar, n, or invn. When *metric* is one of n or invn, no CIs or significance contours are plotted. The default is metric(se).

se specifies that the standard error, $\hat{\sigma}_j$, be used as the precision metric.

invse specifies that the inverse of the standard error, $1/\hat{\sigma}_j$, be used as the precision metric.

var specifies that the variance, $\hat{\sigma}_j^2$, be used as the precision metric.

invvar specifies that the inverse of the variance, $1/\hat{\sigma}_j^2$, be used as the precision metric.

n specifies that the sample size, n_j , be used as the precision metric.

invn specifies that the inverse of the sample size, $1/n_j$, be used as the precision metric.

level(#) specifies the confidence level, as a percentage, for confidence intervals. The default is as declared for the meta-analysis session; see [Declaring a confidence level for meta-analysis](#) in [META] [meta data](#). Also see option level() in [META] [meta set](#). This option may not be combined with option contours().

n(#) specifies the number of points at which to evaluate the CIs or, if option contours() is specified, significance contours. The default is n(300).

metashow and nometashow display or suppress the meta setting information. By default, this information is displayed at the top of the output. You can also specify nometashow with [meta update](#) to suppress the meta setting output for the entire meta-analysis session.

ES line

esopts(*line_options*) affects the rendition of the line that plots the estimated overall effect size; see [G-3] [line_options](#).

CI plot

`ciopts`(*ciopts*) affects the rendition of the (pseudo) CI lines in a funnel plot. *ciopts* are any of the options documented in [G-3] [line_options](#) and option `recast(rarea)` as described in [G-3] [advanced_options](#). This option may not be combined with option `contours()`.

Add plots

`addplot(plot)` allows adding more `graph twoway` plots to the graph; see [G-3] [addplot_option](#).

Y axis, X axis, Titles, Legend, Overall

twoway_options are any of the options documented in [G-3] [twoway_options](#). These include options for titling the graph (see [G-3] [title_options](#)) and for saving the graph to disk (see [G-3] [saving_option](#)).

Remarks and examples

Remarks are presented under the following headings:

- [Introduction](#)
- [Funnel plots](#)
- [Contour-enhanced funnel plots](#)
- [Using meta funnelplot](#)
- [Examples of using meta funnelplot](#)

Introduction

A funnel plot is used to visually explore “small-study effects”. The term small-study effects (Sterne, Gavaghan, and Egger 2000) is used in meta-analysis to describe the cases when the results of smaller studies differ systematically from the results of larger studies. For instance, smaller studies often report larger effect sizes than the larger studies. One of the reasons for the presence of small-study effects is publication bias, also referred to as reporting bias.

For more formal testing of small-study effects, see [META] [meta bias](#). To assess the impact of publication bias on the results, see [META] [meta trimfill](#).

Also see [Publication bias](#) of [META] [Intro](#) for information about publication bias.

Funnel plots

The funnel plot (Light and Pillemer 1984) is a scatterplot of the study-specific effect sizes against measures of study precision. This plot is commonly used to explore publication bias. In the absence of publication bias, the shape of the scatterplot should resemble a symmetric (inverted) funnel.

In a funnel plot, the effect sizes, $\hat{\theta}_j$'s, from individual studies are plotted on the x axis, and measures of study precision such as standard errors, $\hat{\sigma}_j$'s, or sample sizes, n_j 's, are plotted on the y axis (Sterne and Harbord 2016). The line corresponding to the estimated overall effect size is also plotted on a funnel plot. In addition to standard errors and sample sizes, other choices for metrics on the y axis include inverse standard errors, $1/\hat{\sigma}_j$'s; variances, $\hat{\sigma}_j^2$'s; inverse variances, $1/\hat{\sigma}_j^2$'s; and inverse sample sizes, $1/n_j$'s. Sterne and Egger (2001) studied various metrics and found that the standard-error metric performed well in many cases.

In the absence of publication bias (and between-study heterogeneity), the studies should be distributed symmetrically about the overall effect size because the sampling error is random. Also, the effect-size estimates from the smaller studies will be more variable than those from the larger studies. Thus, the scatter will be wider at the base of the plot creating, in the absence of bias, a symmetrical funnel shape or, more precisely, a symmetrical inverted funnel shape. When the statistically nonsignificant results of smaller studies are not published (and thus not included in the meta-analysis), an asymmetrical shape of the funnel plot may be observed. In this case, the estimate of the overall effect size will overestimate the true effect size. See [Sterne, Becker, and Egger \(2005\)](#) for details. Also see [Examples of using meta funnelplot](#) for examples of funnel plots.

[Sutton \(2009\)](#) states that when the y -axis metric is one of standard error, variance, or their inverses, a $(1 - \alpha) \times 100\%$ CI can be formed around the overall estimate. This CI can provide more formal interpretation of the plot. But the author suggests that caution be used when interpreting these CIs because they are formed around the estimate of the overall effect size that may be affected by publication bias. This is one of the reasons why the funnel-plot CIs are often referred to as pseudo CIs.

In general, there may be many reasons for an asymmetric funnel plot such as the choice of the plotted effect size ([Sterne et al. 2011](#)), the presence of a moderator correlated with the study effect and study size ([Peters et al. 2008](#)), or simply chance. One of the more common reasons, however, is the presence of substantial between-study heterogeneity ([Sterne, Gavaghan, and Egger 2000](#)).

The between-study heterogeneity, if present, must be addressed before the exploration of publication bias. For instance, if there are study-level covariates that explain the differences between the studies, their influence can distort a funnel plot if they are not accounted for in the main meta-analysis ([Sutton 2009](#)). Suppose that during our subgroup meta-analysis (see option `subgroup()` in `[META] meta summarize`), we identified a categorical variable that explains most of the heterogeneity between the studies. The exploration of the publication bias should then be performed separately for each group. That is, a separate funnel plot should be constructed for each subgroup. In the case of a continuous variable, some authors suggest constructing a funnel plot based on the residuals, $\hat{\theta}_j - \mathbf{x}_j \hat{\beta}$, on the x axis against their standard errors on the y axis, where the residuals are obtained from a meta-regression that uses this continuous variable as the moderator; see `[META] meta regress` and `[META] meta regress postestimation`.

Contour-enhanced funnel plots

[Peters et al. \(2008\)](#) (also see [Palmer et al. \[2016\]](#)) suggest that contour lines of statistical significance (or significance contours) be added to the funnel plot. These “contour-enhanced” funnel plots are useful for determining whether the funnel-plot asymmetry is potentially caused by publication bias or is perhaps due to other reasons. The contour lines that correspond to certain significance levels ($\alpha = 0.01, 0.05, 0.1$, etc.) of tests of zero effect sizes are overlaid on the funnel plot. Publication bias is suspect if there are studies, especially smaller studies, that are missing in the nonsignificant regions. Otherwise, other reasons may explain the presence of the funnel-plot asymmetry.

Using meta funnelplot

`meta funnelplot` produces funnel plots. By default, it plots effect sizes against the standard errors, but you can specify a different metric in the `metric()` option. For historical reasons, the default meta-analysis model is the common-effect model with the inverse-variance estimation method. You can specify one of `random()`, `common()`, or `fixed()` to use a different model or method.

By default, the CIs are displayed, which correspond to the confidence level as declared by `meta set` or `meta esize`. You can specify a different level in the `level()` option. You can also specify the `ciopts()` option to modify the default look of the CI lines.

Instead of the CIs, you can request a contour-enhanced funnel plot by specifying the desired levels of significance (as a percentage) in the `contours()` option. The default significance contours are based on two-sided significance tests of individual effect sizes. You can use `lower` or `upper` within `contours()` to specify that the significance contours be based on the corresponding one-sided tests. You can also specify `lines` within `contours()` to recast the contours to be displayed as lines instead of shaded area plots.

You can use the `by(varlist)` option to produce separate funnel plots for each group defined by `varlist`. This option is useful after a subgroup analysis (see option `subgroup()` in [META] [meta summarize](#)). If a subgroup analysis identified a categorical variable that explains some of the between-study variability, that variable must be specified in the `by()` option when using `meta funnelplot` to explore publication bias.

You can also change the default look of the effect-size line by specifying the `esopts()` option.

In the next section, we describe some of the uses of `meta funnelplot`.

Examples of using meta funnelplot

Recall the NSAIDS data of 37 trials on the effectiveness and safety of topical nonsteroidal anti-inflammatory drugs for acute pain described in [Effectiveness of nonsteroidal anti-inflammatory drugs \(nsaids.dta\)](#) of [META] [meta](#). In this section, we use its declared version and focus on the demonstration of various options of `meta funnelplot` and explanation of its output.

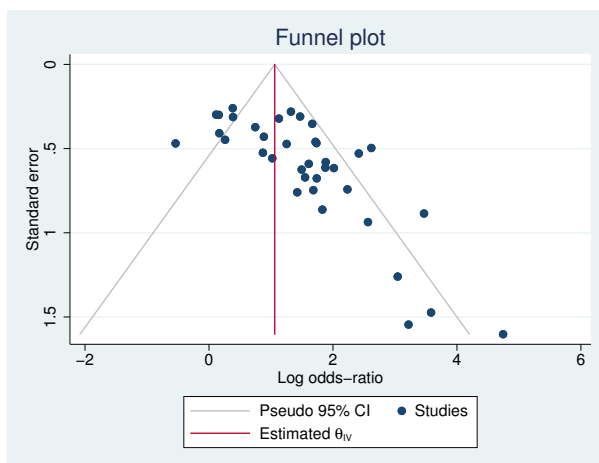
```
. use https://www.stata-press.com/data/r16/nsaidsset
(Effectiveness of nonsteroidal anti-inflammatory drugs; set with -meta esize-)
. meta query, short
-> meta esize nstreat nftreat nscontrol nfcontrol
      Effect-size label:  Log Odds-Ratio
      Effect-size type:  lnoratio
      Effect size:       _meta_es
      Std. Err.:        _meta_se
      Model:             Random-effects
      Method:            REML
```

`meta query, short` reminds us about the main settings of the declaration step. Our data were declared by using `meta esize` with variables `nstreat`, `nftreat`, `nscontrol`, and `nfcontrol` representing the summary data from 2×2 tables, which record the numbers of successes and failures in the treatment and control arms. The computed effect sizes are log odds-ratios; their values and standard errors are stored in the respective [system variables](#) `_meta_es` and `_meta_se`. The declared meta-analysis model is the default random-effects model with the REML estimation method.

Example 1: Funnel plot

To produce a funnel plot for the declared NSAIDS data, we simply type

```
. meta funnelplot
Effect-size label: Log Odds-Ratio
Effect size:      _meta_es
Std. Err.:       _meta_se
Model:           Common-effect
Method:          Inverse-variance
```



The scatterplot of log odds-ratios against their standard errors is produced. The estimated effect-size line and the corresponding pseudo 95% CIs are also plotted. The funnel plot is clearly asymmetric with smaller, less precise studies—studies with larger standard errors—reporting larger effect sizes than the more precise studies. This may suggest the presence of publication bias. The plotted pseudo CI lines are not genuine CI limits, but they provide some insight into the spread of the observed effect sizes about the estimate of the overall effect size. In the absence of publication bias and heterogeneity, we would expect the majority of studies to be randomly scattered within the CI region resembling an inverted funnel shape.

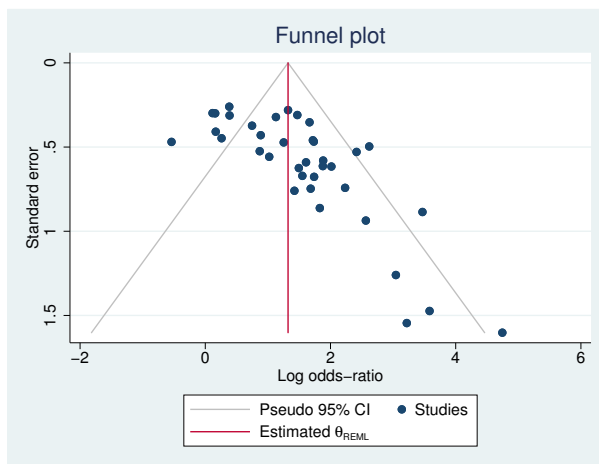
Notice that although the declared meta-analysis model was the random-effects model with the REML estimation method, the default model used by `meta funnelplot` was the common-effect model with the inverse-variance method, as is indicated in the brief output of meta settings reported by the command. This is the model traditionally used with funnel plots in the literature. The reported model and method are used to compute the estimate of the overall effect size, the overall log odds-ratio in our example, which is depicted by the reference (red) effect-size line.

▷ Example 2: Random-effects REML model

In [example 1](#), we pointed out that, for historical reasons, `meta funnelplot` uses the common-effect model with the inverse-variance method by default instead of the declared ones.

If desired, we can obtain the results assuming the declared random-effects model with the REML estimation method by specifying the `random` option.

```
. meta funnelplot, random
Effect-size label: Log Odds-Ratio
Effect size:      _meta_es
Std. Err.:       _meta_se
Model:           Random-effects
Method:          REML
```



From the output of `meta settings`, a random-effects model with the REML method is now used to estimate the overall log odds-ratio. Our conclusion about the existence of potential publication bias remains the same.

For brevity, let's suppress the meta setting information from the output of `meta funnelplot` for the rest of the analysis. We can do this by specifying the `nometashow` option with `meta update` (see [\[META\] meta update](#)).

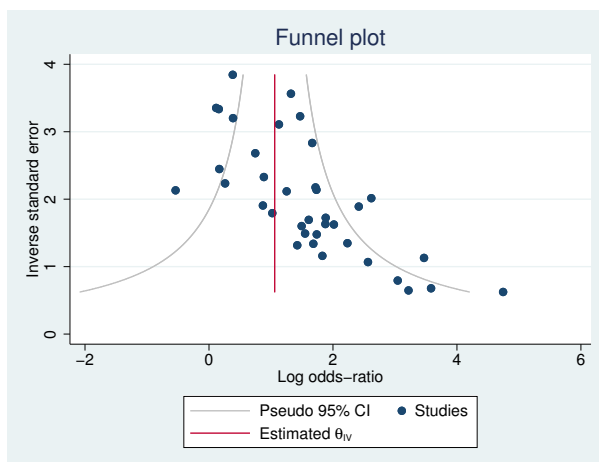
```
. quietly meta update, nometashow
```

Example 3: Inverse-standard-error metric

Continuing with [example 1](#), we can use a different precision metric on the y axis. Using different metrics may reveal different characteristics of the observed studies. For example, for the standard-error metric, more weight is given to the smaller studies, which are more susceptible to publication bias. For the inverse-standard-error metric, more weight is given to the larger studies.

Below, we specify the inverse-standard-error metric by using the `metric(invse)` option.

```
. meta funnelplot, metric(invse)
```



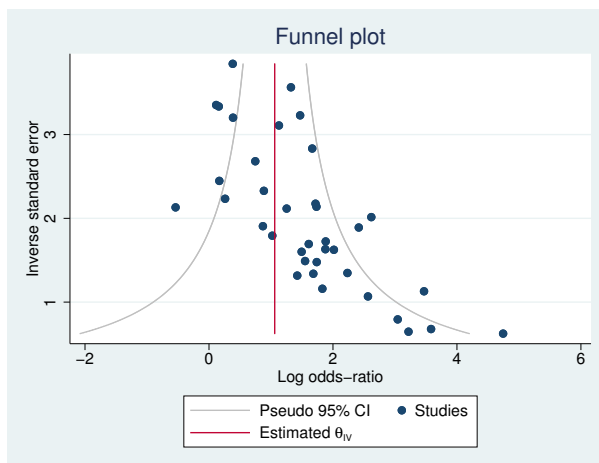
Because our y -axis metric is now $1/\hat{\sigma}_j$, the shape of the plotted CI lines is a hyperbola. The interpretation, however, is similar. We still want to see the majority of studies be concentrated within the regions defined by this hyperbola.

In this metric, the focus is on larger studies with the smaller studies compressed at the bottom. We can see that the asymmetry does not appear to be present for larger studies with, say, $1/\hat{\sigma}_j > 2$. But the asymmetry is present for the smaller studies.

▷ Example 4: The ylabel() option

Continuing with [example 3](#), we can improve the look of the plot by restricting the y -axis values to the observed range. We can do this by specifying the `ylabel(1 2 3)` option:

```
. meta funnelplot, metric(invse) ylabel(1 2 3)
```

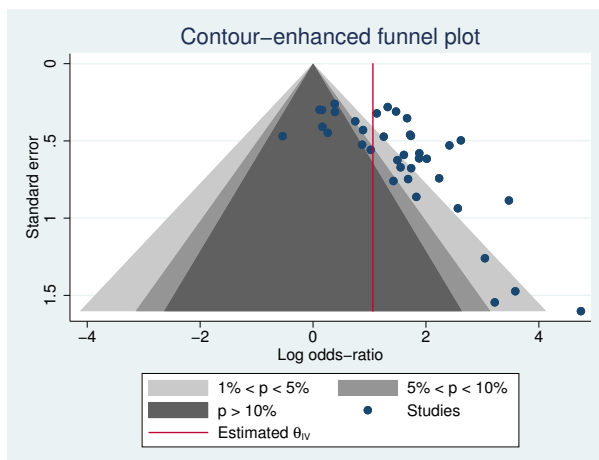


◀

▷ Example 5: Contour-enhanced funnel plot

In [example 11](#) of [\[META\] meta](#), we considered contour-enhanced funnel plots for the NSAIDS data to help us identify whether the asymmetry observed in [example 1](#) is because of publication bias or perhaps some other reasons. Let's obtain the 1%, 5%, and 10% contour-enhanced funnel plots by specifying the `contours(1 5 10)` option:

```
. meta funnelplot, contours(1 5 10)
```



The plotted contour regions define the regions of statistical significance (or nonsignificance) of the individual effect sizes θ_j . That is, if you consider the significance test of $H_0: \theta_j = 0$ for a study j , the contour regions represent the critical regions of such tests for all studies at the specified significance level such as 1%, 5%, and 10% levels in our example. Thus, if a study falls outside a certain region, we have statistical evidence to reject the null hypothesis of no effect at the significance level corresponding to that region.

In our example, for studies in the white region, the null hypothesis of no effect can be rejected at the 1% significance level. That is, the significance tests for these studies would have p -values less than 0.01 or 1%. For studies in the light-gray region, the p -values would be between 1% and 5%. For studies in the darker-gray region, the p -values would be between 5% and 10%. And for studies in the darkest-gray region, the p -values would be larger than 10%.

The plot clearly shows that almost all smaller studies report a statistically significant result, favoring the treatment, either at the 1% or 5% level. On the other hand, some of the larger (more precise) studies (in the darkest-gray region) report nonsignificant results. The hypothetical missing studies—the studies that would make the scatterplot look symmetric with respect to the solid red vertical line—appear to fall in the darkest-gray region corresponding to a p -value of more than 10%. Because we are “missing” small studies in a region of statistical nonsignificance, this suggests that the observed asymmetry in the funnel plot is likely because of publication bias.

Also see [example 13](#) of [\[META\] meta](#).

◀

▷ Example 6: The addplot() option

Continuing with [example 5](#), let’s use the `addplot()` option to add the pseudo 95% CIs around the effect-size line to our contour-enhanced plot. These CIs are computed as follows: $\hat{\theta}_{IV} \pm 1.96 \times y$, where y represents a standard error of $\hat{\theta}_{IV}$, varying within the range of observed standard errors. Here 1.96 corresponds to the $z_{0.975}$ critical value.

We first obtain the estimated value of the overall effect size, which is available in the `r(theta)` scalar from the previous run of `meta funnelplot`. We store it in the `theta` scalar.

```
. display r(theta)
1.0604829
. scalar theta = r(theta)
```

The estimate of the overall effect size may also be obtained from `meta summarize` (see [\[META\] meta summarize](#)):

```
. meta summarize, common(invvariance) nostudies
Meta-analysis summary                Number of studies =    37
Common-effect model
Method: Inverse-variance

      theta: Overall Log Odds-Ratio
```

	Estimate	Std. Err.	z	P> z	[95% Conf. Interval]
theta	1.060483	.0758709	13.98	0.000	.9117788 1.209187

```
. display r(theta)
1.0604829
```

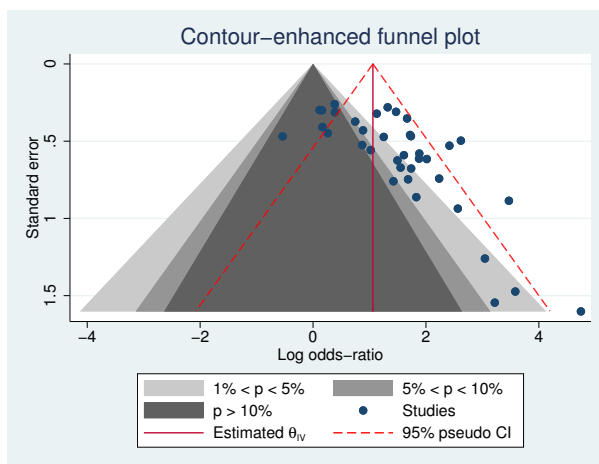
The CI lines can be constructed by using `twoway`'s function command (see [G-2] [graph twoway function](#)):

```
. twoway function theta-1.96*x, horizontal range(0 1.6) ||
>      function theta+1.96*x, horizontal range(0 1.6)
```

In the above, `x` plays the role of `y` in the earlier expression for CIs. We used the `horizontal` option to interchange the roles of `y` and `x` in the function because $\hat{\theta}_j$ appears on the `x` axis and standard errors on the `y` axis in the funnel plot. We also specified the `range()` option so that the range of the plotted function matches the observed range for the standard errors.

We use the above specification in the `addplot()` option with `meta funnelplot`. Because `addplot()` implies a twoway plot, we can omit `twoway` within `addplot()`. We also specify several other options to improve the look of the graph, which we describe later.

```
. local opts horizontal range(0 1.6) lpattern(dash) lcolor("red")
> legend(order(1 2 3 4 5 6) label(6 "95% pseudo CI"))
. meta funnel, contours(1 5 10)
> addplot(function theta-1.96*x, 'opts' || function theta+1.96*x, 'opts')
```



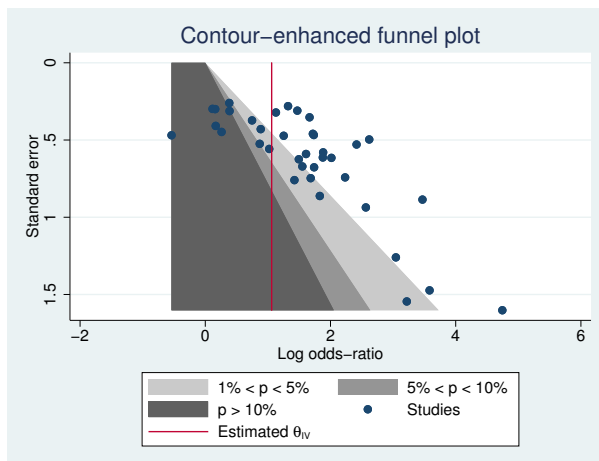
We changed the color and pattern of the CI lines by using options `lpattern()` and `lcolor()`. We used `legend()`'s suboption `order()` to display only the first six keys for the legend to avoid duplicate keys for the two CI plots. We also provided a more descriptive label for the CI legend key. Also, to be more precise, we could have replaced 1.96 in the above with `invnormal(.975)`, which computes the corresponding quantile of the standard normal distribution.

◀

▶ Example 7: Upper one-sided significance contours

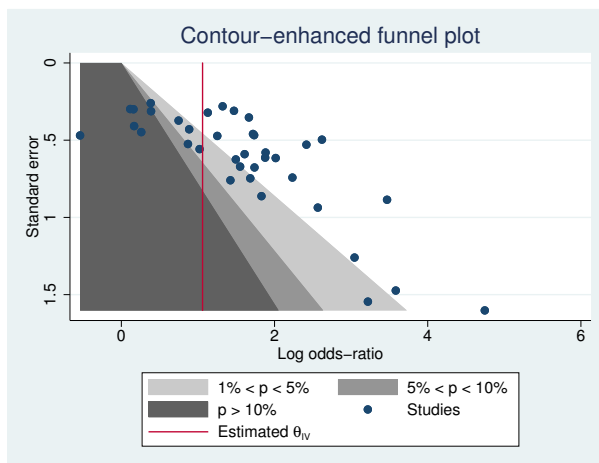
Continuing with [example 5](#), let's produce a contour-enhanced funnel plot based on one-sided significance tests instead of the default two-sided tests. We can specify `lower` or `upper` within `contours()` to produce a funnel plot with contours based on lower or upper one-sided tests. Below, we specify the upper suboption:

```
. meta funnelplot, contours(1 5 10, upper)
```



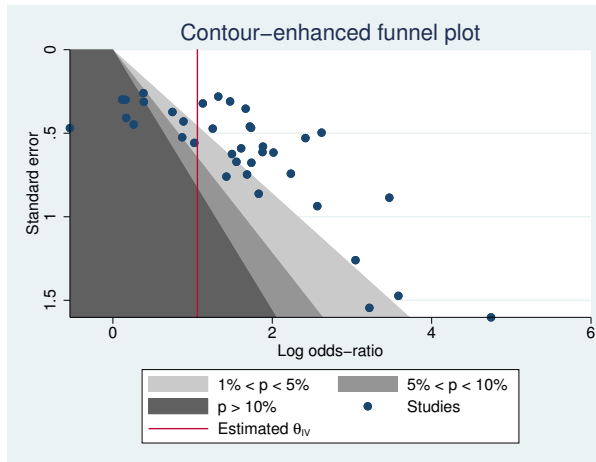
The interpretation of a one-sided contour-enhanced funnel plot is similar to that of a two-sided one. Studies that fall in the upper-tailed region (the white region to the right) are statistically significant at the 1% level based on a one-sided upper-tailed test. The white space on the left is uninformative, and we can suppress it by disallowing the x axis to extend to -2 . This may be done by specifying `xlabel(0(2)6)` (see [G-3] [axis_label_options](#)).

```
. meta funnelplot, contours(1 5 10, upper) xlabel(0(2)6)
```



If we want to suppress all the extra white space around the plot, we can specify `plotregion(margin(zero))` (see [G-3] *region_options*).

```
. meta funnelplot, contours(1 5 10, upper) xlabel(0(2)6)
> plotregion(margin(zero))
```



◀

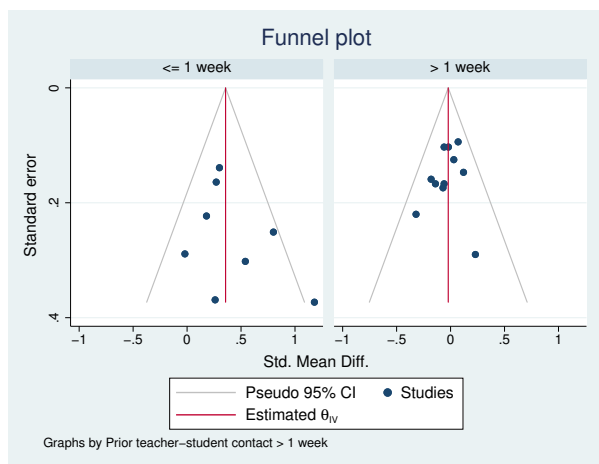
► Example 8: Group-specific funnel plots: option `by()`

In [example 9](#) of [META] [meta summarize](#), we performed subgroup analysis for the pupil IQ data to account for the between-study heterogeneity induced by the binary covariate `week1`, which records whether teachers had prior contact with students for more than 1 week or for 1 week or less. See [Effects of teacher expectancy on pupil IQ \(pupiliq.dta\)](#) of [META] [meta](#) for details about the data.

Let's check for publication bias in these data, accounting for the between-study heterogeneity. We can do this by looking at a funnel plot separately for each category of `week1`. We will use `meta funnel`'s option `by()` to produce such plots.

We use a previously declared version of the dataset.

```
. use https://www.stata-press.com/data/r16/pupiliqset, clear
(Effects of teacher expectancy on pupil IQ; set with -meta set-)
. meta funnelplot, by(week1)
Effect-size label: Std. Mean Diff.
Effect size: stdmdiff
Std. Err.: se
Model: Common-effect
Method: Inverse-variance
```



The above graph shows funnel plots of the subgroups for prior contact of one week or less and more than one week, respectively. These funnels are centered on different effect-size values, but there is little evidence of asymmetry in either plot. We should be careful with our interpretation, however, because we have only a few studies in each plot.

◀

Stored results

`meta funnelplot` stores the following in `r()`:

Scalars

<code>r(theta)</code>	estimated overall effect size
<code>r(xmin)</code>	minimum abscissa of scatter points
<code>r(xmax)</code>	maximum abscissa of scatter points
<code>r(ymin)</code>	minimum ordinate of scatter points
<code>r(ymax)</code>	maximum ordinate of scatter points

Macros

<code>r(model)</code>	meta-analysis model
<code>r(method)</code>	meta-analysis estimation method
<code>r(metric)</code>	metric for the y axis
<code>r(contours)</code>	significance levels of contours

Methods and formulas

`meta funnel` produces a scatterplot of individual effect sizes, $\hat{\theta}_j$'s, on the x axis and measures of study precision on the y axis. The supported measures of precision are the standard errors, $\hat{\sigma}_j$'s (default); inverse standard errors, $1/\hat{\sigma}_j$'s; variances, $\hat{\sigma}_j^2$'s; inverse variances, $1/\hat{\sigma}_j^2$'s; sample sizes, n_j 's; and inverse sample sizes, $1/n_j$'s.

The effect-size reference (solid red) line is plotted at the estimate of the overall effect size. By default, the overall effect size is estimated assuming a common-effect model with the inverse-variance method, but this can be changed by specifying one of `random()`, `common()`, or `fixed()`.

The pseudo $(1 - \alpha) \times 100\%$ CI lines, plotted by default, are constructed as $\hat{\theta} \pm z_{1-\alpha/2}f(y)$, where $\hat{\theta}$ is the estimate of the overall effect size, $z_{1-\alpha/2}$ is the $(1 - \alpha/2)$ th quantile of the standard normal distribution, and $f(y)$ plays the role of the varying standard error of $\hat{\theta}$ as a function of the y -axis values and depends on the chosen metric. When standard errors, $\hat{\sigma}_j$'s, are plotted on the y axis,

$$f(y) = y$$

and the CI curves form straight lines. When variances, $\hat{\sigma}_j^2$'s, are plotted on the y axis,

$$f(y) = \sqrt{y}$$

and the CI curves form a parabola. When inverse standard deviations, $1/\hat{\sigma}_j$'s, or inverse variances, $1/\hat{\sigma}_j^2$, are plotted on the y axis

$$f(y) = \frac{1}{y} \quad \text{and} \quad f(y) = \frac{1}{\sqrt{y}}$$

and the CI curves form a hyperbola.

When the `contours()` option is specified, the contour region corresponding to a significance level α (specified as a percentage in `contours()`) for a two-sided test is defined as the set of points (x, y) ,

$$\left\{ (x, y) : \left| \frac{x}{f(y)} \right| \leq z_{1-\alpha/2} \right\}$$

where $f(y)$ depends on the chosen metric and is defined as before. For upper one-sided tests, the contour region is defined as

$$\left\{ (x, y) : \frac{x}{f(y)} \geq z_{1-\alpha} \right\}$$

and for lower one-sided tests, it is defined as

$$\left\{ (x, y) : \frac{x}{f(y)} \leq z_{\alpha} \right\}$$

The `n(#)` option specifies how many evaluation points are used to construct the CI lines or, when the `contours()` option is specified, the significance contours. By default, 300 points are used for y for CI lines and for each of x and y for contours.

References

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- Sutton, A. J. 2009. Publication bias. In *The Handbook of Research Synthesis and Meta-Analysis*, ed. H. Cooper, L. V. Hedges, and J. C. Valentine, 2nd ed., 435–452. New York: Russell Sage Foundation.

Also see

[META] [meta bias](#) — Tests for small-study effects in meta-analysis

[META] [meta data](#) — Declare meta-analysis data

[META] [meta trimfill](#) — Nonparametric trim-and-fill analysis of publication bias

[META] [meta](#) — Introduction to meta

[META] [Glossary](#)

[META] [Intro](#) — Introduction to meta-analysis

Title

meta bias — Tests for small-study effects in meta-analysis

Description	Quick start	Menu	Syntax
Options	Remarks and examples	Stored results	Methods and formulas
References	Also see		

Description

`meta bias` performs tests for the presence of [small-study effects](#) in a meta-analysis, also known as tests for funnel-plot asymmetry and publication-bias tests. Three regression-based tests and a nonparametric rank correlation test are available. For regression-based tests, you can include moderators to account for potential between-study heterogeneity.

Quick start

Test for small-study effects by using the Egger regression-based test

```
meta bias, egger
```

As above, but include a moderator `x1` to account for between-study heterogeneity induced by `x1`

```
meta bias x1, egger
```

As above, but assume a random-effects model with the empirical Bayes method for estimating τ^2 in the regression-based test

```
meta bias x1, egger random(ebayes)
```

With log risk-ratios, test for small-study effects by using the Harbord regression-based test with moderators `x1` and `x2` to account for between-study heterogeneity

```
meta bias x1 i.x2, harbord
```

With log odds-ratios, test for small-study effects by using the Peters regression-based test and assuming a common-effect model

```
meta bias, peters common
```

Menu

Statistics > Meta-analysis

Syntax

Regression-based tests for small-study effects

Test using meta-analysis model as declared with meta set or meta esize

```
meta bias [moderators] [if] [in], regtest [modelopts]
```

Random-effects meta-analysis model

```
meta bias [moderators] [if] [in], regtest random [(remethod)]
[se(seadj) options]
```

Common-effect meta-analysis model

```
meta bias [if] [in], regtest common [options]
```

Fixed-effects meta-analysis model

```
meta bias [moderators] [if] [in], regtest fixed [multiplicative options]
```

Traditional test

```
meta bias [if] [in], regtest traditional [options]
```

Nonparametric rank correlation test for small-study effects

```
meta bias [if] [in], begg [ [no]metashow detail]
```

<i>regtest</i>	Description
<code>egger</code>	Egger's test
<code>harbord</code>	Harbord's test
<code>peters</code>	Peters's test

modelopts is any option relevant for the declared model.

<i>remethod</i>	Description
<code>reml</code>	restricted maximum likelihood; the default
<code>mle</code>	maximum likelihood
<code>ebayes</code>	empirical Bayes
<code>dlaird</code>	DerSimonian–Laird
<code>sjonkman</code>	Sidik–Jonkman
<code>hedges</code>	Hedges
<code>hschmidt</code>	Hunter–Schmidt

<i>options</i>	Description
Main	
<code>tdistribution</code>	report t test instead of z test
<code>[no]metashow</code>	display or suppress meta settings in the output
<code>detail</code>	display intermediate estimation results
Maximization	
<code>maximize_options</code>	control the maximization process of the between-study variance

moderators may contain factor variables; see [U] 11.4.3 Factor variables.

Options

Main

One of `egger`, `harbord`, `peters`, or `begg` (or their synonyms) must be specified. In addition to the traditional versions of the regression-based tests, their random-effects versions and extensions to allow for moderators are also available.

`egger` (synonym `esphillips`) specifies that the regression-based test of Egger et al. (1997a) be performed. This test is known as the Egger test in the literature. This is the test of the slope in a weighted regression of the effect size, `_meta_es`, on its standard error, `_meta_se`, optionally adjusted for *moderators*. This test tends to have an inflated type I error rate for binary outcomes.

`harbord` (synonym `hesterne`) specifies that the regression-based test of Harbord, Egger, and Sterne (2006) be performed. This test is known as the Harbord test. This is the test of the slope in a weighted regression of Z_j/V_j on $1/\sqrt{V_j}$, optionally adjusting for *moderators*, where Z_j is the score of the likelihood function and V_j is the score variance. This test is used for binary data with effect sizes log odds-ratio and log risk-ratio. It was designed to reduce the correlation between the effect-size estimates and their corresponding standard errors, which is inherent to the Egger test with binary data.

`peters` (synonym `petersetal`) specifies that the regression-based test of Peters et al. (2006) be performed. This test is known as the Peters test in the literature. This is the test of the slope in a weighted regression of the effect size, `_meta_es`, on the inverse sample size, $1/n_j$, optionally adjusted for *moderators*. The Peters test is used with binary data for log odds-ratios. Because it regresses effect sizes on inverse sample sizes, they are independent by construction.

`begg` (synonym `bmazumdar`) specifies that the nonparametric rank correlation test of Begg and Mazumdar (1994) be performed. This is not a regression-based test, so only options `metashow`, `nometashow`, and `detail` are allowed with it. This test is known as the Begg test in the literature. This test is no longer recommended in the literature and provided for completeness.

Options `random()`, `common`, and `fixed`, when specified with `meta bias` for regression-based tests, temporarily override the global model declared by `meta set` or `meta esize` during the computation. Options `random()`, `common`, and `fixed` may not be combined. If these options are omitted, the declared meta-analysis model is assumed; see *Declaring a meta-analysis model* in [META] `meta data`. Also see *Meta-analysis models* in [META] `Intro`.

`random` and `random(remethod)` specify that a random-effects model be assumed for regression-based test; see *Random-effects model* in [META] `Intro`.

remethod specifies the type of estimator for the between-study variance τ^2 . *remethod* is one of `reml`, `mle`, `ebayes`, `dlaird`, `sjonkman`, `hedges`, or `hschmidt`. `random` is a synonym for `random(reml)`. See *Options* in [META] `meta esize` for more information.

`common` specifies that a common-effect model be assumed for regression-based test; see *Common-effect (“fixed-effect”) model* in [META] **Intro**. It uses the inverse-variance estimation method; see *Meta-analysis estimation methods* in [META] **Intro**. Also see the **discussion** in [META] **meta data** about common-effect versus fixed-effects models. `common` is not allowed in the presence of moderators.

`fixed` specifies that a fixed-effects model be assumed for regression-based test; see *Fixed-effects model* in [META] **Intro**. It uses the inverse-variance estimation method; see *Meta-analysis estimation methods* in [META] **Intro**. Also see the **discussion** in [META] **meta data** about fixed-effects versus common-effect models.

`se(seadj)` specifies that the adjustment `seadj` be applied to the standard errors of the coefficients. Additionally, the tests of significance of the coefficients are based on a Student’s *t* distribution instead of the normal distribution. `se()` is allowed only with random-effects models.

`seadj` is `khartung` [, `truncated`]. Adjustment `khartung` specifies that the Knapp–Hartung adjustment (Hartung and Knapp 2001a, 2001b; Knapp and Hartung 2003), also known as the Sidik–Jonkman adjustment (Sidik and Jonkman 2002), be applied to the standard errors of the coefficients. `hknapp` and `sjonkman` are synonyms for `khartung`. `truncated` specifies that the truncated Knapp–Hartung adjustment (Knapp and Hartung 2003), also known as the modified Knapp–Hartung adjustment, be used.

`traditional` specifies that the traditional version of the selected regression-based test be performed. This option is equivalent to specifying options `fixed`, `multiplicative`, and `tdistribution`. It may not be specified with `moderators`.

`multiplicative` performs a fixed-effects regression-based test that accounts for residual heterogeneity by including a multiplicative variance parameter ϕ . ϕ is referred to as an “(over)dispersion parameter”. See *Introduction* in [META] **meta regress** for details.

`tdistribution` reports a *t* test instead of a *z* test. This option may not be combined with option `se()`.

`metashow` and `nometashow` display or suppress the meta setting information. By default, this information is displayed at the top of the output. You can also specify `nometashow` with `meta update` to suppress the meta setting output for the entire meta-analysis session.

`detail` specifies that intermediate estimation results be displayed. For regression-based tests, the results from the regression estimation will be displayed. For the nonparametric test, the results from `ktau` ([R] **spearman**) will be displayed.

Maximization

`maximize_options`: `iterate(#)`, `tolerance(#)`, `nrtolerance(#)`, `nonrtolerance` (see [R] **Maximize**), `from(#)`, and `showtrace`. These options control the iterative estimation of the between-study variance parameter, τ^2 , with random-effects methods `reml`, `mle`, and `ebayes`. These options are seldom used.

`from(#)` specifies the initial value for τ^2 during estimation. By default, the initial value for τ^2 is the noniterative Hedges estimator.

`showtrace` displays the iteration log that contains the estimated parameter τ^2 , its relative difference with the value from the previous iteration, and the scaled gradient.

Remarks and examples

Remarks are presented under the following headings:

Introduction

Using meta bias

Examples of using meta bias

Introduction

As we discussed in *Introduction* of [META] **meta funnelplot**, there is a tendency for smaller studies to report different, often larger, effect sizes than the larger studies. There are various reasons that explain this tendency, but the two more common ones are between-study heterogeneity and publication bias. We covered the between-study heterogeneity in [META] **meta summarize** and [META] **meta regress**. Here we focus on publication bias.

Publication bias often arises when the decision of whether to publish a study depends on the statistical significance of the results of the study. Typically, nonsignificant results from small studies have a tendency of not getting published. See *Publication bias* of [META] **Intro** for details.

The funnel plot ([META] **meta funnelplot**) is commonly used to investigate publication bias or, more generally, **small-study effects** in meta-analysis. The presence of asymmetry in the funnel plot may indicate the presence of publication bias. Graphical evaluation of funnel plots is useful for data exploration but may be subjective when detecting the asymmetry. Thus, a more formal evaluation of funnel-plot asymmetry is desired. Statistical tests were developed for detecting the asymmetry in a funnel plot; they are often called tests for funnel-plot asymmetry. They are also sometimes referred to as tests of publication bias, but this terminology may be misleading because the presence of a funnel-plot asymmetry is not always due to publication bias (for example, Sterne et al. [2011]). Thus, we prefer a more generic term—tests for small-study effects—suggested by Sterne, Gavaghan, and Egger (2000).

There are two types of tests for small-study effects: regression-based tests and a nonparametric rank-based test. The main idea behind these tests is to determine whether there is a statistically significant association between the effect sizes and their measures of precision such as effect-size standard errors.

The Egger regression-based test (Egger et al. 1997b) performs a weighted linear regression of the effect sizes, $\hat{\theta}_j$'s, on their standard errors, $\hat{\sigma}_j$'s, weighted by the precision, $1/\hat{\sigma}_j$'s. The test for the zero slope in that regression provides a formal test for small-study effects. In some cases, such as in the presence of a large true effect or with binary data, the Egger test tends to have an inflated type I error (for example, Harbord, Harris, and Sterne [2016]). Two alternative tests, the Harbord test and the Peters test, were proposed to alleviate the type I error problem in those cases.

The Harbord regression-based test (Harbord, Egger, and Sterne 2006) corresponds to the zero-slope test in a weighted regression of Z_j/V_j 's on $1/\sqrt{V_j}$'s, where Z_j is the score of the likelihood function and V_j is the score variance. The Peters regression-based test (Peters et al. 2006) corresponds to the zero-slope test in a weighted regression of the effect sizes, $\hat{\theta}_j$'s, on the respective inverse sample sizes, $1/n_j$'s. With binary data, these tests tend to perform better than the Egger test in terms of the type I error while maintaining similar power.

The rank correlation Begg test (Begg and Mazumdar 1994) tests whether Kendall's rank correlation between the effect sizes and their variances equals zero. The regression-based tests tend to perform better in terms of type I error than the rank correlation test. This test is provided mainly for completeness.

See Harbord, Harris, and Sterne (2016) and Steichen (2016) for more details about these tests.

As we discussed in [META] [meta funnelplot](#), the presence of between-study heterogeneity may affect the symmetry of a funnel plot. Thus, any statistical method based on the funnel plot will also be affected (Sutton 2009). To account for the between-study heterogeneity, the regression-based tests can be extended to incorporate [moderators](#) that may help explain the heterogeneity (Sterne and Egger 2005).

The traditional version of the regression-based tests used a multiplicative fixed-effects meta-regression to account for [residual heterogeneity](#) (see [Introduction](#) of [META] [meta regress](#)). In addition to adjusting for moderators, a random-effects meta-regression is considered a better alternative to account for residual heterogeneity.

Ioannidis and Trikalinos (2007) provide the following recommendations for when it is appropriate to use small-study tests: a) the number of studies should be greater than 10; b) there should be at least one study with a statistically significant result; c) there should be no significant heterogeneity ($I^2 < 50\%$); and d) the ratio of the maximum to minimum variances across studies should be larger than 4; that is, $\max(\{\hat{\sigma}_j^2\}_{j=1}^K) / \min(\{\hat{\sigma}_j^2\}_{i=1}^K) > 4$. If a) is violated, the tests may have low power. If c) is violated, the asymmetry of the funnel plot may be induced by between-study heterogeneity rather than publication bias. If d) is violated, the funnel plot will look more like a horizontal line than an inverted funnel, and the funnel-asymmetry tests will have an inflated type I error. Also see Sterne et al. (2011) for details.

The results of the tests of small-study effects should be interpreted with caution. In the presence of small-study effects, apart from publication bias, other reasons should also be explored to explain the presence of small-study effects. If small-study effects are not detected by a test, their existence should not be ruled out because the tests tend to have low power.

Also see [META] [meta trimfill](#) for assessing the impact of publication bias on the results.

Using meta bias

`meta bias` performs tests for small-study effects. These tests are also known as the tests for funnel-plot asymmetry and tests for publication bias. You can choose from three regression-based tests: the Egger test for continuous outcomes (option `egger`), the Harbord test for binary outcomes with effect sizes log odds-ratio and log risk-ratio (option `harbord`), and the Peters test for log odds-ratios (option `peters`). You can also perform the Begg nonparametric rank correlation test (option `begg`), but this test is no longer recommended in the meta-analysis literature.

Next, we describe the features that are relevant only to the regression-based tests. These tests are based on meta-regression of effect sizes and their measures of precision.

The default meta-analysis model (and method) are as declared by `meta set` or `meta esize`; see [Declaring a meta-analysis model](#) in [META] [meta data](#). You can change the defaults by specifying one of options `random()`, `common()`, or `fixed()`.

Because the regression-based tests use meta-regression, many of the options of `meta regress` (see [META] [meta regress](#)) apply to `meta bias` as well. For example, you can specify that a multiplicative meta-regression be used by the test with option `multiplicative`. And you can specify to use the t test instead of a z test for inference with option `tdistribution`.

The regression-based tests support the `traditional` option, which specifies that the tests be performed as originally published. This option is a shortcut for `fixed`, `multiplicative`, and `tdistribution`.

To account for between-study heterogeneity when checking for publication bias, you can specify moderators with the regression-based tests.

Examples of using meta bias

Recall the pupil IQ data (Raudenbush and Bryk 1985; Raudenbush 1984) described in *Effects of teacher expectancy on pupil IQ (pupiliq.dta)* of [META] **meta**. Here we will use its declared version and will focus on the demonstration of various options of **meta bias** and explanation of its output.

```
. use https://www.stata-press.com/data/r16/pupiliqset
(Effects of teacher expectancy on pupil IQ; set with -meta set-)
. meta query, short
-> meta set stdmdiff se , studylabel(study1bl) eslabel(Std. Mean Diff.)
Effect-size label: Std. Mean Diff.
Effect-size type: Generic
Effect size: stdmdiff
Std. Err.: se
Model: Random-effects
Method: REML
```

From the meta summary, our data were declared by using **meta set** with variables **stdmdiff** and **se** specifying the effect sizes and their standard errors, respectively. The declared meta-analysis model is the default random-effects model with the REML estimation method.

Examples are presented under the following headings:

- Example 1: Small-study effects due to a confounding moderator*
- Example 2: Traditional tests and detailed output*
- Example 3: Harbord's test for small-study effects*

► Example 1: Small-study effects due to a confounding moderator

Our main focus is on investigating the potential presence of small-study effects by using a regression-based test. Because we are working with continuous data, we will use the Egger test.

```
. meta bias, egger
      Effect-size label: Std. Mean Diff.
      Effect size:   stdmdiff
      Std. Err.:    se

Regression-based Egger test for small-study effects
Random-effects model
Method: REML
H0: beta1 = 0; no small-study effects
      beta1 =      1.83
SE of beta1 =      0.724
      z =      2.53
Prob > |z| =      0.0115
```

From the output header, the regression-based test uses the declared random-effects model with REML estimation to account for residual heterogeneity. The estimated slope, $\hat{\beta}_1$, is 1.83 with a standard error of 0.724, giving a test statistic of $z = 2.53$ and a p -value of 0.0115. This means that there is some evidence of small-study effects.

In [example 9](#) of [\[META\] meta summarize](#), we used subgroup-analysis on binary variable `week1`, which records whether teachers had prior contact with students for more than 1 week or for 1 week or less, to account for between-study heterogeneity. It explained most of the heterogeneity present among the effect sizes, with generally higher effect sizes in the low contact group.

Moderators that can explain a substantial amount of the heterogeneity should be included in the regression-based test as a covariate. By properly accounting for heterogeneity through the inclusion of `week1`, we can test for small-study effects due to reasons other than heterogeneity. We include factor variable `week1` as a moderator as follows:

```
. meta bias i.week1, egger
      Effect-size label: Std. Mean Diff.
      Effect size:   stdmdiff
      Std. Err.:    se

Regression-based Egger test for small-study effects
Random-effects model
Method: REML
Moderators: week1
H0: beta1 = 0; no small-study effects
      beta1 =      0.30
SE of beta1 =      0.729
      z =      0.41
Prob > |z| =      0.6839
```

Now that we have accounted for heterogeneity through moderator `week1`, the Egger test statistic is 0.41 with a p -value of 0.6839. Therefore, we have strong evidence to say that the presence of small-study effects was the result of heterogeneity induced by teacher-student prior contact time. ◀

► Example 2: Traditional tests and detailed output

For illustration, we perform the traditional version of the Egger regression-based test by specifying the `traditional` option. We also use the `detail` option to report the meta-regression results used to construct the Egger test.

```
. meta bias, egger traditional detail
      Effect-size label: Std. Mean Diff.
            Effect size: stdmdiff
            Std. Err.: se
Fixed-effects meta-regression          Number of obs =      19
Error: Multiplicative                  Dispersion phi =     1.69
Method: Inverse-variance              Model F(1,17) =     4.17
                                       Prob > F =      0.0571
```

_meta_es	Coef.	Std. Err.	t	P> t	[95% Conf. Interval]	
_meta_se	1.627717	.7975212	2.04	0.057	-.0549052	3.31034
_cons	-.1797108	.126835	-1.42	0.175	-.4473093	.0878876

```
Test of residual homogeneity: Q_res = chi2(17) = 28.77   Prob > Q_res = 0.0367
```

```
Regression-based Egger test for small-study effects
```

```
Fixed-effects model
```

```
Method: Inverse-variance
```

```
H0: beta1 = 0; no small-study effects
```

```
      beta1 =      1.63
```

```
SE of beta1 =      0.798
```

```
      t =      2.04
```

```
Prob > |t| =      0.0571
```

The traditional version also suggests the presence of small-study effects, but its p -value, 0.0571, is larger than that from [example 1](#).

Running the above command is identical to typing

```
. meta regress _meta_se, fixed multiplicative tdistribution
      Effect-size label: Std. Mean Diff.
            Effect size: stdmdiff
            Std. Err.: se
Fixed-effects meta-regression          Number of obs =      19
Error: Multiplicative                  Dispersion phi =     1.69
Method: Inverse-variance              Model F(1,17) =     4.17
                                       Prob > F =      0.0571
```

_meta_es	Coef.	Std. Err.	t	P> t	[95% Conf. Interval]	
_meta_se	1.627717	.7975212	2.04	0.057	-.0549052	3.31034
_cons	-.1797108	.126835	-1.42	0.175	-.4473093	.0878876

```
Test of residual homogeneity: Q_res = chi2(17) = 28.77   Prob > Q_res = 0.0367
```

The header and coefficient table from `meta bias`'s detailed output is identical to that produced by `meta regress` (see [\[META\] meta regress](#)).

◀

▶ Example 3: Harbord's test for small-study effects

In [example 1](#) of [\[META\] meta funnelplot](#), we explored the presence of publication bias in the NSAIDS data, which was described in *Effectiveness of nonsteroidal anti-inflammatory drugs (nsaids.dta)* of [\[META\] meta](#). The contour-enhanced funnel plot from [example 5](#) of [\[META\] meta funnelplot](#) revealed that the funnel-plot asymmetry was caused by the absence of small studies in the region where the tests of the log odds-ratios equal to zero were not statistically significant. This may suggest the presence of publication bias. We can explore this more formally by performing a test for small-study effects.

We use the declared version of the NSAIDS dataset.

```
. use https://www.stata-press.com/data/r16/nsaidsset, clear
(Effectiveness of nonsteroidal anti-inflammatory drugs; set with -meta esize-)
. meta query, short
-> meta esize nstreat nftreat nscontrol nfcontrol
    Effect-size label:  Log Odds-Ratio
    Effect-size type:  lnoratio
    Effect size:       _meta_es
    Std. Err.:        _meta_se
    Model:             Random-effects
    Method:           REML
```

The declared effect size is log odds-ratio, so we will use the Harbord regression-based test to investigate whether the small-study effects (or funnel-plot asymmetry) is present in these data.

```
. meta bias, harbord
    Effect-size label:  Log Odds-Ratio
    Effect size:       _meta_es
    Std. Err.:        _meta_se

Regression-based Harbord test for small-study effects
Random-effects model
Method: REML

H0: beta1 = 0; no small-study effects
    beta1 =          3.03
SE of beta1 =       0.741
    z =            4.09
    Prob > |z| =     0.0000
```

The p -value is less than 0.0000, so we reject the null hypothesis of no small-study effects. It is difficult to be certain whether the small-study affects are driven by publication bias because of the presence of substantial heterogeneity in these data (see [META] [meta summarize](#)). Note that the regression-based test assumed an (REML) random-effects model, which accounts for heterogeneity present among the studies. If we had access to study-level covariates for these data that could explain some of the between-study variability, we could have specified them with `meta bias`.

◀

Stored results

For regression-based tests, `meta bias` stores the following in `r()`:

Scalars

<code>r(beta1)</code>	estimate of the main slope coefficient
<code>r(se)</code>	standard error for the slope estimate
<code>r(z)</code>	z statistic
<code>r(t)</code>	t statistic
<code>r(p)</code>	two-sided p -value

Macros

<code>r(testtype)</code>	type of test: <code>egger</code> , <code>harbord</code> , or <code>peters</code>
<code>r(model)</code>	meta-analysis model
<code>r(method)</code>	meta-analysis estimation method
<code>r(moderators)</code>	moderators used in regression-based tests

Matrices

<code>r(table)</code>	regression results
-----------------------	--------------------

For Begg's test, `meta bias` stores the following in `r()`:

Scalars	
<code>r(score)</code>	Kendall's score estimate
<code>r(se_score)</code>	standard error of Kendall's score
<code>r(z)</code>	z test statistic
<code>r(p)</code>	two-sided p -value
Macros	
<code>r(testtype)</code>	<code>begg</code>

Methods and formulas

Methods and formulas are presented under the following headings:

Regression-based tests

Egger's linear regression test

Harbord's test for log odds-ratios or log risk-ratios

Peters's test for log odds-ratios

Begg's rank correlation test

Let K be the number of studies for a given meta-analysis. For the j th study, $\hat{\theta}_j$ denotes the estimated effect size, and $\hat{\sigma}_j^2$ denotes the effect-size (within-study) variance. The tests are applicable to any type of effect size as long as it is asymptotically normally distributed.

For binary data, also consider the following 2×2 table for the j th study.

group	event	no event	size
treatment	a_j	b_j	$n_{1j} = a_j + b_j$
control	c_j	d_j	$n_{2j} = c_j + d_j$

The total sample size for the j th study is denoted by $n_j = n_{1j} + n_{2j}$.

Regression-based tests

Regression-based tests use meta-regression to examine a linear relationship between the individual effect sizes and measures of study precision such as the effect-size standard errors, possibly adjusting for moderators that explain some of the between-study variability.

In the subsections below, we provide the traditional versions of the regression-based tests. The extensions of traditional versions include the support of other models such as a random-effects model and the support of moderators.

In the presence of moderators, the test for small-study effects is the test of $H_0: \beta_1 = 0$ in the corresponding meta-regression with the following linear predictor,

$$\mathbf{x}_j\boldsymbol{\beta} = \beta_0 + \beta_1 m_j + \beta_2 x_{2,j} + \cdots + \beta_{p-1} x_{p-1,j}$$

where $x_{2,j}, \dots, x_{p-1,j}$ represent the *moderators* specified with `meta bias` and $m_j = \hat{\sigma}_j$ for the Egger test, $m_j = 1/\sqrt{V_j}$ for the Harbord test, and $m_j = 1/n_j$ for the Peters test. See the subsections below for details about these tests. Also see [Sterne and Egger \(2005\)](#).

The computations of regression-based tests are based on the corresponding meta-regression models; see [Methods and formulas](#) of `[META] meta regress`.

The formulas below are based on [Harbord, Harris, and Sterne \(2016\)](#), [Sterne and Egger \(2005\)](#), and [Peters et al. \(2010\)](#).

Egger's linear regression test

The formulas and discussion in this subsection are based on [Sterne and Egger \(2005\)](#).

The test proposed by [Egger, Davey Smith, Schneider, and Minder \(1997b\)](#) is based on a simple linear regression of the standard normal variate, which is defined as the individual effect-size estimate divided by its standard error, against the study precision, which is defined as the reciprocal of the standard error:

$$E\left(\frac{\hat{\theta}_j}{\hat{\sigma}_j}\right) = b_0 + b_1 \frac{1}{\hat{\sigma}_j} \quad (1)$$

The Egger test of no small-study effects is the test of $H_0: b_0 = 0$.

Linear regression model (1) is equivalent to the weighted linear regression of the effect sizes $\hat{\theta}_j$'s on their standard errors $\hat{\sigma}_j$'s,

$$E\left(\hat{\theta}_j\right) = b_1 + b_0 \hat{\sigma}_j \quad (2)$$

with weights inversely proportional to the variances of the effect sizes, $w_j = 1/\hat{\sigma}_j^2$. Note that the intercept b_0 in regression (1) corresponds to the slope in the weighted regression (2). Therefore, Egger test for small-study effects corresponds to a test of a linear trend in a funnel plot (see [\[META\] meta funnelplot](#)) of effect sizes against their standard errors.

Let's denote $\beta_0 = b_1$ and $\beta_1 = b_0$. The statistical model for the traditional Egger's test, as it originally appeared in the literature ([Egger et al. 1997b](#)), is given by

$$\hat{\theta}_j = \beta_0 + \beta_1 \hat{\sigma}_j + \epsilon_j \quad \text{weighted by } w_j = 1/\hat{\sigma}_j^2, \text{ where } \epsilon_j \sim N(0, \hat{\sigma}_j^2 \phi)$$

and ϕ is the overdispersion parameter as defined in multiplicative meta-regression; see [Introduction](#) of [\[META\] meta regress](#).

Egger's test for small-study effects is the test of $H_0: \beta_1 = 0$, and the null hypothesis is rejected if

$$t_{\text{egger}} = \left| \frac{\hat{\beta}_1}{\widehat{\text{SE}}(\hat{\beta}_1)} \right| > t_{K-2, 1-\alpha/2}$$

where $t_{K-2, 1-\alpha/2}$ is the $(1 - \alpha/2)$ th quantile of the Student's t distribution with $K - 2$ degrees of freedom. The above test is performed when you specify options `egger` and `traditional`.

□ Technical note

[Sterne and Egger \(2005\)](#) point out that, originally, [Egger et al. \(1997b\)](#) used a weighted version of (1) with weights equal to the inverse of the variances of effect sizes ($1/\hat{\sigma}_j^2$'s). The authors strongly recommend that this version of the test not be used because it does not have a theoretical justification. □

Harbord's test for log odds-ratios or log risk-ratios

Consider the fixed-effects model $\hat{\theta}_j \sim N(\theta, \hat{\sigma}_j^2)$. For a study j , let Z_j be the first derivative (score) and V_j be the negative second derivative (Fisher's information) of the model log likelihood with respect to θ evaluated at $\theta = 0$ (Whitehead and Whitehead 1991; Whitehead 1997).

For binary data, Harbord, Egger, and Sterne (2006) proposed a modification of the Egger test based on the intercept in an unweighted regression of $Z_j/\sqrt{V_j}$ against $\sqrt{V_j}$:

$$E\left(\frac{Z_j}{\sqrt{V_j}}\right) = b_0 + b_1\sqrt{V_j} \quad (3)$$

When the effect of interest is the log odds-ratio,

$$Z_j = \frac{a_j - (a_j + c_j)n_{1j}}{n_j} \text{ and } V_j = \frac{n_{1j}n_{2j}(a_j + c_j)(b_j + d_j)}{n_j^2(n_j - 1)}$$

Note that Z_j and V_j are the numerator and denominator of the log Peto's odds-ratio as defined in *Methods and formulas* of [META] **meta esize**.

When the effect of interest is the log risk-ratio,

$$Z_j = \frac{a_j n_j - (a_j + c_j)n_{1j}}{b_j + d_j} \text{ and } V_j = \frac{n_{1j}n_{2j}(a_j + c_j)}{n_j(b_j + d_j)}$$

Whitehead (1997) showed that when θ_j is small and n_j is large, $\hat{\theta}_j \approx Z_j/V_j$ and $\hat{\sigma}_j^2 \approx 1/V_j$. In this case, the Harbord regression model (3) is equivalent to Egger's regression model (1). Thus, Harbord's test becomes equivalent to Egger's test when all studies are large and have small effect sizes (Harbord, Harris, and Sterne 2016).

As with Egger's test, if we use the weighted version of regression model (3) and denote $\beta_0 = b_1$ and $\beta_1 = b_0$ in that model, the statistical model for the Harbord test, as it originally appeared in the literature, is given by

$$\frac{Z_j}{V_j} = \beta_0 + \beta_1 \frac{1}{\sqrt{V_j}} + \epsilon_j \quad \text{weighted by } w_j = V_j, \text{ where } \epsilon_j \sim N\left(0, \frac{\phi}{V_j}\right)$$

where ϕ is the overdispersion parameter as defined in multiplicative meta-regression; see *Introduction* of [META] **meta regress**.

Then, the traditional Harbord test is the test of $H_0: \beta_1 = 0$, and its null hypothesis is rejected if $t_{\text{harbord}} = \left| \hat{\beta}_1 / \text{SE}(\hat{\beta}_1) \right| > t_{K-2, 1-\alpha/2}$. This test can be performed when you specify options **harbord** and **traditional**.

Peters's test for log odds-ratios

Peters et al. (2006) provide a test based on the following model:

$$\hat{\theta}_j = \beta_0 + \beta_1 \frac{1}{n_j} + \epsilon_j \quad \text{weighted by } w_j = (a_j + c_j)(b_j + d_j)/n_j, \text{ where } \epsilon_j \sim N(0, \hat{\sigma}_j^2 \phi)$$

$\hat{\theta}_j = \ln(\widehat{\text{OR}}_j)$, and ϕ is the overdispersion parameter as defined in multiplicative meta-regression; see *Introduction* of [META] **meta regress**.

The traditional Peters test is the test of $H_0: \beta_1 = 0$, and its null hypothesis is rejected if $t_{\text{peters}} = \left| \widehat{\beta}_1 / \text{SE}(\widehat{\beta}_1) \right| > t_{K-2, 1-\alpha/2}$. This test can be performed when you specify options **peters** and **traditional**.

When the test is based on the random-effects model, the weights are given by $w_j = 1/(\hat{\sigma}_j^2 + \hat{\tau}^2)$.

Begg's rank correlation test

Consider the standardized effect sizes

$$\hat{\theta}_j^s = \frac{\hat{\theta}_j - \hat{\theta}_{\text{IV}}}{\sqrt{v_j^s}}$$

where

$$\hat{\theta}_{\text{IV}} = \frac{\sum_{j=1}^K \hat{\theta}_j / \hat{\sigma}_j^2}{\sum_{j=1}^K 1 / \hat{\sigma}_j^2}$$

and

$$v_j^s = \text{Var}(\hat{\theta}_j - \hat{\theta}_{\text{IV}}) = \hat{\sigma}_j^2 - \left(\sum_{j=1}^K \hat{\sigma}_j^{-2} \right)^{-1}$$

The Begg test (Begg and Mazumdar 1994) is Kendall's rank correlation test of independence between $\hat{\theta}_j^s$'s and $\hat{\sigma}_j^2$'s; see *Methods and formulas* of [R] **spearman**.

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Also see

- [META] [meta data](#) — Declare meta-analysis data
- [META] [meta funnelplot](#) — Funnel plots
- [META] [meta regress](#) — Meta-analysis regression
- [META] [meta summarize](#) — Summarize meta-analysis data
- [META] [meta trimfill](#) — Nonparametric trim-and-fill analysis of publication bias
- [META] [meta](#) — Introduction to meta
- [META] [Glossary](#)
- [META] [Intro](#) — Introduction to meta-analysis

Title

meta trimfill — Nonparametric trim-and-fill analysis of publication bias

Description	Quick start	Menu	Syntax
Options	Remarks and examples	Stored results	Methods and formulas
References	Also see		

Description

`meta trimfill` performs the nonparametric “trim-and-fill” method to account for publication bias in meta-analysis. The command estimates the number of studies potentially missing from a meta-analysis because of publication bias, imputes these studies, and computes the overall effect-size estimate using the observed and imputed studies. It can also provide a funnel plot, in which omitted studies are imputed.

Quick start

Perform the trim-and-fill analysis of publication bias under the default setting specified in either `meta set` or `meta esize`

```
meta trimfill
```

As above, and request a funnel plot

```
meta trimfill, funnel
```

Specify that the number of missing studies be estimated using the rightmost-run estimator instead of the default linear estimator

```
meta trimfill, estimator(run)
```

Specify that the estimation of the overall effect size be based on the fixed-effects inverse-variance method during the iteration step and random-effects DerSimonian–Laird method during the pooling step of the trim-and-fill algorithm

```
meta trimfill, itermethod(fixed) poolmethod(dlaird)
```

Specify that studies in the right side of the funnel plot (with large effect sizes) be suppressed and need to be imputed

```
meta trimfill, right
```

Menu

Statistics > Meta-analysis

Syntax

```
meta trimfill [if] [in] [, options]
```

<i>options</i>	Description
Main	
<code>estimator(<i>estimator</i>)</code>	estimator for the number of missing studies; default is <code>linear</code>
<code>left</code>	impute studies on the left side of the funnel plot
<code>right</code>	impute studies on the right side of the funnel plot
<code>funnel[(<i>funnelopts</i>)]</code>	draw funnel plot
Options	
<code>level(#)</code>	set confidence level; default is as declared for meta-analysis
<code>eform_option</code>	report exponentiated results
<code>[no]metashow</code>	display or suppress meta settings in the output
<code>display_options</code>	control column formats
Iteration	
<code>random[(<i>remethod</i>)]</code>	random-effects meta-analysis to use for iteration and pooling steps
<code>common</code>	common-effect meta-analysis to use for iteration and pooling steps; implies inverse-variance method
<code>fixed</code>	fixed-effects meta-analysis to use for iteration and pooling steps; implies inverse-variance method
<code>itermethod(<i>method</i>)</code>	meta-analysis to use for iteration step
<code>poolmethod(<i>method</i>)</code>	meta-analysis to use for pooling step
<code>iterate(#)</code>	maximum number of iterations for the trim-and-fill algorithm; default is <code>iterate(100)</code>
<code>[no]log</code>	display an iteration log from the trim-and-fill algorithm

<i>estimator</i>	Description
<code>linear</code>	linear estimator, L_0 ; the default
<code>run</code>	run estimator, R_0
<code>quadratic</code>	quadratic estimator, Q_0 (rarely used)

<i>remethod</i>	Description
<code>reml</code>	restricted maximum likelihood; the default
<code>mle</code>	maximum likelihood
<code>ebayes</code>	empirical Bayes
<code>dlaird</code>	DerSimonian–Laird
<code>sjonkman</code>	Sidik–Jonkman
<code>hedges</code>	Hedges
<code>hschmidt</code>	Hunter–Schmidt

Options

Main

`estimator`(*estimator*) specifies the type of estimator for the number of missing studies. *estimator* is one of `linear`, `run`, or `quadratic`. The default is `estimator(linear)`.

`linear` specifies that the “linear” estimator, L_0 , be used to estimate the number of missing studies. This is the default estimator.

`run` specifies that the rightmost-run estimator, R_0 , be used to estimate the number of missing studies.

`quadratic` specifies that the “quadratic” estimator, Q_0 , be used to estimate the number of missing studies. This estimator is not recommended in the literature and provided for completeness.

Duval and Tweedie (2000a) found that the L_0 and R_0 estimators perform better in terms of mean squared error (MSE) than the Q_0 estimator, with L_0 having the smallest MSE in certain cases. They also found that R_0 tends to be conservative in some cases. Therefore, L_0 is chosen to be the default, but the authors recommend that all estimators be considered in practice. Also see *Estimating the number of missing studies in Methods and Formulas* for details about the estimators.

`left` and `right` specify the side of the funnel plot, where the missing studies are to be imputed. By default, the side is chosen based on the results of the traditional Egger test—if the estimated slope is positive, `left` is assumed; otherwise, `right` is assumed. Only one of `left` or `right` is allowed.

`left` assumes that the leftmost (smallest) effect sizes have been suppressed and specifies to impute them.

`right` assumes that the rightmost (largest) effect sizes have been suppressed and specifies to impute them.

`funnel` and `funnel`(*funnelopts*) specify to draw a funnel plot that includes the imputed studies.

funnelopts are any options as documented in [META] [meta funnelplot](#), except `random[()]`, `common[()]`, `fixed[()]`, `by()`, `[no]metashow`, and `addplot()`.

Options

`level(#)` specifies the confidence level, as a percentage, for confidence intervals. The default is as declared for the meta-analysis session; see *Declaring a confidence level for meta-analysis* in [META] [meta data](#). Also see option `level()` in [META] [meta set](#).

eform_option is one of `eform`, `eform(string)`, `or`, or `rr`. It reports exponentiated effect sizes and transforms their respective confidence intervals, whenever applicable. By default, the results are displayed in the metric declared with `meta set` or `meta esize` such as log odds-ratios and log risk-ratios. *eform_option* affects how results are displayed, not how they are estimated and stored.

`eform(string)` labels the exponentiated effect sizes as *string*; the other options use default labels. The default label is specific to the chosen effect size. For example, option `eform` uses `Odds Ratio` when used with log odds-ratios declared with `meta esize` or `Risk Ratio` when used with the declared log risk-ratios. Option `or` is a synonym for `eform` when log odds-ratio is declared, and option `rr` is a synonym for `eform` when log risk-ratio is declared. If option `eslabel(eslab)` is specified during declaration, then `eform` will use the `exp(eslab)` label or, if `eslab` is too long, the `exp(ES)` label.

`metashow` and `nometashow` display or suppress the meta setting information. By default, this information is displayed at the top of the output. You can also specify `nometashow` with `meta update` to suppress the meta setting output for the entire meta-analysis session.

`display_options`: `cformat(%fmt)`; see [R] [Estimation options](#).

Iteration

Options `random()`, `common`, and `fixed`, when specified with `meta trimfill`, temporarily override the global model declared by `meta set` or `meta esize` during the computation. These options specify that the same method be used during both iteration and pooling steps. To specify different methods, use options `itermethod()` and `poolmethod()`. Options `random()`, `common`, and `fixed` may not be combined. If these options are omitted, the declared meta-analysis model is assumed; see [Declaring a meta-analysis model](#) in [META] [meta data](#). Also see [Meta-analysis models](#) in [META] [Intro](#).

`random` and `random(remethod)` specify that a random-effects model be assumed for meta-analysis during iteration and pooling steps of the trim-and-fill algorithm; see [Random-effects model](#) in [META] [Intro](#).

remethod specifies the type of estimator for the between-study variance τ^2 . *remethod* is one of `reml`, `mle`, `ebayes`, `dlaird`, `sjonkman`, `hedges`, or `hschmidt`. `random` is a synonym for `random(reml)`. See [Options](#) in [META] [meta esize](#) for more information.

`common` specifies that a common-effect model be assumed for meta-analysis during iteration and pooling steps of the trim-and-fill algorithm; see [Common-effect \(“fixed-effect”\) model](#) in [META] [Intro](#). It uses the inverse-variance estimation method; see [Meta-analysis estimation methods](#) in [META] [Intro](#). Also see the [discussion](#) in [META] [meta data](#) about common-effect versus fixed-effects models.

`fixed` specifies that a fixed-effects model be assumed for meta-analysis during iteration and pooling steps of the trim-and-fill algorithm; see [Fixed-effects model](#) in [META] [Intro](#). It uses the inverse-variance estimation method; see [Meta-analysis estimation methods](#) in [META] [Intro](#). Also see the [discussion](#) in [META] [meta data](#) about fixed-effects versus common-effect models.

`itermethod(method)` specifies the meta-analysis method to use during the iteration step of the trim-and-fill algorithm. The default is the method declared for meta-analysis; see [Declaring a meta-analysis model](#) in [META] [meta data](#). Also see [Trim-and-fill algorithm](#) in [Methods and formulas](#). This option may not be combined with `random()`, `common`, or `fixed`.

method is one of the random-effects meta-analysis methods, *remethod*; or a common-effect inverse-variance method, `common`; or a fixed-effects inverse-variance method, `fixed`; see [Options](#) in [META] [meta set](#) for details.

`poolmethod(method)` specifies the meta-analysis method to use during the pooling step of the trim-and-fill algorithm. The default is to use the method declared for meta-analysis; see [Declaring a meta-analysis model](#) in [META] [meta data](#). Also see [Trim-and-fill algorithm](#) in [Methods and formulas](#). This option may not be combined with `random()`, `common`, or `fixed`.

method is one of the random-effects meta-analysis methods, *remethod*; or a common-effect inverse-variance method, `common`; or a fixed-effects inverse-variance method, `fixed`; see [Options](#) in [META] [meta set](#) for details.

`iterate(#)` specifies the maximum number of iterations for the trim-and-fill algorithm. The default is `iterate(100)`. When the number of iterations equals `iterate()`, the algorithm stops and presents the current results. If convergence is not reached, a warning message is also displayed. If convergence is declared before this threshold is reached, the algorithm will stop when convergence is declared.

`noLog` and `log` specify whether an iteration log showing the progress of the trim-and-fill algorithm is to be displayed. By default, the log is suppressed but you can specify `log` to display it.

Remarks and examples

Remarks are presented under the following headings:

Introduction

Using meta trimfill

Examples of using meta trimfill

Introduction

Publication bias is a serious problem in meta-analysis. It arises when the decision whether to publish a study depends on the statistical significance of the results of the study. Typically, more significant findings are more likely to get published. See *Publication bias* of [META] [Intro](#) for details.

Publication bias can be assessed visually with a funnel plot ([META] [meta funnelplot](#)). More formal tests for the presence of publication bias or, more generally, of small-study effects are also available ([META] [meta bias](#)). The focus of this entry is on assessing the impact of the publication bias on the results. One of the methods popular in practice is the so-called trim-and-fill method, introduced by [Duval and Tweedie \(2000a, 2000b\)](#).

The main goal of the trim-and-fill method is to evaluate the impact of publication bias on our final inference. The idea of the method is to iteratively estimate the number of studies potentially missing because of publication bias at the iteration stage. Then, at the final pooling stage, impute (fill in) the effect sizes and effect-size standard errors for these studies and use the completed set of studies to compute the overall effect-size estimate. For details, see *Trim-and-fill algorithm*.

Meta-analysis literature does not provide definitive recommendations for which model should be used during the iteration stage of the method. [Duval \(2005\)](#) points out that a random-effects model was recommended initially ([National Research Council 1992](#); [Sutton et al. 1998](#)), but a common-effect model was found later to provide a more conservative approach. What is meant by conservative? Random-effects models tend to give more weight to less precise studies than common-effect models ([Poole and Greenland 1999](#)). But less precise (smaller) studies are more likely to exhibit publication bias. In general, neither model outperforms the other in all situations. Thus, meta-analysts are advised to try both in practice and compare the results.

Just like other methods for detecting publication bias such as funnel plots and tests for the funnel-plot asymmetry, the trim-and-fill method is sensitive to the presence of substantial between-study heterogeneity (for example, [Peters et al. \[2007\]](#)). The method is agnostic to the reasons for the funnel-plot asymmetry, be it publication bias or between-study heterogeneity. It merely detects the asymmetry and attempts to correct it. Unlike the tests for the funnel-plot asymmetry ([META] [meta bias](#)), the trim-and-fill method does not allow accounting for the moderators that could potentially explain the heterogeneity. For categorical moderators, however, you can perform the method separately for each category. In any case, you should take potential heterogeneity into account when interpreting the final estimate of the effect size from the trim-and-fill method. See the *Trim and fill* section in [Deeks, Higgins, and Altman \(2017\)](#) for details.

[Duval \(2005\)](#) states that there are researchers who are not comfortable with using imputed (fictional) studies to adjust the meta-analysis results for publication bias ([Begg 1997](#)). Indeed, one of the shortcomings of this method is that it treats “imputed” effect sizes as if they were observed in the final meta-analysis and thus potentially underestimates their corresponding standard errors.

Note that the emphasis of the trim-and-fill method is on sensitivity analyses and not on trying to recover the missing study values. The actual imputed values are not of interest. The primary goal of the method is to explore the impact of missing studies on the overall effect-size estimate. This method should be used purely as a guide to which meta-analysis appears to be more susceptible to publication bias.

Using meta trimfill

`meta trimfill` implements the trim-and-fill method of [Duval and Tweedie \(2000a, 2000b\)](#). It supports three estimators: `linear`, `run`, and `quadratic`, which can be specified in the `estimator()` option.

It can impute studies on the left side of the funnel plot with option `left` or on the right with option `right`. By default, it assumes the side where the missingness occurs based on the traditional Egger test; see [\[META\] meta bias](#). Specifically, if the slope from the Egger test is positive, then `left` is assumed; otherwise, `right` is assumed.

You can use option `funnel` or `funnel()` to draw a funnel plot, which would include the imputed studies. You can customize the default look of the funnel plot by specifying `funnel()`.

You can use `eform_option` to exponentiate the results reported in the log metric such as log odds-ratios and log risk-ratios.

By default, the model declared with either `meta set` or `meta esize` is assumed during both the iteration and pooling stages. You can change this by specifying one of `random()`, `common`, or `fixed`. You can also specify a different model for the iteration stage in the `itermethod()` option and for the pooling stage in the `poolmethod()` option.

Examples of using meta trimfill

Here we use `metatrim.dta` to demonstrate the usages of the trim-and-fill method. (This dataset is based on `pupiliq.dta`; it contains a subset of variables renamed to have generic names and fewer observations.) Also see [example 13](#) of [\[META\] meta](#) for the trim-and-fill analysis of the [NSAIDS dataset](#).

► Example 1: A basic trim-and-fill analysis

Consider the dataset `metatrim.dta`. In this dataset, each study consists of two groups, and the standardized difference in means between the two groups is recorded, as well as the standard error for this difference. We begin by declaring the effect sizes and their standard errors with `meta set`.

```

. use https://www.stata-press.com/data/r16/metatrim
(Subset of pupiliq.dta)
. meta set stdmdiff se
Meta-analysis setting information
Study information
  No. of studies: 16
  Study label: Generic
  Study size: N/A
  Effect size
    Type: Generic
    Label: Effect Size
    Variable: stdmdiff
  Precision
  Std. Err.: se
    CI: [_meta_cil, _meta_ciu]
  CI level: 95%
Model and method
  Model: Random-effects
  Method: REML

```

We perform a trim-and-fill analysis to see whether any studies are estimated to be missing:

```

. meta trimfill
  Effect-size label: Effect Size
  Effect size: stdmdiff
  Std. Err.: se
Nonparametric trim-and-fill analysis of publication bias
Linear estimator, imputing on the left
Iteration                                Number of studies = 19
  Model: Random-effects                   observed = 16
  Method: REML                            imputed = 3
Pooling
  Model: Random-effects
  Method: REML

```

Studies	Effect Size	[95% Conf. Interval]	
Observed	0.119	-0.018	0.256
Observed + Imputed	0.034	-0.150	0.219

The model in the iteration and pooling steps is a random-effects model with the REML estimation as declared by `meta set`. These models may be individually controlled by using options `itermethod()` and `poolmethod()`. By default, estimation of the number of missing studies K_0 was based on the linear estimator. This can be changed using the `estimator()` option.

The mean effect size based on the 16 observed studies is 0.119 with a 95% CI of $[-0.018, 0.256]$. Three hypothetical studies, $\widehat{K}_0 = 19 - 16 = 3$, are estimated to be missing and are imputed. If these three studies were included in the meta-analysis, the funnel plot would be more symmetrical. After imputing the studies, we obtain an updated estimate (based on the 19 studies, observed plus imputed) of the mean effect size of 0.034 with a 95% CI $[-0.150, 0.219]$.

► Example 2: Funnel plot

Continuing with [example 1](#), we can request a funnel plot by specifying the `funnel` option.

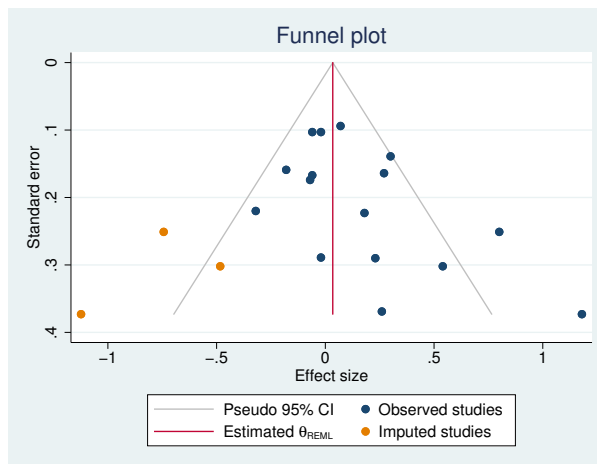
```
. meta trimfill, funnel
      Effect-size label: Effect Size
      Effect size: stdmdiff
      Std. Err.: se

Nonparametric trim-and-fill analysis of publication bias
Linear estimator, imputing on the left

Iteration                                Number of studies =    19
  Model: Random-effects                    observed =          16
  Method: REML                             imputed =           3

Pooling
  Model: Random-effects
  Method: REML
```

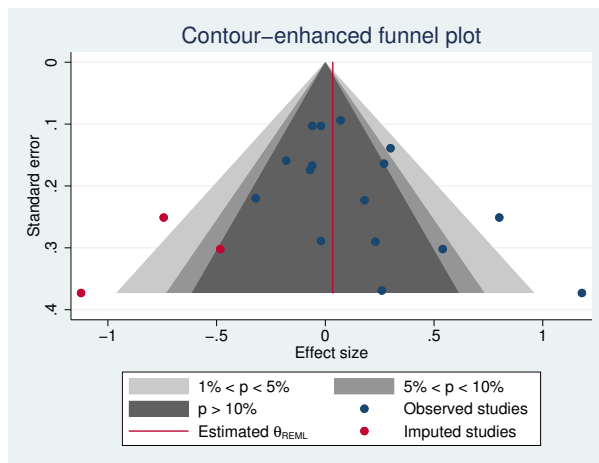
Studies	Effect Size	[95% Conf. Interval]	
Observed	0.119	-0.018	0.256
Observed + Imputed	0.034	-0.150	0.219



The imputed studies are shown in orange in the above funnel plot. Only one of the three imputed studies lies within the 95% pseudo CI; see [\[META\] meta funnelplot](#) for the interpretation of the funnel plot.

We may be interested in a contour-enhanced funnel plot of the completed set of studies (observed and imputed) to visually assess whether the imputed studies fall in regions of statistical significance. This is done by specifying the `contour(1 5 10)` suboption within `funnel()` in `meta trimfill`. We suppress the output from the command with `quietly`.

```
. quietly meta trimfill, funnel(contour(1 5 10))
```



The plot reveals that two of the three imputed studies fall in the white region corresponding to a p -value less than 1%; see [example 5](#) of [\[META\] meta funnelplot](#) for more examples of contour-enhanced funnel plots.

◀

► Example 3: Specifying different pooling and iteration methods

By default, the same meta-analysis methods will be used for both the pooling and iteration steps, but we can specify other methods with the `poolmethod()` and `itermethod()` options. For example, below we specify the random-effects DerSimonian–Laird method for the pooling step and the fixed-effects inverse-variance method for the iteration step:

```
. meta trimfill, itermethod(fixed) poolmethod(dlaird)
Effect-size label: Effect Size
Effect size: stdmdiff
Std. Err.: se

Nonparametric trim-and-fill analysis of publication bias
Linear estimator, imputing on the left

Iteration                                Number of studies =    19
Model: Fixed-effects                       observed =    16
Method: Inverse-variance                   imputed =     3

Pooling
Model: Random-effects
Method: DerSimonian-Laird
```

Studies	Effect Size	[95% Conf. Interval]	
Observed	0.117	-0.016	0.249
Observed + Imputed	0.033	-0.120	0.186

The estimates are only slightly smaller than what they were in [example 1](#), where we used the random-effects REML estimation method in both steps.

◀

▷ Example 4: Specifying the estimator

By default, the linear estimator is used to estimate the number of missing studies. Let's use the rightmost-run estimator and see how the results differ:

```
. meta trimfill, estimator(run)
      Effect-size label: Effect Size
      Effect size: stdmdiff
      Std. Err.: se

Nonparametric trim-and-fill analysis of publication bias
Run estimator, imputing on the left

Iteration                                Number of studies =    18
  Model: Random-effects                    observed =         16
  Method: REML                              imputed =          2

Pooling
  Model: Random-effects
  Method: REML
```

	Studies	Effect Size	[95% Conf. Interval]	
	Observed	0.119	-0.018	0.256
	Observed + Imputed	0.059	-0.124	0.242

The mean effect size based on the 16 observed studies is still 0.119, as it was in [example 1](#), but the updated estimate that includes the imputed studies is large now, 0.059. Also, the estimated number of missing studies is 2 in this example instead of 3.

◀

Stored results

`meta trimfill` stores the following in `r()`:

Scalars

`r(K_total)` total number of studies (observed plus imputed)
`r(K_observed)` number of observed studies
`r(K_imputed)` number of imputed studies
`r(converged)` 1 if trim-and-fill algorithm converged, 0 otherwise

Macros

`r(estimator)` type of estimator for the number of missing studies
`r(side)` side of the funnel plot with missing studies; `left` or `right`
`r(itermethod)` meta-analysis estimation method used during iteration step
`r(poolmethod)` meta-analysis estimation method used during final pooling step
`r(level)` confidence level for CIs

Matrices

`r(table)` trim-and-fill table of results
`r(imputed)` matrix of effect sizes and their standard errors for imputed studies

Methods and formulas

For each study j , $j = 1, \dots, K$, let $\hat{\theta}_j$ be the effect size and $\hat{\sigma}_j^2$ be its squared standard error (or within-study variance). The goal is to estimate an overall effect size, θ , from the sample of the effect sizes, accounting for (potentially) suppressed studies with extreme values from the meta-analysis.

The formulas and discussion below are based on Duval and Tweedie (2000b), Duval (2005), and Steichen (2000).

Suppose that there are K observed studies and K_0 relevant studies that are potentially missing from the meta-analysis because of publication bias. The goal is to estimate the value of K_0 as well as the effect size from the “completed” set of $K + K_0$ studies. The formulas below are based on the assumption that the K_0 effect-size values that are missing are the most extreme smallest values. That is, the studies with “nonsignificant” results are the ones being suppressed. This is also equivalent to assuming that the studies are missing from the left side of the funnel plot (option `left`). If missing studies are expected to be missing on the right side of the funnel (option `right`), Duval (2005) indicates that the same formulas below can be applied after multiplying the effect sizes by -1 .

Note that the default behavior of `meta trimfill` is to assume the side where the missingness occurs based on the traditional Egger test; see *Egger’s linear regression test* in [META] **meta bias**. Specifically, if the slope from the Egger test is positive, then option `left` is assumed, and vice versa.

Methods and formulas are presented under the following headings:

Estimating the number of missing studies
Trim-and-fill algorithm

Estimating the number of missing studies

We follow the description and notation from Duval and Tweedie (2000b) and Steichen (2000). Let $X_j = \hat{\theta}_j - \hat{\theta}$ and denote the ranks of $|X_j|$ ’s as $r_j^* \in \{1, 2, \dots, K\}$. Let $\gamma^* \geq 0$ be the length of the rightmost run of ranks associated with positive values of X_j ’s such that $\gamma^* = K - r_h^*$, where r_h^* is the absolute rank of the most negative X_j with the corresponding index h ; see Rothstein et al. (2005, 137) for an example illustrating the computation of γ^* manually.

The “trimmed” rank test statistic for the observed K values is

$$T_K = \sum_{X_j > 0} r_j^*$$

The following estimators of K_0 can be considered based on the above quantities:

$$\begin{aligned} R_0 &= \gamma^* - 1 \\ L_0 &= \frac{4T_K - K(K + 1)}{2K - 1} \\ Q_0 &= K - \frac{1}{2} - \sqrt{2K^2 - 4T_K + \frac{1}{4}} \end{aligned}$$

Because K_0 must be an integer, the above estimators are rounded as follows,

$$\begin{aligned} R_0^+ &= \max(0, R_0) \\ L_0^+ &= \max\{0, \text{round}(L_0)\} \\ Q_0^+ &= \max\{0, \text{round}(Q_0)\} \end{aligned}$$

where $\text{round}(x)$ is x rounded to the nearest integer.

Duval and Tweedie (2000a) compared the behavior of the MSEs of the estimators R_0^+ , L_0^+ , and Q_0^+ using simulated data and found that L_0^+ and R_0^+ seem to perform better than Q_0^+ . They also found that R_0 tends to be conservative in some cases. In general, the authors recommend that all estimators be considered in practice. Also see Duval and Tweedie (2000b), Duval (2005), and Steichen (2016) for more details about these estimators.

Trim-and-fill algorithm

Without loss of generality, we assume that the observations, $\hat{\theta}_j$'s, are sorted in ascending order.

The steps of the trim-and-fill method below are based on Duval and Tweedie (2000b) and Steichen (2016).

Step 1. At the initial iteration $l = 1$, compute $\hat{\theta}^{(1)}$ using the meta-analysis model as declared with `meta set` or `meta esize`; the model may be changed using the `itermethod()` option.

Compute the centered values

$$\hat{\theta}_j^{(1)} = \hat{\theta}_j - \hat{\theta}^{(1)} \quad j = 1, \dots, K$$

and estimate $\hat{K}_0^{(1)}$ using the default L_0^+ estimator applied to the set of centered values $\hat{\theta}_j^{(1)}$; that is, set $X_j = \hat{\theta}_j^{(1)}$ in the previous section. Other estimators for $\hat{K}_0^{(1)}$ may be specified using the `estimator()` option.

Step 2. At the iteration $l \geq 2$, remove $\hat{K}_0^{(l-1)}$ values from the right end of the set of values $\hat{\theta}_j$, and estimate $\hat{\theta}^{(l)}$ based on the trimmed “symmetric” set of $K - \hat{K}_0^{(l-1)}$ values: $\{\hat{\theta}_1, \dots, \hat{\theta}_{K-\hat{K}_0^{(l-1)}}\}$. Compute the next set of centered values

$$\hat{\theta}_j^{(l)} = \hat{\theta}_j - \hat{\theta}^{(l)} \quad j = 1, \dots, K$$

Step 3. Repeat step 2 until an iteration $l = L$, at which $\hat{K}_0^{(L)} = \hat{K}_0^{(L-1)}$ (and thus, $\hat{\theta}^{(L)} = \hat{\theta}^{(L-1)}$). Set $\hat{K}_0 = \hat{K}_0^{(L)}$.

Step 4. Finally, compute the \hat{K}_0 imputed symmetric values

$$\hat{\theta}_j^* = 2\hat{\theta}^{(L)} - \hat{\theta}_{K-j+1} \quad j = 1, \dots, \hat{K}_0$$

and the corresponding imputed within-study standard errors,

$$\hat{\sigma}_j^* = \hat{\sigma}_{K-j+1} \quad j = 1, \dots, \hat{K}_0$$

Compute the final overall effect-size estimate using the default meta-analysis method or `poolmethod()`, if specified, based on the “completed” dataset $\{\hat{\theta}_1, \dots, \hat{\theta}_K, \hat{\theta}_1^*, \dots, \hat{\theta}_{\hat{K}_0}^*\}$.

References

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Also see

- [META] [meta bias](#) — Tests for small-study effects in meta-analysis
- [META] [meta data](#) — Declare meta-analysis data
- [META] [meta funnelplot](#) — Funnel plots
- [META] [meta summarize](#) — Summarize meta-analysis data
- [META] [meta](#) — Introduction to meta
- [META] [Glossary](#)
- [META] [Intro](#) — Introduction to meta-analysis

Glossary

Begg test, Begg and Mazumdar test. A nonparametric rank correlation test for funnel-plot asymmetry of [Begg and Mazumdar \(1994\)](#). It tests whether Kendall's rank correlation between the effect sizes and their variances equals zero. The regression-based tests such as the [tend to perform better](#) in terms of type I error than the rank correlation test. This test is no longer recommended in the literature and provided mainly for completeness. See [\[META\] meta bias](#).

between-study sample size. The number of studies in a meta-analysis.

between-study variability. Also known as between-study heterogeneity; see [heterogeneity](#).

bubble plot. A scatterplot of effect size against a continuous covariate (moderator) in the meta-regression. The size of points representing the studies is proportional to study weights from a fixed-effects or, optionally, random-effects meta-analysis.

clinical heterogeneity. According to [Deeks, Higgins, and Altman \(2017\)](#), it is “variability in the participants, interventions and outcomes studied”. Clinical variation will lead to [heterogeneity](#) if the effect size is affected by any of these varying factors.

Cochrane's *Q* statistic. See [Q statistic](#).

Cohen's *d*. An effect-size measure introduced by [Cohen \(1988\)](#) for continuous outcomes. It is a standardized mean difference where the difference between the two group means is usually divided by the standard deviation pooled across both groups. See [Standardized mean difference](#) of [Methods and formulas](#) in [\[META\] meta esize](#).

combined effect size. See [overall effect size](#).

common-effect meta-analysis model. A meta-analysis model that assumes that a single (common) true effect size underlies all the [primary study](#) results. See [Common-effect \(“fixed-effect”\) model](#) in [\[META\] Intro](#).

cumulative meta-analysis. Cumulative meta-analysis performs multiple meta-analyses by accumulating studies one at a time. The studies are first ordered with respect to the variable of interest, the ordering variable. Meta-analysis summaries are then computed for the first study, for the first two studies, for the first three studies, and so on. The last meta-analysis will correspond to the standard meta-analysis using all studies. See [\[META\] meta summarize](#).

cumulative overall effect sizes. In the context of cumulative meta-analysis, cumulative (overall) effect sizes refer to the overall effect sizes computed by accumulating one study at a time. That is, the first overall effect size is simply the individual effect size of the first study. The second overall effect size is the overall effect size computed based on the first two studies. The third overall effect size is the overall effect size computed based on the first three studies. And so on. The last effect size in a cumulative meta-analysis corresponds to the overall effect size computed using all studies in a standard meta-analysis.

DerSimonian–Laird's method. A noniterative, random-effects estimator of the between-study variance parameter that does not make any assumptions about the distribution of random effects. This method was introduced in [DerSimonian and Laird \(1986\)](#). Historically, random-effects meta-analysis has been based solely on this method. See [Noniterative methods](#) of [Methods and formulas](#) in [\[META\] meta summarize](#).

effect size. A numerical summary of the group differences or of association between factors. For example, effect sizes for two-group comparisons include standardized and unstandardized mean differences, odds ratio, risk ratio, hazard ratio, and correlation coefficient. See [\[META\] meta esize](#).

Egger test. A regression-based test for funnel-plot asymmetry of (Egger et al. 1997). This is the test of a slope coefficient in a weighted regression of the effect sizes on their standard errors. See [META] [meta bias](#).

fixed-effects meta-analysis model. A meta-analysis model that assumes effect sizes are different across the studies and estimates a weighted average of their true values. This model is not valid for making inferences about studies beyond those included in the meta-analysis. See *Fixed-effects model* in [META] [Intro](#).

fixed-effects meta-regression. [Meta-regression](#) that assumes a fixed-effects meta-analysis model. This regression model does not account for residual heterogeneity. See *Introduction* in [META] [meta regress](#).

forest plot. A forest plot is a graphical representation of the results of a meta-analysis. In addition to meta-analytic summary such as overall effect size and its confidence interval and heterogeneity statistics and tests, it includes study-specific effect sizes and confidence intervals. See [META] [meta forestplot](#).

funnel plot. The funnel plot is a scatterplot of the study-specific effect sizes against measures of study precision. This plot is commonly used to explore small-study effects or publication bias. In the absence of small-study effects, the shape of the scatterplot should resemble a symmetric inverted funnel. See [META] [meta funnelplot](#).

Glass's Δ . An effect-size measure introduced by Smith and Glass (1977) for continuous outcomes. It is a standardized mean difference where the difference between the two group means is divided by the sample standard deviation of the control group. Another variation of this statistic uses the sample standard deviation of the treatment group for the standardization. See *Standardized mean difference* of *Methods and formulas* in [META] [meta esize](#).

grey literature. In the context of meta-analysis, grey literature refers to the literature that is difficult to obtain; it is thus rarely included in a meta-analysis.

H^2 statistic. A statistic for assessing heterogeneity. A value of $H^2 = 1$ indicates perfect homogeneity among the studies. See *Heterogeneity measures* of *Methods and formulas* in [META] [meta summarize](#).

Hedges's g . An effect-size measure introduced by Hedges (1981) for continuous outcomes. It is a Cohen's d statistic adjusted for bias. See *Standardized mean difference* of *Methods and formulas* in [META] [meta esize](#).

heterogeneity. In a meta-analysis, statistical heterogeneity, or simply heterogeneity, refers to the variability between the study-specific effect sizes that cannot be explained by a random variation. See *Heterogeneity* in [META] [Intro](#).

heterogeneity parameter. In a random-effects meta-analysis, the variance of the random effects, τ^2 , is used to account for the between-study heterogeneity. It is often referred to as the "heterogeneity parameter".

homogeneity. The opposite of [heterogeneity](#).

homogeneity test. A test based on Cochran's Q statistic for assessing whether effect sizes from studies in a meta-analysis are homogeneous. See *Homogeneity test* of *Methods and formulas* in [META] [meta summarize](#).

I^2 statistic. A statistic for assessing heterogeneity. It estimates the proportion of variation between the effect sizes due to heterogeneity relative to the pure sampling variation. $I^2 > 50$ indicates substantial heterogeneity. See *Heterogeneity measures* of *Methods and formulas* in [META] [meta summarize](#).

intervention effects. See *effect size*.

inverse-variance method. A method of estimating the overall effect size as a weighted average of the study-specific effect sizes by using the weights that are inversely related to the variance (Whitehead and Whitehead 1991). This method is applicable to all meta-analysis models and all types of effect sizes.

L'Abbé plot. A scatterplot of the summary outcome measure such as log odds in the control group on the x axis and of that in the treatment group on the y axis. It is used with binary outcomes to inspect the range of group-level summary outcome measures among the studies to identify excessive heterogeneity. See [META] [meta labbeplot](#).

large-strata limiting model. A model assumption for binary data in which the number of studies remains fixed but similar cell sizes in the 2×2 tables increase. See Robins, Breslow, and Greenland (1986).

Mantel–Haenszel method. In the context of meta-analysis, the Mantel–Haenszel method combines odds ratios, risk ratios, and risk differences. This method performs well in the presence of sparse data. For nonparse data, its results are similar to those of the inverse-variance method. It was introduced by Mantel and Haenszel (1959) for odds ratios and extended to risk ratios and risk differences by Greenland and Robins (1985). See *Mantel–Haenszel method for binary outcomes* of *Methods and formulas* in [META] [meta summarize](#).

meta data. meta data are the data that were meta set (or declared) by either [meta set](#) or [meta esize](#). meta data store key variables and characteristics about your meta-analysis specifications, which will be used by all meta commands during your meta-analysis session. Thus, declaration of your data as meta data is the first step of your meta-analysis in Stata. This step helps minimize mistakes and saves you time—you need to specify the necessary information only once. Also see [META] [meta data](#).

meta settings. Meta settings refers to the meta-analysis information specified during the declaration of the meta data via [meta set](#) or [meta esize](#). This includes the declared effect size, meta-analysis model, estimation method, confidence level, and more. See *Declaring meta-analysis information* in [META] [meta data](#) for details.

meta-analysis. The statistical analysis that combines quantitative results from multiple individual studies into a single result. It is often performed as part of a systematic review. See *Brief overview of meta-analysis* in [META] [Intro](#).

meta-regression. A weighted regression of study effect sizes on study-level covariates or moderators. You can think of it as an extension of standard meta-analysis to incorporate the moderators to account for between-study heterogeneity. See [META] [meta regress](#).

methodological heterogeneity. Variability in study design and conduct (Deeks, Higgins, and Altman 2017). See *Heterogeneity* in [META] [Intro](#).

moderator. A moderator is a study-level covariate that may help explain between-study heterogeneity. If the moderator is categorical, its effect may be investigated by a subgroup analysis (see [META] [meta summarize](#)); if the moderator is continuous, its effect may be investigated by a meta-regression. See [META] [meta regress](#).

multiple subgroup analyses. Subgroup analysis performed separately for each of multiple categorical variables. See [META] [meta summarize](#).

multiplicative dispersion parameter. In a fixed-effects meta-regression, the multiplicative dispersion parameter is a multiplicative factor applied to the variance of each effect size to account for [residual heterogeneity](#). See *Introduction* of [META] [meta regress](#).

multiplicative meta-regression. A fixed-effects meta-regression that accounts for [residual heterogeneity](#) through a dispersion parameter ϕ applied (multiplicatively) to each effect-size variance. See [Introduction](#) of [\[META\] meta regress](#).

narrative review. In a narrative review, the conclusion about the findings from multiple studies is given by a person, an expert in a particular field, based on his or her research of the studies. This approach is typically subjective and does not allow to account for certain aspects of the studies such as study heterogeneity and publication bias.

odds ratio. A ratio of the odds of a success in one group (treatment group) to those of another group (control group). It is often used as an effect size for comparing binary outcomes of two groups. See [\[META\] meta esize](#).

overall effect size. The main target of interest in meta-analysis. Its interpretation depends on the assumed meta-analysis model. In a common-effect model, it is the common effect size of the studies. In a fixed-effects model, it is a weighted average of the true study-specific effect sizes. In a random-effects model, it is the mean of the distribution of the effect sizes. The overall effect size is usually denoted by θ in the output. Also see [Meta-analysis models](#) in [\[META\] Intro](#).

Peto's method. A method for combining odds ratios that is often used with sparse 2×2 tables. This method does not require a [zero-cell adjustment](#). See [Peto's method for odds ratios](#) of [Methods and formulas](#) in [\[META\] meta summarize](#).

pooled effect size. See [overall effect size](#).

prediction interval. In a random-effects meta-analysis, a $100(1 - \alpha)\%$ prediction interval indicates that the true effect sizes in $100(1 - \alpha)\%$ of new studies will lie within the interval. See [Prediction intervals](#) of [Methods and formulas](#) in [\[META\] meta summarize](#).

primary study. The original study in which data are collected. An observation in a meta-analysis represents a primary study.

pseudo confidence interval. Pseudo confidence intervals refer to the confidence intervals as constructed by the standard funnel plot. See [\[META\] meta funnelplot](#).

publication bias. Publication bias is known in the meta-analysis literature as an association between the likelihood of a publication and the statistical significance of a study result. See [Publication bias](#) in [\[META\] Intro](#).

Q statistic. The test statistic of the [homogeneity test](#). See [Homogeneity test](#) of [Methods and formulas](#) in [\[META\] meta summarize](#).

random-effects meta-analysis model. A meta-analysis model that assumes that the study effects are random; that is, the studies used in the meta-analysis represent a random sample from a larger population of similar studies. See [Random-effects model](#) in [\[META\] Intro](#).

random-effects meta-regression. [Meta-regression](#) that assumes a random-effects meta-analysis model. This regression model accounts for residual heterogeneity via an additive error term. See [Introduction](#) in [\[META\] meta regress](#).

randomized controlled trial. A randomized controlled trial is an experiment in which participants are randomly assigned to two or more different treatment groups. Randomized controlled trials are commonly used in clinical research to determine the effectiveness of new treatments. By design, they avoid bias in the treatment estimates.

rate ratio. See [risk ratio](#).

relative risk. See [risk ratio](#).

reporting bias. Systematic difference between the studies selected in a meta-analysis and all the studies relevant to the research question of interest. Also see [publication bias](#).

residual heterogeneity. In the meta-regression context, this is the remaining variability between the studies not accounted for by the moderators. It is usually captured by the [heterogeneity parameter](#) in a random-effects meta-regression or by a [multiplicative dispersion parameter](#) in a fixed-effects meta-regression.

risk ratio. A ratio of the success probability in one group (treatment) to that of another group (control). It is often used as an effect size for comparing binary outcomes of two groups. See [\[META\] meta esize](#).

sensitivity analysis. In the context of meta-analysis, sensitivity analyses are used to assess how robust the meta-analysis results are to assumptions made about the data and meta-analysis models. See [\[META\] meta summarize](#).

significance contours. In the context of a funnel plot ([\[META\] meta funnelplot](#)), significance contours (or contour lines of statistical significance) are the contour lines corresponding to the tests of significance of individual effect sizes for a given significance level $\alpha = c/100$. In other words, if a study falls in the shaded area of a c -level contour, it is considered not statistically significant at the α level based on a test of significance of the study effect size.

single subgroup analysis. Subgroup analysis performed for one categorical variable. See [\[META\] meta summarize](#).

small-study effects. Small-study effects arise when the results of smaller studies differ systematically from the results of larger studies. See [Introduction of \[META\] meta funnelplot](#).

sparse data. For binary data, a 2×2 table is considered sparse if any of the cell counts are small.

sparse data limiting model. A model assumption for binary data in which the number of 2×2 tables (studies) increases but the cell sizes remain fixed. See [Robins, Breslow, and Greenland \(1986\)](#).

statistical heterogeneity. See [heterogeneity](#).

study precision. Study precision is a function of a study sample size or study variability. Typically, study precision is measured by the inverse of the effect-sizes standard errors, $1/\hat{\sigma}_j$, but other measures are also used. For instance, in a funnel plot, multiple precision metrics such as variances and sample sizes are considered. More precise studies (with larger sample sizes and smaller variances) are assigned larger weights in a meta-analysis.

subgroup analysis. A subgroup analysis divides the studies into groups and then estimates the overall effect size for each of the groups. The goal of subgroup analysis is to compare the overall effect sizes and explore heterogeneity between the subgroups. See [\[META\] meta summarize](#) and [\[META\] meta forestplot](#).

subgroup heterogeneity. In the context of meta-analysis, subgroup heterogeneity is between-study heterogeneity induced by the differences between effect sizes of groups defined by one or more categorical variables. See [\[META\] meta](#) and [\[META\] meta summarize](#).

summary data. In the context of [meta-analysis](#), we use the term summary data to mean summary statistics that are used to compute the effect sizes and their standard errors for each study in the meta-analysis. For example, for a two-group comparison of continuous outcomes, the summary data contain the number of observations, means, and standard deviations in each group for each study. For a two-group comparison of binary outcomes, the summary data contain the 2×2 tables for each study. See [\[META\] meta esize](#).

summary effect. See [overall effect size](#).

systematic review. A procedure that uses systematic and well-defined methods to find, select, and evaluate relevant research studies to answer a specific research question. It typically involves collecting and analyzing summary data of the selected studies. Meta-analysis is the statistical analysis used as part of a systematic review.

trim-and-fill method. A method of testing and adjusting for publication bias in meta-analysis; see [META] **meta trimfill**.

zero-cell adjustment. Adjustment made to cells of 2×2 tables containing zero cells. In the meta-analysis of binary data, zero-cell counts pose difficulties when computing odds ratios and risk ratios. Therefore, it is common to make zero-cell adjustments, such as adding a small number to all cells containing zeros. See *Zero-cells adjustments* in *Methods and formulas* of [META] **meta esize**.

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Subject and author index

See the [combined subject index](#) and the [combined author index](#) in the *Glossary and Index*.